

UNIVERSIDADE FEDERAL DE PERNAMBUCO  
CENTRO DE TECNOLOGIA E GEOCIÊNCIAS  
DEPARTAMENTO DE OCEANOGRAFIA

Biologia populacional de *Pomacanthus paru* (Teleostei: Pomacanthidae) e análise da sustentabilidade de captura do *bycatch* de peixes ornamentais.



Caroline Vieira Feitosa



**UNIVERSIDADE FEDERAL DE PERNAMBUCO**

**CENTRO DE TECNOLOGIA E GEOCIÊNCIAS**

**DEPARTAMENTO DE OCEANOGRAFIA**

**Biologia populacional de *Pomacanthus paru* (Teleostei: Pomacanthidae) e análise da sustentabilidade de captura do *bycatch* de peixes ornamentais.**

Caroline Vieira Feitosa

Tese submetida ao curso de Doutorado do Programa de Pós Graduação em Oceanografia da Universidade Federal de Pernambuco, como requisito para obtenção do título de Doutor em Oceanografia.

Orientadora: Dr<sup>a</sup> Maria Elisabeth de Araújo  
Co-Orientadora: Dr<sup>a</sup> Beatrice Padovani Ferreira

Fevereiro/2009

Feitosa, Caroline Vieira

Biologia populacional de *Pomacanthus paru* (Teleostei: Pomacanthidae) e análise da sustentabilidade de captura do bycatch de peixes ornamentais. / Caroline Vieira Feitosa . – Recife : O Autor, 2009.

108 folhas : il., fig., tab.

Tese (doutorado) – Universidade Federal de Pernambuco. CTG. Oceanografia, 2009.

Inclui bibliografia

1. Biologia populacional – Reprodução, idade e crescimento 2. *Pomacanthus paru*. 3. Bycatch. 4. Sustentabilidade de captura. I. Título.

551.46

CDU (2.ed.)

UFPE

551.46

CDD (22.ed.)

3C - 2009 - 048



UNIVERSIDADE FEDERAL DE PERNAMBUCO  
CENTRO DE TECNOLOGIA E GEOCIÊNCIAS  
DEPARTAMENTO DE OCEANOGRAFIA

**Biologia populacional de *Pomacanthus paru* (Teleostei: Pomacanthidae) e análise da sustentabilidade de captura do *bycatch* de peixes ornamentais.**

Caroline Vieira Feitosa

Tese submetida em 11 de fevereiro de 2009 ao curso de Doutorado do Programa de Pós Graduação em Oceanografia da Universidade Federal de Pernambuco, como requisito para obtenção do título de Doutor em Oceanografia e aprovada pela seguinte banca examinadora:

---

Dr. Francisco Marcante Santana Silva

---

Dr<sup>a</sup>. Maria Elisabeth de Araújo

---

Dr. Paulo Jorge Parreira dos Santos

---

Dr. Ronaldo Bastos Francini Filho

---

Dr<sup>a</sup> Simone Ferreira Teixeira

Dedico esta tese a minha querida vó Luda  
(*in memoriam*), por todo o amor e pela  
alegria ao vivenciar minhas conquistas  
acadêmicas, mesmo torcendo a  
distância...

## **AGRADECIMENTOS**

Ao Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) pela bolsa de estudo concedida. A Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco pelo financiamento do projeto.

A minha orientadora Maria Elisabeth de Araújo pela “adoção”, por todos os ensinamentos durante minha vida acadêmica, além da grande amizade. Uma orientadora que sempre torceu e vibrou a cada conquista da sua “pupila”. Muito obrigada!!

A minha co-orientadora Beatrice Padovani Ferreira por todas as valiosas críticas e oportunidades. Sempre contribuiu com sugestões brilhantes! Agradeço também pelos divertidos momentos de trabalho e lazer em Tamandaré.

Aos meus pais Hedilberto e Solange por todo o apoio, paciência e confiança depositada em mim.

A Myrna, secretária do Curso de Pós Graduação em Oceanografia por todo o suporte ao longo do curso.

Aos pescadores da colônia de Itamaracá, em especial Gulu, Chico e Carlinhos, pela ajuda na amostragem dos peixes.

A todos que me ajudaram no processamento das amostras: Bebeto, Lula, Beth, Fernando, Adriane, Henrique, Rodrigo. Ao Seu Mano e ao Beto que sempre me ajudaram na limpeza do laboratório após o processamento das amostras.

Ao Prof. Diógenes e Alex Benício (UFPE – Departamento de Farmácia) pelos ensinamentos histológicos.

À Prof<sup>a</sup> Raquel Coimbra (UFRPE – Departamento de Pesca) pela estrutura cedida no Laboratório de Genética Aplicada (LAGA) para extração de DNA. Especialmente, agradeço a Patrícia Lima que me ensinou e me acompanhou durante o processo de extração do DNA. Ao Dr. Miguel Martins Moreira (INCA – Pesquisa básica) pelo sequenciamento das amostras e sugestões no capítulo de genética.

Ao Prof. Fernando Feitosa por ter me ajudado na “caça” ao microscópio com luz UV.

Ao Prof. Marcelo dos Santos Guerra Filho (UFPE) pela estrutura cedida no Laboratório de Citogenética Vegetal. Agradeço também a Ana Emília Barros e Silva e a Gabriela Corina Cabral pelas instruções e auxílio no manuseio do microscópio com luz UV.

Ao Bili e a Ana pelas indispensáveis informações e dicas na pesquisa com otólitos. Ao Nino, Alexandre e Simone pela ajuda com as leituras dos otólitos e conversas agradáveis no laboratório.

A Adriane pelo fundamental auxílio durante o experimento de validação.

Ao Henrique Maranhão pela ajuda com os mapas e referências, além da fiel companhia durante as longas jornadas de laboratório.

A Nice pelo apoio na alimentação dos peixes durante o período de validação. Não falhava nem aos domingos!

A minha grande amiga Ana Lídea pelo companheirismo, amizade, longas conversas e risadas ao longo desses anos. E que venham muitos carnavais, coleguinha!!!

A minha amiga cearense/pernambucana Danise pelo help nas análises estatísticas e pela companhia maravilhosa que com aquele sotaque e expressões faz com que eu me sinta em casa.

Ao Lula por todo o apoio e compreensão durante boa parte da pesquisa. Coleta, processamento de amostra, montagem dos aquários... Sempre vinha passear e acabava entrando no trabalho junto comigo. Muito obrigada!

As coleguinhas Alê, Cris, Dan, Irlanda, Lálá e Laís por me ajudarem a repor ou a perder o restinho das energias nos finais de semana.

Aos amigos dos laboratórios Henrique, Nicole, Cabelo, Adri, Jana, Jorge, Andréa, Beth Cabral, Andreza, Nino, Dudu, Simone e Alexandre, pelas divertidas horas de trabalho, boas conversas, incentivo e sugestões.

As inúmeras pessoas que, ao longo desses quatro anos, de alguma forma contribuíram para a conclusão desta pesquisa. Muito obrigada!



## Sumário

	Página
Resumo	II
Abstract	III
Lista de Figuras	IV
Lista de Tabelas	VI
Introdução	1
<b>Chapter 1</b> - Reproduction of the French Angelfish <i>Pomacanthus paru</i> (Teleostei: Pomacanthidae) from Northeastern Brazil.	10
Introduction	12
Material and methods	14
Results	18
Discussion	24
<b>Chapter 2</b> - Age and growth of the French angelfish <i>Pomacanthus paru</i> (Teleostei: Pomacanthidae) from Northeastern Brazil.	37
Introduction	38
Material and methods	40
Results	43
Discussion	48
<b>Chapter 3</b> - Diverse Mitochondrial DNA variability in sympatric populations of <i>Pomacanthus</i> (Actinopterygii: Pomacanthidae).	61
Introduction	63
Material and methods	65
Results	68
Discussion	70
<b>Chapter 4</b> - A rapid new method for assessing sustainability of ornamental fish <i>bycatch</i> from coral reefs.	82
Introduction	83
Material and methods	84
Results	90
Discussion	94
Considerações finais	105
Apêndice	108

## Resumo

*Pomacanthus paru* tem grande importância para o aquarismo brasileiro, além de ser a segunda espécie marinha mais exportada desde 2000. Os objetivos desta pesquisa foram os seguintes: (a) descrever a biologia reprodutiva e determinar a idade e o crescimento de *P. paru*, (b) analisar a diversidade genética de *P. paru* e *P. arcuatus* a partir do DNA mitocondrial, (c) registrar as espécies de peixes ornamentais capturadas como *bycatch* através da pesca com armadilhas e propor uma metodologia para determinar a sustentabilidade de captura das principais espécies ornamentais marinhas no Brasil. Amostragens semanais foram realizadas durante o período de março 2006 a fevereiro 2007. Os exemplares menores de 15 cm foram comprados de coletores de peixes ornamentais. Os peixes foram medidos, pesados, e as gônadas e os otólitos foram removidos. Amostras de músculo e do fígado foram armazenadas para o estudo genético. A região 16S rDNA foi amplificada e sequências do ADN mitocondrial foram utilizadas para a análise dos haplótipos. Para a proposição da metodologia de sustentabilidade de captura, cada espécie foi avaliada segundo dois grupos de critérios: (1) vulnerabilidade relativa à captura pelas armadilhas; (2) resiliência da população relacionada à mortalidade devido à pressão pesqueira. No total, 236 indivíduos foram analisados, sendo 139 fêmeas, 86 machos e 11 que não tiveram o sexo definido. *P. paru* é uma espécie gonocorística, que se reproduz ao longo do ano, com as fêmeas apresentando dois picos de desova (Fevereiro-Março/Setembro-Outubro). As fêmeas foram mais abundantes em todos os meses, exceto em outubro. O  $L_{50}$  é atingido aos 30 cm para as fêmeas e 35 cm para os machos. A idade dos indivíduos amostrados de *P. paru* variou de 1 a 27 anos. *P. paru* atinge o comprimento máximo lentamente e tem um longo ciclo de vida. Para *P. paru* seis haplótipos foram encontrados, enquanto em *P. arcuatus* apenas um, incluindo os espécimes do Caribe. Foram registradas como *bycatch*, 19 espécies pertencentes a 10 famílias. As espécies cujas capturas foram menos sustentáveis foram *Holacanthus tricolor*, *H. ciliaris*, *Chaetodon striatus* e *P. paru*. O manejo de *P. paru* deve incluir medidas baseadas no tamanho de primeira maturação sexual, bem como nos parâmetros de crescimento aqui estabelecidos. Os resultados relacionados à genética representam um primeiro passo para se compreender as estruturas populacionais de espécies tão próximas filogenética e ecologicamente como *P. paru* e *P. arcuatus*. A nova metodologia aqui proposta deverá ser uma ferramenta útil e alternativa para o manejo e avaliação da sustentabilidade de captura das espécies oriundas de pescarias multi-específicas, com poucos dados disponíveis e que ocorrem em áreas de alta riqueza, como os recifes.

**Palavras-chave:** *Pomacanthus paru*, reprodução, idade, crescimento, *bycatch*

## Abstract

*Pomacanthus paru* presents great importance in the Brazilian aquarium trade, being the second most exported species since 2000. The aims of this research were (a) to describe the reproductive biology and to determine the age and growth, (b) to analyze the genetic diversity of *P. paru* and *P. arcuatus* from mt-DNA, (c) to record ornamental fish *bycatch* and to propose a methodology to assess the catch sustainability of ornamental fish *bycatch* from the major Brazilian families in the aquarium trade. Weekly samplings were performed from March 2006 to February 2007. Individuals smaller than 15 cm were bought from collectors of ornamental fish. The fish were measured, weighed and the gonads and otoliths were removed. Samples of tissue from muscle and liver were stored for the genetic study. A region of the 16S rDNA was amplified and sequences of the mtDNA were used for haplotypes analysis. To assess the catch sustainability of ornamental fish *bycatch*, each species was assessed based on two groups of criteria: (1) relative vulnerability to capture; and (2) population resilience regarding mortality due to fishing efforts. Of the 236 individuals sexed, 139 were female, 86 were male and 11 were indefinite. *P. paru* is a gonochoric species, which is reproductive active all year-round, with females presenting two spawning peaks (February-March/September-October). Females were more abundant at all months, except October. The  $L_{50}$  is reached at 30 cm for females and 35 cm for males. The ages of the fish in the sample varied from 1 to 27 years. *P. paru* attains its maximal size slowly and has a long lifespan. For *P. paru* six haplotypes were recorded, while only one was registered for *P. arcuatus*, including Caribbean species. Nineteen species from 10 families were recorded. The species least likely to have sustainable catch rates were *Holacanthus tricolor*, *H. ciliaris*, *Chaetodon striatus* and *P. paru*. The management of this latter species should include measures based on its size at maturity as well as in growth parameters established in this research. The considerations regarding genetic represent a first step toward understanding the population structures of species as phylogenetically and ecologically close as *Pomacanthus paru* and *P. arcuatus*. The proposed methodology is a useful alternative for management efforts and the assessment of catch sustainability in newly-developed multi-species fisheries with limited available data, which occur in areas of high species richness such as reefs.

**Key-words:** *Pomacanthus paru*, reproduction, age, growth, *bycatch*

## Lista de Figuras

	Página
<b>Chapter 1</b> - Reproduction of the French Angelfish <i>Pomacanthus paru</i> (Teleostei: Pomacanthidae) from Northeastern Brazil.	
Figure 1 - Location of the sampled site.	14
Figure 2 - Size distribution between sexes of sampled individuals with bars indicating standard error.	19
Figure 3 - Sampled individuals distribution of female and male of <i>Pomacanthus paru</i> among bimesters.	19
Figure 4 - Mean total length distribution of <i>Pomacanthus paru</i> .	20
Figure 5 - Bimonthly changes of the mean gonadosomatic index (GSI) in female and male <i>Pomacanthus paru</i> with bars and values indicating the standard error and the number of Individuals analyzed for each stage, respectively.	21
Figure 6 - Bimonthly percentage frequencies of occurrence of gonadal developmental stages in female and male <i>Pomacanthus paru</i> . The values inform the number individual analyzed for each stage.	22
Figure 7: Percentage of individuals in spawning and immature stage versus length class and mean gonad weight for female and male (A) and graphic showing the $L_{50}$ for female and male of <i>Pomacanthus paru</i> .	23
<b>Chapter 2</b> - Age and growth of the French angelfish <i>Pomacanthus paru</i> (Teleostei: Pomacanthidae) from Northeastern Brazil.	
Figure 1 - Location of the sampled area.	41
Figure 2 - Whole (A) and sectioned (B) <i>sagittae</i> otoliths of a 8 and a 11 years old <i>Pomacanthus paru</i> (a = anterior, p = posterior, d = dorsal, v = ventral, di = distal, pr = proximal).	44
Figure 3 - Sectioned otolith treated with tetracycline observed under UV light (A) and under an external light source and UV light (B). TM – tetracycline mark, OM – opaque mark.	45
Figure 4 - Relation between <i>sagittae</i> otolith weight and standard length for <i>Pomacanthus paru</i> . SL = standard length (cm), OW = otolith weight.	46
Figure 5 - Relation between otolith weight and age for <i>Pomacanthus paru</i> . A = age years, OW = otolith weight.	46
Figure 6 - Relation of weight to length for <i>Pomacanthus paru</i> . SL = standard length (cm), W = total weight (g).	47
Figure 7 - Von Bertalanffy growth curve fitted to length at age data of female and male of <i>Pomacanthus paru</i> .	48
<b>Chapter 3</b> - Diverse Mitochondrial DNA variability in sympatric populations of <i>Pomacanthus</i> (Actinopterygii: Pomacanthidae).	
Figure 1 - Location of sampled area. CE - Ceará State, PE - Pernambuco State, AL - Alagoas State	66

Figure 2 - Neighbor-Joining (A) and Parsimony (B) topologies based on 16S rDNA mitochondrial sequences for the haplotypes of <i>Pomacanthus paru</i> , <i>Pomacanthus arcuatus</i> and other <i>Pomacanthus</i> species. <i>C. flavissimus</i> - <i>Centropyge flavissimus</i> and <i>C. dubolayi</i> - <i>Chaetodontoplus duboulayi</i> were used as outgroups. Localities where the haplotypes were found are indicated between parenthesis: AL – Alagoas state, Brazil; CE – Ceará state, Brazil; PE – Pernambuco state, Brazil; and Caribbean. DC indicates specimens of <i>P. paru</i> with grayish coloration.	69
Figure 3 – Parsimony network of the mitochondrial haplotypes from <i>Pomacanthus paru</i> and <i>Pomacanthus arcuatus</i> . The circles represent the haplotypes and the diameter is proportional to the number of specimens carrying each haplotype, except for <i>P. arcuatus</i> (Ha 1). Nucleotide substitutions between haplotypes were indicated.	70
<b>Chapter 4</b> - A rapid new method for assessing sustainability of ornamental fish <i>bycatch</i> from coral reefs.	
Figure 1 - Location of sampled area.	85
Figure 2 - Length distributions for the four analyzed species. P values refer to normality.	91
Figure 3 - Correspondence analysis showing the relationship between species and criteria used to evaluate the recovery capacity. Caption: <i>Hol. cil.</i> – <i>Holacanthus ciliaris</i> , <i>Hol. tri.</i> - <i>Holacanthus tricolor</i> <i>Cha. str.</i> – <i>Chaetodon striatus</i> , <i>Pom. par.</i> – <i>Pomacanthus paru</i> , Max. Size – Maximum size.	94

## Lista de Tabelas

	Página
<b>Chapter 1</b> - Reproduction of the French Angelfish <i>Pomacanthus paru</i> (Teleostei: Pomacanthidae) from Northeastern Brazil.	
Table 1 - Macroscopic and histological stages used to describe ovaries and testes of <i>Pomacanthus paru</i> based in Murphy and Taylor (1990), West (1990) and Yamaguchi <i>et al.</i> (2006).	16
Table 2 - Minimum, maximum and mean values of batch (BF), total (TF) and relative (RF) fecundity of <i>Pomacanthus paru</i> .	24
<b>Chapter 3</b> - Diverse Mitochondrial DNA variability in sympatric populations of <i>Pomacanthus</i> (Actinopterygii: Pomacanthidae).	
Table 1 - Variable sites found in haplotypes of <i>P. paru</i> (Hp 1-6) and <i>P. acutatus</i> (Ha 1) and number of individuals sharing a same haplotype per locality. Sequences from the Belize specimen belonging to Hp 4 haplotype and the specimen from <i>P. arcuatus</i> from the Caribbean were obtained from GenBank.	68
<b>Chapter 4</b> - A rapid new method for assessing sustainability of ornamental fish <i>bycatch</i> from coral reefs.	
Table 1 - The criteria used to assess the susceptibility of <i>bycatch</i> species to capture and mortality due to traps and their recovery. For each criterion the weighting score and definition of the three ranks is given.	89
Table 2 - Specie registered as <i>bycatch</i> , with the respective common names.	92
Table 3 - The ranking of the species on the criteria on the resilience axis.	93
Table 4 - The length at first capture, length at maturity, probability of individuals caught are below of the length at maturity, probability of breeding, mean and maximum length (Max. length) of the species and the mortality index (Z) of each.	93

## Introdução

Os peixes da família Pomacanthidae, conhecidos como peixes-anjo, possuem o corpo alto e comprimido lateralmente, algumas vezes quase circular (Cervigón, 1993). São bastante parecidos com os peixes borboleta da família Chaetodontidae (Smith & Heemstra, 1991) e Ehippidae (Menezes & Figueiredo, 1985). Apresentam boca pequena, terminal e protátil, extremidade inferior do pré-opérculo com um forte e largo espinho e escamas ctenóides pequenas (Menezes & Figueiredo, 1985; Cervigón, 1993). São de tamanho médio e possuem padrões de colorido variados e contrastantes, diferindo notadamente entre os jovens e adultos (Nelson, 1994). É uma das famílias mais procuradas pelos aquaristas, sendo o colorido a principal razão desta popularidade (Allen et al., 1998).

Os pomacantídeos distribuem-se por todas as latitudes tropicais e subtropicais. Estão geralmente associados às águas costeiras com desenvolvimento de recifes ou pedras (Cervigón, 1993; Allen, 1979) em profundidades inferiores a 20m e raramente abaixo dos 50m (Nelson, 2006). A família Pomacanthidae possui 82 espécies e oito gêneros (Nelson, 2006) com mais da metade das espécies sendo pertencentes aos gêneros *Centropyge* e *Pomacanthus* (Nelson, 1994). Destas, 89% do total ocorrem nos oceanos Índico e Pacífico. No Atlântico, a maioria das espécies ocorre na porção oeste deste oceano, comumente referidas para o mar do Caribe. Apenas uma espécie é exclusiva do oeste da África e outra do Atlântico Central (Ascensão). Para Austrália são registradas 24 espécies, sendo considerado o país com a maior diversidade de pomacantídeos (Allen et al., 1998). O gênero *Pomacanthus* está representado por 13 espécies, além de 4 híbridos (Allen et al., 1998). Estas espécies podem ser encontradas em águas rasas em regiões de recifes e pedras, em grupos ou isoladamente (Menezes & Figueiredo, 1985).

Estudos sobre a ecologia dos pomacantídeos são numerosos (e.g. Randall & Hartman, 1968; Hourigan et al., 1989; Roberts et al., 1992; Sakai & Kohda, 1997; Kuwamura, 1998;

Kulbick & Sarramégna, 1999; Sazima et al., 1999; Aburto-Oropeza et al., 2000; Hamagushi et al., 2002; Bellwood et al., 2004). Entretanto, informações sobre seus aspectos populacionais como idade, crescimento e longevidade são relativamente escassas. A única espécie da família avaliada quanto à idade e o crescimento foi *Pomacanthus imperator*, onde as escamas foram utilizadas para determinação destes parâmetros (Chung & Woo, 1999). Os pomacantídeos foram geneticamente estudados principalmente no Atlântico Sul. Foram realizadas análises com marcadores citogenéticos (Galetti et al., 2006), enzimáticos (Chung & Woo, 1998), e mitocondriais (Bellwood et al., 2004; Bowen et al., 2006), principalmente com o objetivo de responder questões taxonômicas e evolutivas. Uma vez que os pomacantídeos encontram-se sob forte pressão pesqueira, informações genéticas continuam sendo necessárias a fim de obter uma medida acurada da biodiversidade (Affonso & Galetti, 2007).

Dentre as espécies de pomacantídeos, aquelas do gênero *Centropyge* são as mais estudadas principalmente no que se refere ao desenvolvimento gonadal e comportamento reprodutivo (Bauer & Bauer, 1981; Moyer et al., 1983; Aldenhoven, 1986; Sakai, 1986; Sakai & Kohda, 1997). Alguns estudos foram realizados enfocando aspectos reprodutivos para os gêneros *Holacanthus* e *Pomacanthus*, tais como tamanho de primeira maturação sexual, fecundidade (n = 2) (Aiken, 1983) e época de desova (n = 21) (Munro et al., 1973). Porém, o tamanho amostral de *P. paru* foi considerado pequeno pelos autores para realizar comparações com outras pesquisas.

Os principais componentes da dieta destes indivíduos são as esponjas e algas (Randall, 1967; Randall & Hartman, 1968). Quando jovens, complementam sua dieta retirando ectoparasitas de peixes de maior porte (Allen et al., 1998; Sazima et al., 1999). São espécies que normalmente formam pares, entretanto pequenos haréns foram registrados (Allen et al., 1998).



A espécie *P. paru* (Bloch, 1787) distribui-se desde a Flórida até o sudeste do Brasil. É relativamente comum no litoral brasileiro (Menezes & Figueiredo, 1985), sendo mais abundante na região Sudeste quando comparada a Nordeste (Ferreira et al., 2004). Ocorre em profundidades entre 5 a 100 m (Allen, 1979). Atinge cerca de 40 cm de comprimento e quando adulto, possui o corpo enegrecido, tendo a cabeça uma coloração mais clara e as margens das escamas amareladas. Nos jovens o corpo é totalmente negro com cinco faixas amarelas verticais (Allen et al., 1998). O jovem de *P. paru* é muito parecido com a espécie *P. arcuatus*, diferindo no número de raios e principalmente na nadadeira caudal, que é truncada posteriormente em *P. arcuatus* e arredondada em *P. paru* (Menezes & Figueiredo, 1985).

O conhecimento do ciclo de vida de espécies como *P. paru*, que são extraídas diretamente do ambiente natural para o aquarismo, é fundamental para a sua conservação. Embora *P. paru* não se encontre na lista de espécies ameaçadas de extinção da IUCN (IUCN, 2008), ela é muito explorada no Brasil pela indústria aquariofilista (Monteiro-Neto et al., 2003). Devido à pressão pesqueira sobre os jovens, a abundância de *P. paru* em alguns pontos próximo a Guarapari (ES) está inversamente relacionada com a distância da costa, enquanto o número de adultos não difere (Gasparini et al., 2005). A abundância desta espécie é significativamente maior em áreas protegidas (Gasparini et al., 2005). Nos últimos anos, *P. paru* vem sendo listada como a segunda espécie mais exportada pelo Brasil (Monteiro-Neto et al., 2003; IBAMA, 2007). Entre o período de 1995 a 2000, somente o Estado do Ceará exportou 22.969 espécimes de *P. paru* (Monteiro-Neto et al., 2003) e, em 2007, 13.422 indivíduos foram exportados pelo Brasil (IBAMA, 2007). O ordenamento destas espécies vem sendo realizado através de uma instrução normativa do IBAMA (IN56/04), onde foram estabelecidas cotas específicas de exportação. Atualmente a cota de captura deste indivíduo é de 2.500 indivíduos/ano/empresa (IBAMA, 2004).

Grande parte das espécies marinhas comercializadas no aquarismo é de origem tropical e extraída dos recifes. O potencial para a sobreexploração desta atividade é alto, uma vez que indivíduos juvenis, na maioria das vezes, menores que 10 cm de comprimento e de muito valor são retirados em grandes quantidades (Wood, 2001). Além dos efeitos diretos causados pela extração de peixes para o aquarismo é consenso os danos provocados no ecossistema marinho pelas práticas destrutivas utilizadas para coletar estes organismos (Wood, 1985, 2001; Nottingham et al., 2005). Somada à pressão pesqueira sofrida por esta espécie por meio do aquarismo, *P. paru* é constantemente capturada como *bycatch* em armadilhas de pesca no Nordeste do Brasil (Feitosa et al., 2008). As armadilhas têm sido responsáveis pelo colapso de muitos recursos recifais (Wolff et al., 1999), principalmente devido à pouca seletividade do petrecho. No caso específico de *P. paru*, espécimes de 15 a 40 cm de comprimento foram pescadas pelas armadilhas (Feitosa et al., 2008). Dentre os peixes ornamentais capturados por este petrecho, aqueles dos gêneros *Acanthurus*, *Chaetodon*, *Bodianus*, *Holacanthus* e *Pomacanthus* são frequentemente os mais capturados (Garrison et al., 1994; Wolff et al., 1999).

A sobreexploração de espécies de peixes marinhos pela captura para fins ornamentais e a destruição de habitats são apontados como os principais problemas gerados pelo aquarismo. Portanto, recomenda-se que sejam realizados levantamentos estatísticos sobre o número de peixes ornamentais envolvidos no mercado, além de estudos sobre a biologia e dinâmica populacional destas espécies (Andrews, 1990; Edwards & Shepherd, 1992; Wood, 2001).

A demanda global por peixes ornamentais tem resultado no aumento direto dos índices de coletas nos estoques naturais (Ogawa & Brown, 2001). Pouco se sabe sobre a dinâmica populacional destas espécies, pois a quantidade de espécies exportadas é baseada em entrevistas com coletores que muitas vezes não falam a realidade. A situação das populações de peixes recifais é inconclusiva, principalmente devido a variações sazonais

naturais, bem como as alterações no esforço de pesca para um dado ano de estudo (Ogawa & Brown, 2001).

Nas últimas reuniões técnicas, promovidas pelo IBAMA, sobre a exploração de peixes ornamentais marinhos, foi enfatizada a carência de pesquisas sobre a biologia e dinâmica populacional das principais espécies exportadas pelo Brasil (e.g. *Holacanthus ciliaris* e *Pomacanthus paru*) (IBAMA, 2000, 2003). Desta forma, espera-se que os resultados obtidos na presente pesquisa forneçam subsídios fundamentais à criação de medidas de regulamentação, bem como proporcionem dados necessários ao adequado manejo de suas populações.

## Referências

- Aburto-Oropeza, O.; Sala, E.; Sánchez-Ortiz, C. 2000. Feeding behavior, habitat use & abundance of the angelfish *Holacanthus passer* (Pomacanthidae) in the Southern Sea of Cortés. *Environmental Biology of Fishes*, 57: 435-442.
- Aiken, K. 1983. The biology, ecology & bionomics of the butterfly & angelfishes, Chaetodontidae. p. 155 – 165. In: Munro, J. L. (ed). *Caribbean coral reef fisheries resource*. Iclarm studies & reviews 7. Manila, Philippines.
- Affonso, P. R. A. M.; Galetti Jr., P. M. 2007. Genetic diversity of three ornamental reef fishes (Families Pomacanthidae & Chaetodontidae) from the Brazilian coast. *Brazilian Journal of Biology*, 67: 925-933.
- Aldenhoven, J. M. 1986. Different reproductive behavior strategies in sex changing coral reef fish *Centropyge bicolor* (Pomacanthidae). *Australian Journal of Marine & Freshwater Research*, 37: 353-360.
- Allen, G. R. 1979. *Butterfly and angelfishes of the world*. Mergus Publishers, Melle.
- Allen, G.R.; Steene, R.; Allen, M. 1998. *A guide to angelfishes & butterflyfishes*. Odyssey Publishing, USA/ Tropical Reef Research, Australia.

- Andrews, C. 1990. The ornamental fish trade & conservation. *Journal of Fish Biology*, 37: 53-59.
- Bauer, J. A.; Bauer, S. E. 1981. Reproductive biology of pigmy angelfishes of the genus *Centropyge* (Pomacanthidae). *Bulletin of Marine Science*, 31: 495-513.
- Belwood, D. R.; van Herwerden, L.; Konow, N. 2004. Evolution & biogeography of marine angelfishes (Pisces: Pomacanthidae). *Molecular Phylogenetics & Evolution*, 33: 140-155.
- Bowen, B. W.; Muss, A.; Rocha, L. A.; Grant, W. S. 2006. Shallow mtDNA Coalescence in Atlantic Pygmy Angelfishes (Genus *Centropyge*) Indicates a Recent Invasion from the Indian Ocean. *Journal of Heredity*, 97: 1–12.
- Cervigón, F. 1993. Los peces marinos de Venezuela. Fundación Científica de Los Roques, Venezuela.
- Chung, K. C.; Woo, N. Y. S. 1998. Phylogenetic relationships of the Pomacanthidae (Pisces: Teleostei) inferred from allozyme variation. *Journal of Zoology*, 246: 215-231.
- Chung, K.C.; Woo, N.Y.S. 1999. Age and growth by scale analysis of *Pomacanthus imperator* (Teleostei: Pomacanthidae) from Dongsha Islands, southern China. *Environmental Biology of Fishes*, 55: 399-412.
- Edwards, A.J.; Shepherd, A.D. 1992. Environmental implications of aquarium-fish collection in the Maldives, with proposals for regulation. *Environmental Conservation*, 19: 61-72.
- Feitosa, C. V.; Ferreira, B. P.; Araújo, M. E. 2008. A rapid new method for assessing sustainability of ornamental fish by-catch from coral reefs. *Marine and Freshwater Research*, 59: 1092-1100.
- Ferreira, C.E.L.; Floeter, S.R.; Gasparini, J.L.; Joyeux, J.C.; Ferreira, B.P. 2004. Trophic structure patterns of Brazilian reef fishes: a latitudinal comparison. *Journal of Biogeography*, 31: 1093 – 1106.
- Galetti Jr., P. M.; Molina, W. F.; Affonso, P.A. M.; Aguilar, C. T., 2006. Assessing genetic diversity of Brazilian reef fishes by chromosomal and DNA markers. *Genetica*, 126: 161-177.

- Garrison, V. H.; Rogers, C. S.; Beets, J. 1994. Of reef fish, overfishing and *in situ* observations of fish traps in St. John, U. S. Virgin Islands. *Revista de Biologia Tropical*, 46: 41 – 59.
- Gasparini, J. L.; Floeter, S. R.; Ferreira, C. E. L.; Sazima, I. 2005. Marine ornamental trade in Brazil. *Biodiversity Conservation*, 14: 2883-2899.
- Hamagushi, Y.; Sakai, Y.; Takasu, F.; Shigesada, N. 2002. Modelling spawning strategy for sex change under social control in harem angelfishes. *Behavioral ecology*, 13: 75-82.
- Hourigan, T. F.; Stanton, F. G.; Motta, P. J.; Kelley, C. D.; Carlson, B. 1989. The feeding ecology of three species of Caribbean angelfishes (family Pomacanthidae). *Environmental Biology of Fishes*, 24: 105-116.
- IBAMA. 2000. Projeto Peixes ornamentais marinhos: ordenamento da captura e comercialização. Relatório final, Tamandaré-PE.
- IBAMA. 2003. Relatório da reunião sobre a exploração de peixes ornamentais marinhos no Brasil. Fortaleza- CE.
- IBAMA. 2004. Instrução normativa nº 14, de 18 de fevereiro de 2004.
- IBAMA. 2007. Peixes ornamentais marinhos: Estatística de exportação por espécie para 2007. [http://www.ibama.gov.br/recursos-pesqueiros/wp-content/files/quantidades \\_exportadas \\_2007 \\_marinho.pdf](http://www.ibama.gov.br/recursos-pesqueiros/wp-content/files/quantidades_exportadas_2007_marinho.pdf)
- IUCN 2008. 2008 IUCN Red List of Threatened Species. <[www.iucnredlist.org](http://www.iucnredlist.org)>. Downloaded on 26 January 2009.
- Kulbicki, M; Sarramégn, S. 1999. Comparison of density estimates derived from a strip transect and distance sampling for underwater visual censuses: a case study of Chaetodontidae e Pomacanthidae. *Aquatic Living Resources*, 12: 315-325.
- Kuwamura, T. 1998. New aspects of sex change among reef fishes: recent studies in Japan. *Environmental Biology of Fishes*, 52: 125-135.

- Menezes, N.; Figueiredo, J. L. 1985. Manual de peixes marinhos do sudeste do Brasil. V. Teleostei (4). EDUSP, São Paulo, 105p.
- Monteiro Neto, C.; Cunha, F. E. A.; Nottingham, M. C.; Araújo, M. E.; Rosa, I. L. & Barros, G. M. L. 2003. Analysis of the marine ornamental fish trade at Ceará State, northeast Brazil. *Biodiversity and Conservation*, 12: 1287 – 1295.
- Moyer, J. T.; Thresher, R. E.; Colin, P. L. 1983. Courtship, spawning and inferred social organization of American angelfishes (Genera *Pomacanthus*, *Holacanthus* and *Centropyge*; Pomacanthidae). *Environmental Biology of Fishes*, 9: 25-39.
- Munro, J.L.; Gaut, V.C.; Thompson, R.; Reeson, P. 1973. The spawning seasons of Caribbean reef fishes. *Journal of Fish Biology*, 5: 69-84.
- Nelson, J. S. 1994. *Fishes of the world*. John Wiley & Sons Inc, USA. 600pp.
- Nelson, J. S. 2006. *Fishes of the world*. John Wiley & Sons Inc, USA. 622pp.
- Nottingham, M. C.; Barreto, L. M.; Araujo, M. E.; Monteiro-Neto, C.; Cunha, F. E.; Rosa, I. M. L.; Alencar, C. A. G. 2005. A exploração de peixes ornamentais marinhos no Estado do Ceará, Brasil: captura, manutenção nas empresas e exportação. *Boletim Técnico- Científico do CEPENE*, 13: 53-73.
- Ogawa, T.; Brown, C. L. 2001. Ornamental reef fish aquaculture & collection in Hawaii. *Aquarium Sciences and Conservation*, 3: 151-169, 2001.
- Randall, J. E. 1967. Food habits of reef fishes of the West Indies. *Studies in Tropical Oceanography* (Miami), 5: 665-847.
- Randall, J. E.; Hartman, W. D. 1968. Sponge-feeding fishes of the West Indies. *Marine Biology*, 1: 216-225.
- Roberts, C. M.; Shepherd, A. R. D.; Ormond, R. F. G. 1992. Large-Scale Variation in Assemblage Structure of Red Sea Butterflyfishes and Angelfishes. *Journal of Biogeography*, 19: 239-250.

Sakai, Y. 1986. Fecundity of female angelfish, *Centropyge ferrugatus*, independent of body size: field collection of spawned eggs. Ichthyological Research, 43: 186-189.

Sakai, Y.; Kohda, M. 1997. Harem structure of the protogynous angelfish, *Centropyge ferrugatus* (Pomacanthidae). Environmental Biology of Fishes, 49: 333 – 339.

Sazima I.; Moura R.L.; Sazima, C. 1999. Cleaning activity of juvenile angelfish, *Pomacanthus paru*, on the reefs of the Abrolhos Archipelago, western South Atlantic. Environmental Biology of Fishes, 56, 399-407.

Smith, M. M.; Heemstra, P. C. 1991. Smith's sea fishes. Southern Book Publishers, Johannesburg, 1048p., 144pls.

Wolff, N.; Grober-Dunsmore, R.; Rogers, C. S.; Beets, J. 1999. Management implications of fish trap effectiveness in adjacent coral reef & gorgonian habitats. Environmental Biology of Fish, 55: 81-90.

Wood, E. 1985. Exploitation of coral reef fishes for the aquarium trade, a report to the Marine Conservation Society UK, 129p.

Wood, E. 2001. Collection of coral reef fish for aquaria: global trade, conservation issues and management strategies. Marine Conservation Society, Herefordshire, UK.

**Capítulo 1 - Reprodução do Paru preto *Pomacanthus paru* (Teleostei: Pomacanthidae) no nordeste do Brasil.**

**Chapter 1\* - Reproduction of the French Angelfish *Pomacanthus paru* (Teleostei: Pomacanthidae) from Northeastern Brazil.**

\* Capítulo em formato de artigo e padronizado de acordo com as normas da revista *ICES Journal of Marine Science*.



**Reproduction of the French Angelfish *Pomacanthus paru* (Teleostei: Pomacanthidae) from Northeastern Brazil.**

Caroline Vieira Feitosa<sup>a, b</sup>, Beatrice Padovani Ferreira<sup>a</sup>, Maria Elisabeth de Araújo<sup>a</sup>,

**Abstract**

*Pomacanthus paru* is an important species for the Brazilian aquarium trade, being the second most exported. The aim of this study was to describe the reproductive biology of *P. paru*. Fish were caught as bycatch in trap fishery. Weekly samplings were performed from March 2006 to February 2007. Total length (TL), standard length (SL) and body weight (BW) were recorded. Gonadosomatic index (GSI) and batch fecundity were calculated. Individuals were sexed and the maturity stages were determined. The French angelfish is a gonochoristic fish and presents size dimorphism. Sex ratios were biased toward female at all months, except October. The L50 were different between sexes, being maturity reached at 30cm for females and 35 cm for males. *Pomacanthus paru* is reproductively active throughout the year, with females presenting two distinct spawning peaks. This species is mainly targeted by the aquarium trade, but it is also caught as bycatch. Despite the determination of the size at maturity, this information is not relevant for the aquarium trade management, as most commercialized ornamental fish are juveniles under 10 cm in standard length. It can be suggested that conservationist issues should be based on capture per area and the establishment of protected areas.

**Key words:** *Pomacanthus paru*, sex ratio, size at maturity, size dimorphism, spawning season.

a - Departamento de Oceanografia, Centro de Tecnologia e Geociências, Universidade Federal de Pernambuco, 50670-901, Recife, Pernambuco, Brazil.

25 b- Corresponding author - Email: carol\_feitosa@hotmail.com. Tel.: + 55 – 81 –  
26 21267223; Fax: + 55 – 81 – 32278227

## 27    **Introduction**

28            Knowledge on fish reproduction is especially important for targeted species.  
29    Size at first maturity, reproductive behavior and fecundity are essential characteristics  
30    for the management of fisheries (Wootton, 1979; Jennings *et al.*, 2001).

31            Pomacanthids, or angelfish, are among the most colorful and widely recognized  
32    group of reef fish (Thresher, 1982), and a favorite for divers and aquarists. Angelfish  
33    occur throughout tropical and subtropical seas and are associated with coral reefs. The  
34    biology of Pomacanthidae family has caused special concern among reef ichthyologists,  
35    particularly due to its habit of forming harems (Allen *et al.*, 1998). The genera  
36    *Centropyge* and *Geniakanthus* (the smallest angelfish), are the most studied in terms of  
37    gonad development and reproductive behavior (Bauer and Bauer, 1981; Moyer *et al.*,  
38    1983; Aldenhoven, 1986; Sakai, 1986; Sakai and Kohda, 1997). The reproductive  
39    behavior of angelfish is relatively well known but there is little information on the  
40    spawning habits of *Pomacanthus* angelfish (Allen, 1979). *Pomacanthus* angelfish are  
41    known to be gonochoristic (Moyer *et al.*, 1983; Thresher, 1984) but further studies are  
42    needed to state this for all members of the genus (Michael, 2004).

43            *Pomacanthus paru* is distributed from Florida, the Bahamas and northern Gulf  
44    of Mexico to Santa Catarina State - Brazil (Menezes *et al.*, 2003). It is also reported for  
45    St. Paul's Rocks (Feitoza *et al.*, 2003) and Ascension Island (Maugé, 1990) in the  
46    Central Atlantic and Gulf of Guinea. Maximal length is 41 cm (Cervigón, 1993). This  
47    species occurs in waters from 5 to 100 m, but juveniles and subadults are more common  
48    in shallow waters (Allen *et al.*, 1998; Michael, 2004). The French angelfish is an  
49    omnivore. Sponges are its main food item, constituting nearly 75% of its diet (Randall,  
50    1967). It appears that adult usually form stable pairs, although in areas where the fish is  
51    more common it is known to form small harems (Allen *et al.*, 1998; Michael, 2004).

52 Most species collected for the Brazilian aquarium trade belong to the  
53 Pomacanthidae family (Araújo and Albuquerque Filho, 2005). *Pomacanthus paru* is an  
54 important species in this trade and has been one of the most exported species since 2000  
55 (Araújo and Albuquerque Filho, 2005; Nottingham *et al.*, 2000; Monteiro-Neto *et al.*,  
56 2003). During the period of 1995 and 2000, only the Ceará State (Northeastern Brazil)  
57 exported 22. 969 specimens of *P. paru* (Monteiro-Neto *et al.*, 2003). This species is  
58 managed through a Federal Act (56/04) established in 2 004 with the purpose of  
59 managing activity through the establishment of export quotas. Besides the impact from  
60 aquarium trade on juveniles, this species is frequently caught by traps. Trap fisheries  
61 catch a broad length range of ornamental species (e.g. *Holacanthus ciliaris* and  
62 *Pomacanthus paru*), including individuals that make up the breeding stock (Feitosa *et*  
63 *al.*, 2008). There is no regulation for this type of fishery in Brazil.

64 Studies on reproduction are very important for fishery management (Fonteles –  
65 Filho, 1989), as management decisions are based on reproductive parameters, such as  
66 size at maturity. There have been few studies on the sexuality, sexual maturity,  
67 fecundity (Aiken, 1983) and spawning season of pomacanthids (Munro *et al.*, 1973).  
68 For *P. paru* the sample sizes in the two studies cited were very small (four and 21  
69 individuals, respectively). Thus, there is no substantial information on the population  
70 structure of this species.

71 The aim of the present study was to describe the reproductive biology of the *P.*  
72 *paru*. The specific goals of the study were to investigate (a) gonad development, (b)  
73 spawning season, (c) gonad morphology and histology, (d) fecundity and (e) size at  
74 sexual maturity in order to provide useful information for the management of the fishery  
75 targeting this species.

76

## 77 Materials and methods

### 78 Sample collection

79 The fish were collected from commercial fishing boats of the fishing colony of  
80 Itamaracá Island located on Pilar beach ( $7^{\circ}45.30'S$   $34^{\circ}49.44'W$ ) (state of Pernambuco,  
81 Northeastern Brazil – Fig. 1). The specimens were caught as bycatch while trap fishing  
82 for the spotted goatfish (*Pseudupeneus maculatus*) and parrotfish (*Sparisoma* spp). In  
83 this fishery, traps are deployed unbaited next to reefs and hauled 6 hours later.  
84 Occasionally, depending on factors such as wind and currents, some traps are left for  
85 longer periods and only hauled after 48 hours.



86  
87 Figure 1: Location of the sampled site.

88 Weekly samplings were carried out from March 2006 to February 2007. As  
89 individuals smaller than 15 cm are not caught by traps, they were bought from fish  
90 collectors between September 2006 and February 2007.

91           The fresh sampled fish were placed in polystyrene chests with ice and taken to  
92   the Laboratório de Nécton e Aquicultura of the Departamento de Oceanografia of  
93   Universidade Federal de Pernambuco. Total length (TL), standard length (SL) and body  
94   height (BH) and weight (BW) were recorded from the specimens to the nearest 1 mm  
95   and to the nearest 0.01 g. The gonadosomatic index (GSI) was used as an index of  
96   reproductive activity and was calculated as follows:  $[GSI = (GW/BW) \times 100]$ , in which  
97   GW is the gonadal weight (g) and BW is the fish weight. For the batch fecundity  
98   analysis, 0.100 g of the ovary were removed and separated in glycerine with the aid of a  
99   brush and knife; and the hydrated oocytes in this sample were counted (Hunter and  
100   Macewicz, 1985a; Hunter and Goldberg, 1980; Vazzoler, 1996). Relative fecundity was  
101   expressed as the number of eggs/gram of ovary-free body weight. Oocyte diameter was  
102   measured under a microscope at 100 x with a video camera attachment, using the  
103   ImageLab software program (Softium).

#### 104   **Sample processing**

105           The gonads were dissected from the fish, weighed to the nearest 0.01 g, fixed in  
106   Bouin's solution for 24 h and transferred to 70% ethanol for later sectioning. The mid  
107   portion of the gonad was embedded in paraffin, following standard histological  
108   techniques. The paraffin blocks were sectioned transversely at 6 µm thickness by a  
109   rotary microtome. Duplicate slides were prepared for each tissue and stained with  
110   Mayer's hematoxylin-eosin, following standard histological procedures. Individuals  
111   were sexed and the maturity stages were determined adopting the stages proposed by  
112   Murphy and Taylor (1990), West (1990) and Yamaguchi *et al.* (2006) (Table 01). Sex  
113   and reproductive condition were determined for each individual based on histological  
114   evaluations under a microscope at 40 – 400 x magnification.

115

116 Table 1: Macroscopic and histological stages used to describe ovaries and testes of  
 117 *Pomacanthus paru* based in Murphy and Taylor (1990), West (1990) and Yamaguchi *et*  
 118 *al.* (2006).

Gonad stage by sex	Main characteristics
Females	
Immature	Difficult to determine sex macroscopically. Ovaries small and threadlike, transparent. Most advanced oocytes are at peri-nucleolus stage or yolk vesicle stage.
Maturing	The gonads present reduced dimensions ( $\approx 6$ cm in length and 1.80 cm in width). Ovaries varies in color from pink to pale yellow. Most oocytes are in early vitellogenesis stage. Oocytes in late vitellogenesis stage also can be seen. Besides, brown bodies are also observed.
Ripening	Ovaries are yellow in color. An increase in blood vessels, volume and size of the structure are recorded. Most oocytes are in late vitellogenesis stage. Few oocytes are in early vitellogenesis stage. Brown bodies are also registered.
Spawning	Gonads very developed. Ovaries coloration varies from yellow to orange. Migratory nucleus, hydrating or hydrated oocytes. Brown bodies recorded.
Resting	Most advanced oocytes are at peri-nucleolus stage or yolk vesicle stage. Due to the diameter of the gonad and the thickness of the gonad wall, it was possible to differ the resting ovaries differ from the immature ones.

Gonad stage by sex	Main characteristics
Males	
Immature	Difficult to determine sex macroscopically. Testes small and threadlike, transparent. Testes with spermatogonia in the first spermatogenesis stage.
Maturing	Testes transparent or pale white. Tissue predominantly comprised of primary and secondary spermatocytes. Few quantities of spermatids in lobules.
Ripening	White testes. Tissue consists predominantly of spermatocytes, spermatids and spermatozoa. Spermatozoa present in lobules, but none in spermatic ducts.
Spawning	White testes, enlarged. Mature spermatozoa fill the spermatic ducts.
Spent	Testes dull brown in colour. Developed lobules containing few remaining sperm.
Resting	Flat, white-grayish testes. Spermatogonia in the first spermatogenesis stage.

119

## 120 **Statistical analysis**

121       The Mann-Whitney test was used to determine possible sexual dimorphism  
122 based on data on standard length and body height for females and males. A  $\chi^2$  goodness-  
123 of-fit test was performed to compare female to male (F: M) ratios (per month and within  
124 size groups), with a hypothesized sex-ratio of 1:1. Differences in size at maturity and  
125 size between sexes within length classes were performed using the Mann-Whitney test.

126       The Kruskal-Wallis test was used to determine differences in gonadosomatic  
127 indices among months. Batch fecundity was calculated using the following equation:



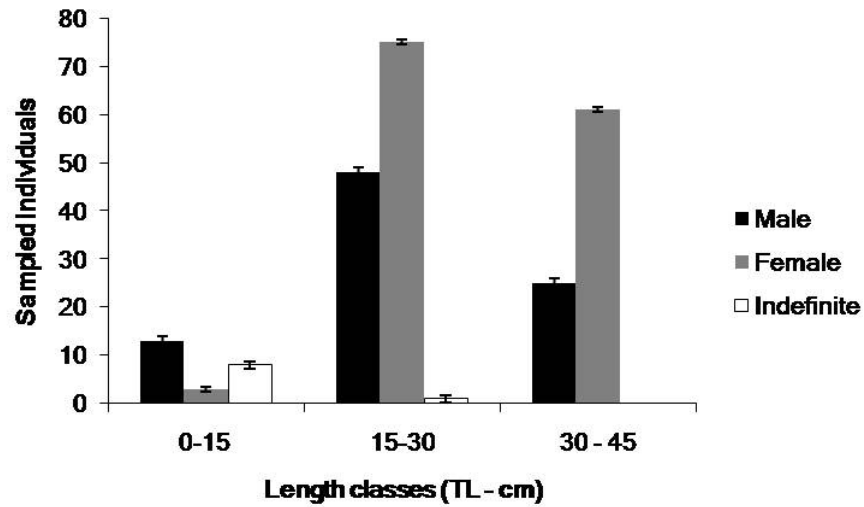
128 BF =  $nWg/w$ , in which: BF = batch fecundity, n = number of hydrated oocytes in the  
129 sample, Wg = the gonad weight and w = the sample weight. The number of batches was  
130 calculated based on the method proposed by Hunter and Macewicz (1985b). The  
131 significance level for all analyses was set at  $P < 0.05$ . The BioEstat 3.0 software  
132 program (Instituto de Desenvolvimento Sustentável Mamirauá – Amazonas) was used  
133 for these analyses.

134

## 135 **Results**

### 136 **Sex ratio and size composition**

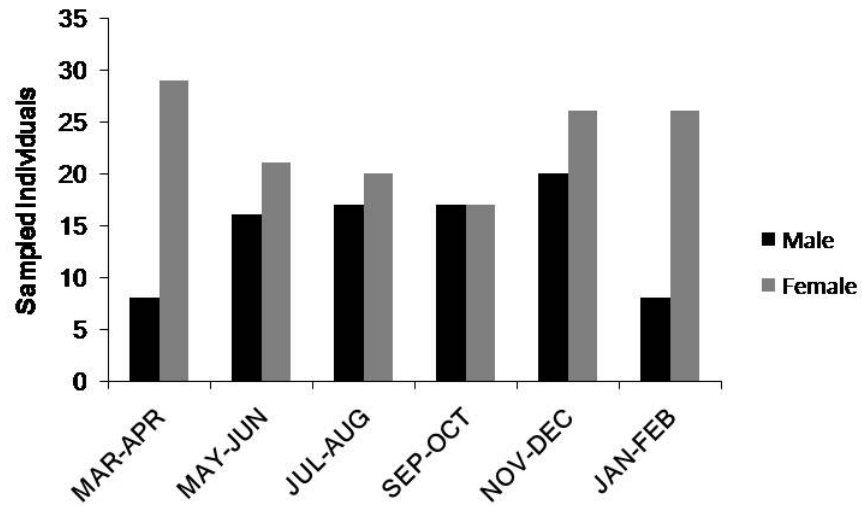
137 The French angelfish, *Pomacanthus paru*, is a gonochoristic fish. Differences  
138 between sexes were found in total length ( $Z(U) = 3.1200$ ,  $P = 0.0018$ ), standard length  
139 ( $Z(U) = 4.2672$ ,  $P = 0.0000$ ), body height ( $Z(U) = 2.96$ ,  $P = 0.0030$ ). Among 236  
140 individuals sexed, 139 were female (58.89%), 86 were male (36.44%) and 11 were  
141 undefined (4.66%) (Fig.2). Sex ratios were biased toward females in all bimesters,  
142 except September-October. Bimonthly sex ratios were statistically significant only in  
143 March-Apr/06 ( $\chi^2 = 6.48$ ,  $P = 0.01$ ) and January-February/07 ( $\chi^2 = 5.12$ ,  $P = 0.02$ ), where  
144 females were 3.63-fold and 3.25-fold, more abundant than males (Fig. 3). Differences in  
145 sex ratios for size classes were recorded only among these length classes [15 - 30 cm ( $Z$   
146 ( $U) = 4.19$ ,  $P = 0.0000$ ); 30 - 45 cm ( $Z(U) = 4.8131$ ,  $P = 0.0000$ )] (Fig. 4).



147

148 Figure 2: Size distribution between sexes of sampled individuals with bars indicating  
 149 standard error.

150



151

152

153 Figure 3: Sampled individuals distribution of female and male of *Pomacanthus paru*  
 154 among bimesters.

155

156

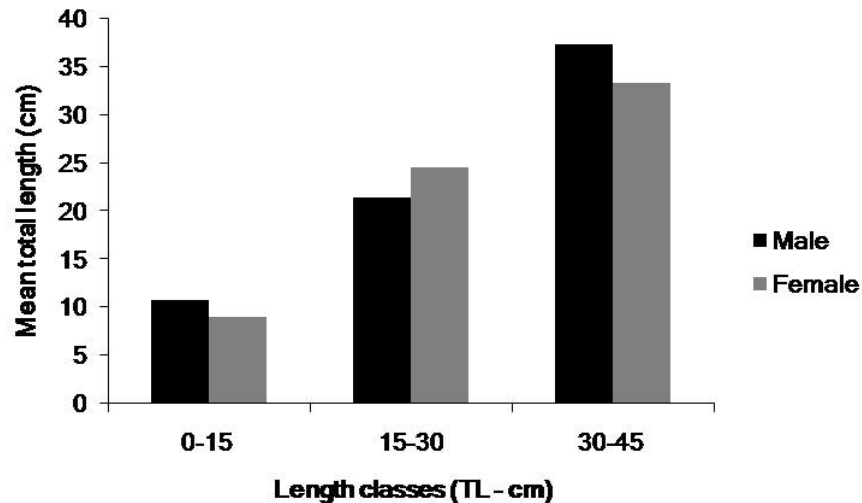


Figure 4: Mean total length distribution of *Pomacanthus paru*.

#### Spawning season and gonadal development

Monthly changes in mean GSI for males and females are presented in Fig. 5 and were not statistically significant for males (K-W test male  $H = 4.6852$   $P = 0.4555$ ), but significant for females in bimesters Mar-Apr and Nov-Dec (K-W test female  $H = 13.4832$   $P < 0.05$ ). The analysis of GSI for females indicates an increase beginning in January-February, with a peak in March-April, and gradual decline until August. GSI attained high levels in September-October, but declined in November-December and then began to rise. For males, the GSI is the highest in March-Apr, as observed in females. The GSI then declined abruptly and continued nearly the same until March.

The immature females occurred from April to January, but the highest frequency was found between July-August and September-October. Maturing and ripening stages occurred throughout the year, except in June, September, October and December for the maturing stage. The spawning stage was dominant in September-October and from January until April. In this period, the GSI increased, mainly between January and April

and in October. Resting ovaries were recorded throughout the year, with the exception of July, September and February. However, the spawning stage was recorded in these months (Fig. 6).

Males in immature stages were only recorded in a few months. The maturing stage was not observed in April, December or February, but spawning and spent stages were recorded. The ripening stage was more frequent in July-August and November-December. From May until October, the spawning stage was dominant. Individuals in the spent stage were recorded throughout the year. Resting testes were not recorded only in March-April and September-October (Fig. 6).

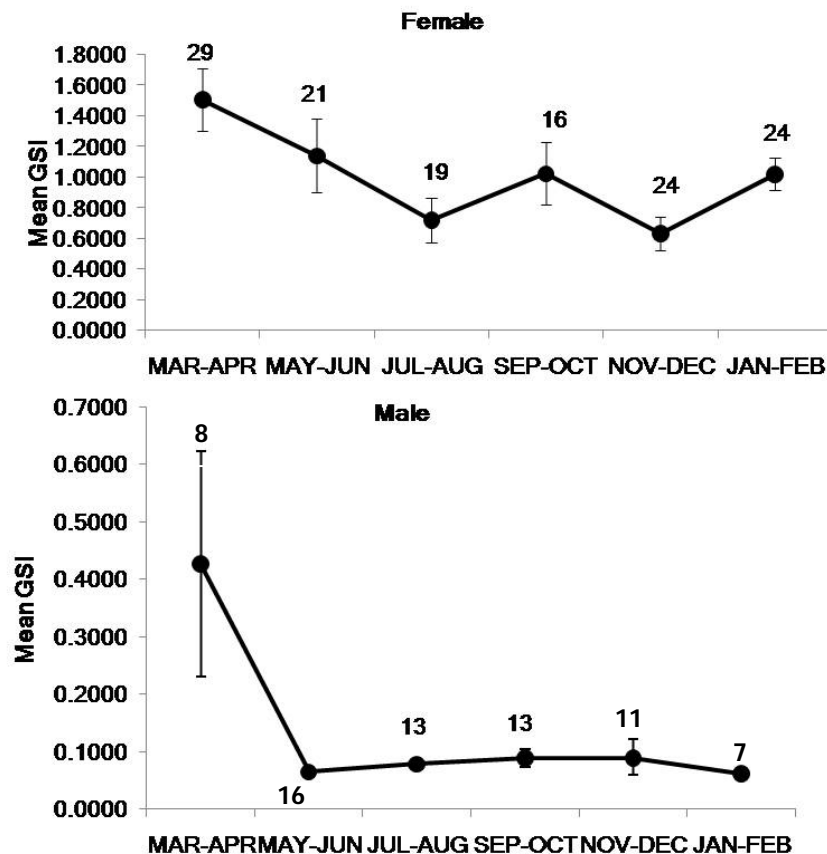
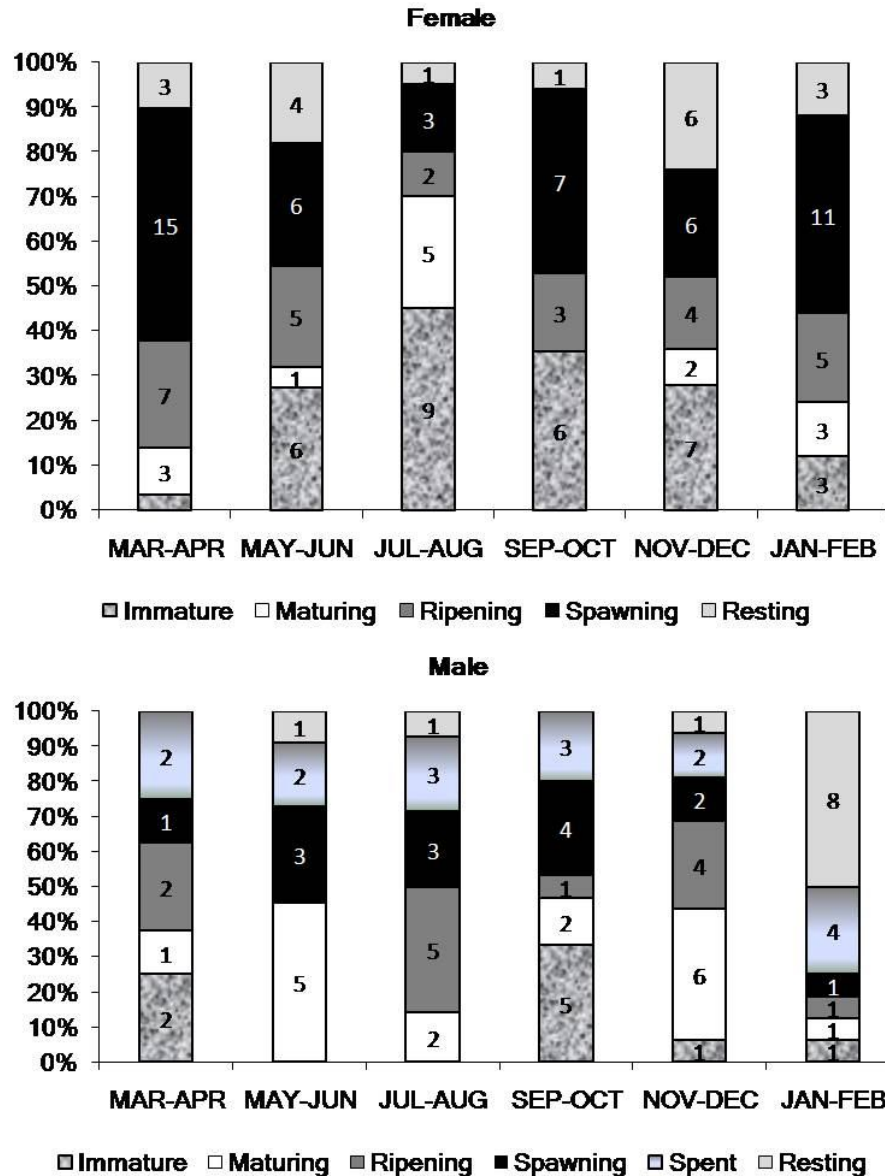


Figure 5: Bimonthly changes of the mean gonadosomatic index (GSI) in female and male *Pomacanthus paru* with bars and values indicating the standard error and the number of individuals analyzed for each stage, respectively.



188

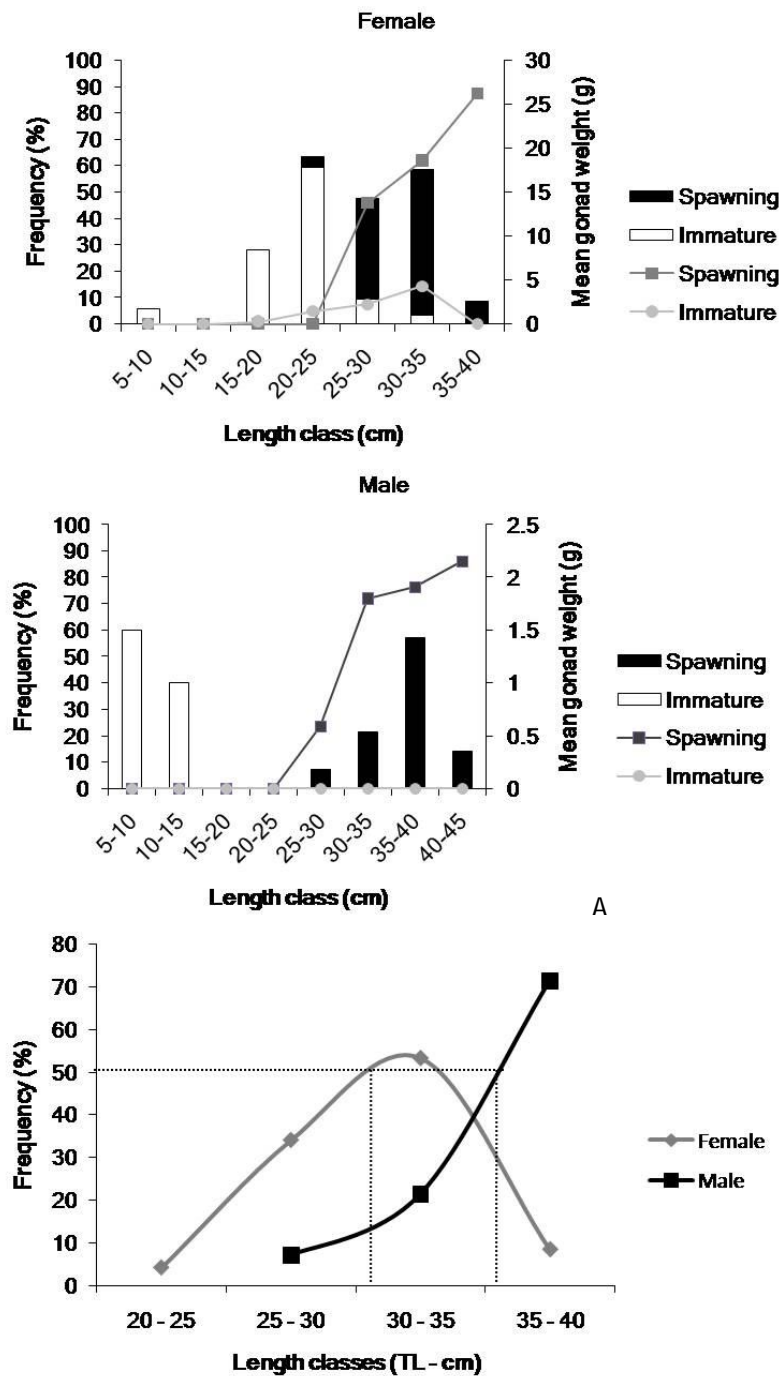
189

190 Figure 6: Bimonthly percentage frequencies of occurrence of gonadal developmental  
 191 stages in female and male *Pomacanthus paru*. The values inform the number individual  
 192 analyzed for each stage.

### 193 Size at maturity

194 Individuals fit for reproduction appeared at 20 to 25 cm for females and 25 to 30  
 195 cm TL for males. The L50 were different between sexes, being maturity reached at

196 30cm for females and 35 cm for males ( $Z(U) = 4.3532$   $P = 0.0000$ ). The smallest mature  
 197 female and male were 23.18 cm TL and 25.24 cm TL, respectively (Fig. 7).



198

199

Figure 7: Percentage of individuals in spawning and immature stage versus length class and mean gonad weight for female and male (A) and graphic showing the L<sub>50</sub> for female and male of *Pomacanthus paru* (B).

## Fecundity

Table 2 displays estimates of the minimum, maximum and mean values for the batch and relative fecundities. Hydrating oocytes ranged in size from 350 to 800µm.

Table 2: Minimum, maximum and mean values of batch (BF), total (TF) and relative (RF) fecundities of *Pomacanthus paru*.

	TL* (cm)	BW* (g)	BF	RF
Minimum value	23.18	428	109.6	4.73
Maximum value	34.24	1 354	29 475	860.83
Mean value ± SE			3.751.62 ±1.396.78	118.48 ± 41.42

\*TL = total length, BW = body weight

## Discussion

### Sex ratio and size composition

In the present study, *Pomacanthus paru* was classified as a gonochoric species. An overlap in size distributions, similar mean sizes of adult males and females and the lack of histological evidence for hermaphroditism support this sexual pattern. Hermaphroditism is a common process in tropical fish families and is recorded in, at least, eighteen families (Ross, 1990). In the family Pomacanthidae, protogynous hermaphroditism has been found in three genera: *Centropyge* (Moyer and Nazakono, 1978; Bruce, 1980), *Geniakanthus* (Bruce, 1980) and *Holacanthus* (Hourigan and Kelley, 1985, Nottingham *et al.*, 2003). However, it has been suggested that *Pomacanthus* species are gonochoristic (Michael, 2004). In most gonochoric fish, size

distributions of females and males overlap broadly. However, the population structure alone is not a reliable indicator of sexual pattern, as bimodal size may be observed in gonochoric species that possess size dimorphism (Sadovy, 1986) or differential rates of growth, maturation, or mortality by sex (Sadovy and Shapiro, 1987).

Species that occur in pairs are commonly gonochoric, as was observed for *Pomacanthus paru* in the present study. However, the gonochoric pattern also occurs in fishes with group-spawning mating systems, where all males have access to females, and sperm competition is intense (Warner, 1982). The mating system of the genus *Pomacanthus* depends on the population density in a given location. In some areas, *P. paru* form stable pairs, whereas in locations in which the species is abundant this fish forms harems (Michael, 2004). However, some authors state that *Pomacanthus paru* is consistently found in pairs, regardless of whether the species is abundant or not (Moyer *et al.*, 1983). Despite the common occurrence of pairs, spawning involves pairs and solitary individuals, as observed by Thresher (1980) and Moyer *et al.* (1983).

Sexual dimorphism is uncommon among marine teleost fish. It can be manifested as changes in body shape during growth, which is related to protogynous hermaphroditism. However, in gonochoristic species, size differences between sexes have also been recorded (García-Cagide *et al.*, 2001), as was observed for *Pomacanthus paru*. Sexual size dimorphism is a ubiquitous characteristic of angelfish, for which males are generally larger than females. Size dimorphism and sexual dichromatism have been recorded in some species of the genus *Pomacanthus* (Thresher, 1982). Feitosa *et al.* (unpublished data - chapter 2) observed a different pattern of growth between sexes, in which males continued to grow, while females already reached the asymptotic length. Reinforcing the size dimorphism for *Pomacanthus*, Moyer *et al.* (1983) stated that



245 sexual size dimorphism is not conspicuous in most adult pairs of *P. paru*, however after  
246 a close examination, a slight size difference between paired individuals can be noticed.

247 Sex ratio is a population characteristic that plays an important role in  
248 reproduction. In species of snappers, jacks and grunts, females are more abundant in  
249 almost all length classes and reach a larger size than males (García-Cagide *et al.*, 2001).  
250 In these cases, female dominance may be explained by a greater survivorship, but also  
251 by differences in habitat preference (García-Cagide *et al.*, 2001). According to Aburto-  
252 Oropeza *et al.* (2000), the sex ratio of angelfish populations (proportion of sexes and  
253 sizes) seems to be determined by the mating system. In the family Pomacanthidae, there  
254 are different reproductive styles among genera (Allen, 1979) and most are protogynous  
255 hermaphrodite (Thresher, 1984). Few studies, however, have addressed the role of other  
256 factors, such as food availability, in the structuring of angelfish communities (Arellano-  
257 Martínez *et al.*, 1999; Aburto-Oropeza *et al.*, 2000).

258 Aburto-Oropeza *et al.* (2000) observed differences in habitat preference between  
259 sexes in the angelfish *Holacanthus passer*, for which a significant habitat overlap was  
260 only recorded between large females and small males. For this angelfish, sex differed  
261 significantly over the size range, with females prevailing at smaller lengths and males at  
262 larger lengths (Arellano-Martínez *et al.*, 1999). It may be suggested that there is a  
263 habitat preference between sexes in *Pomacanthus paru*, as a similar pattern to that  
264 described by Arellano-Martínez *et al.* (1999) was recorded in the samples of the present  
265 study. In a study performed in Caribbean waters, Aiken (1983) found that the sex ratios  
266 of the various species of chaetodontids and pomacanthids caught in traps were female  
267 biased, mainly in *Pomacanthus arcuatus* (M:F – 1:2.51) and *Pomacanthus paru* (M:F –  
268 1:3.28). The author attributes these variations to differences in sample site depths.

269           Other explanations for this pattern are that females enter fish traps more readily  
270 than males (Gaut and Munro, 1983) or that females fit for reproduction move into the  
271 sampling area and are more susceptible to traps, due to their reproductive status.  
272 Females are especially more abundant in samples in months related to the spawning  
273 peak. As smaller individuals between 0 and 15 cm TL (length class dominated by  
274 males) were not caught by traps, the female dominance of *P. paru* may be explained by  
275 these factors.

276

### 277 **Spawning season and gonadal development**

278           Although female of *Pomacanthus paru* has two reproductive peaks and male  
279 one, the species is probably capable of spawning throughout the year, based on the  
280 occurrence of females in the spawning stage and males in the spent stage in nearly every  
281 month. In Caribbean waters, ripe individuals of *P. paru* have been collected in May,  
282 July, August, October and November and from January to August, with a maximal  
283 proportion in April for *Holacanthus ciliaris* (Munro *et al.*, 1973). For *Pomacanthus*  
284 *arcuatus*, the greatest proportions of ripe fishes were collected in October and January,  
285 but according to Munro *et al.* (1973), there is no discernible seasonal trend in the data.  
286 Ripe individuals of *Holacanthus tricolor* were found in all months, with the exception  
287 of December for which there is no data. According to Moyer *et al.* (1983), *H. tricolor*  
288 spawns most of the year in Puerto Rico. Arellano-Martínez and Ceballos-Vásquez  
289 (2001) also collected ripe fishes of *Holacanthus passer* in April. Although ripe *P. paru*  
290 were collected in every month in the present study, the data presented are in agreement  
291 with those obtained by Munro *et al.* (1973), Moyer *et al.* (1983) and Arellano-Martínez  
292 *et al.* (2001) with regard to spawning months.

293           The gonadosomatic index value is an indicator of the duration of the  
294 reproductive season, but caution should be exercised in its interpretation due to regional  
295 and temporal variations (Jons and Miranda, 1997). In fact, seasonal variations in GSI  
296 must be analyzed together with other factors, such as the monthly proportion of fish in  
297 different stages of gonad development (García-Cagide *et al.*, 2001). In the present study,  
298 the GSI index demonstrated a unique and common peak (March – April) for both sexes.  
299 Two peaks were recorded for females, - a strong peak in January-February and March-  
300 April and a weak one in September-October. A high frequency of males in spawning  
301 stages was recorded in May until October, when the GSI index values were low. In  
302 *Hyporhamphus australis*, some gonads have been found in spawning condition outside  
303 their respective GSI-defined peak spawning periods (Hughes and Stewart, 2006). For  
304 two species of tropical Atlantic hemiramphids it has been demonstrated that, while the  
305 spawning of young females is strongly seasonal, older female fish spawn throughout the  
306 year (McBride and Thurman, 2003). Moyer *et al.* (1983) found a daily spawning cycle  
307 that occurs throughout all or most of the lunar cycle in six American species and fifteen  
308 Western Pacific species of angelfishes. Many tropical fish are intermittent spawners  
309 and, for such individuals, the GSI provides information for only a certain period of the  
310 reproductive cycle, rather than entire year (García-Cagide *et al.*, 2001). Despite the  
311 values of this index, the data of the present study support a pattern of reproductive  
312 activity throughout the year, with two spawning peaks.

### 313 **Size at maturity**

314           Size at maturity is influenced by environmental and genetic factors (Wootton,  
315 1979), but fishing pressure can also affect this parameter (Jennings *et al.*, 2001). Based  
316 on GSI index of *Holacanthus passer*, Arellano-Martinez and Ceballos-Vásquez (2001)  
317 affirm that due to the gonad size, male fish requires less stored energy for gonadal

development and it may start to mature earlier than females. However, the results of the present study demonstrated that *P. paru* females mature earlier than males. Spawning stage first appeared in females at 23 cm TL and in males at 25 cm TL. Besides, the  $L_{50}$  for females was 5 cm smaller than males. The total length of the smallest mature *Pomacanthus paru* female caught in Caribbean waters by Aiken (1983) was 23 cm. The author suggests that males tend to mature at a larger size than females, as no male was captured. The results of the present study corroborate with Aiken (1983).

### **Fecundity**

According to Hunter and Goldberg (1980), the number of hydrated eggs in ovaries prior to ovulation should give the most accurate estimate of fecundity and this method has been used on several *Engraulius* species (Hunter and Goldberg, 1980; Hunter *et al.*, 1984; Hunter and Macewicz, 1985a), as well as on the chub mackerel (*Scomber japonicus*) (Peña *et al.*, 1986) and dwarf herring (García-Cagide *et al.*, 2001). The fecundity obtained in the present study was much lower than that described by Aiken (1983), who found a mean value of 34 200. This difference may be due to the sample size, as Aiken (1983) only analyzed two individuals, whereas 44 females were analyzed in the present study. Among this total, one fish had values similar (29 475) to that determined by Aiken (1983). It may be suggested that the sample size used by Hourigan and Kelley (1985) was insufficient for a comparative analysis. The authors only sampled ten *Holacanthus tricolor* females, - five in May and five in September. The authors recorded a higher number of hydrated oocytes:  $15\,330 \pm 5\,490$  in May and  $35\,840 \pm 18\,480$  in September. Similarities were recorded between the present study and that performed by Arellano – Martínez *et al.* (2006) for *Pomacanthus zonipectus*, in which these authors found a mean values of  $79.4 \pm 9.2$  for relative fecundity. The range

size of hydrating oocytes found in the present study corroborates the results obtained by Hourigan and Kelley (1985) for *Holacanthus tricolor* (363 – 721  $\mu\text{m}$ ) and by Arellano-Martínez *et al.* (2006) for *Pomacanthus zonipectus* ( $675 \pm 1.2 \mu\text{m}$ ).

Besides the sample size issues, the definition of spawning stage without determinate the minimal quantity of these oocytes for this stage should be revised. This definition, based on the simple presence of hydrated oocytes, may be influencing the higher variation recorded in the present results of fecundity.

### Considerations

The present study provides important information on the reproductive biology of *P. paru*. The French angelfish is a gonochoristic species. Some hypotheses, such as habitat preference between sexes, greater female susceptibility to trap fisheries and female migration to sampling areas, may explain the role of population sex structure of *Pomacanthus paru*. However, further studies are needed to clear up this issue. This species is mainly targeted by the aquarium trade. Species-based quotas have been established by the Brazilian environmental protection agency (IBAMA) to ensure that harvests are maintained at a sustainable level. Despite the determination of the size at maturity in the present study, this information is not relevant for the aquarium trade management, as most commercialized ornamental fish are juveniles under 10 cm in standard length. It can be suggested that conservationist issues should be based on quantitative data (capture) per area and the establishment of protected areas.

*Pomacanthus paru* is also commonly caught as bycatch in trap fishery, an unregulated multi-specific activity. Specimens captured as bycatch are not landed and are usually returned to the sea (many still alive, according to the fishermen). The survival of these individuals is doubtful, as the traps are hauled quickly to surface and problems related to decompression have often been reported. Little is known regarding

the impact of this activity, being difficult to manage it. Monitoring and management of this fishery, a growing activity with catches exported to the international market, is mandatory in order to assess the impact of this kind of fishery on both target and non-target species.

## Acknowledgements

This study was sponsored by the Brazilian Scientific Council - Conselho Nacional de Pesquisa e Desenvolvimento (CNPq) and Pernambuco State Scientific Council - Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco (FACEPE).

## References

- Aburto-Oropeza, O.; Sala, E. and Sánchez-Ortiz, C. 2000. Feeding behavior, habitat use, and abundance of the Angelfish *Holacanthus passer* (Pomacanthidae) in the Southern Sea of Cortés. *Environmental Biology of Fishes*, 57: 435-442.
- Aiken, K. 1983. The biology, ecology and bionomics of the butterfly and angelfishes, Chaetodontidae. pp. 155 – 165. *In*: Caribbean coral reef fisheries resource. Ed by J. L. Munro. Iclarm studies and reviews 7, Manila, Philippines, 283pp.
- Aldenhoven, J. M. 1986. Different reproductive behavior strategies in sex changing coral reef fish *Centropyge bicolor* (Pomacanthidae). *Australian Journal of Marine and Freshwater Research*, 37: 353-360.
- Allen, G.R. 1979. Butterfly and angelfishes of the world, vol. 2. Mergus, Melle. 352 pp.
- Allen, G.R.; Steene, R. and Allen, M. 1998. Introducing angelfishes. pp. 8 – 12. *In*: A guide to angelfishes and butterflyfishes. Odyssey Publishing: USA/ Tropical Reef Research: Australia, 250pp.

392 Araújo, M. E. and Albuquerque Filho, A. C. 2005. Biologia das principais espécies de  
 393 peixes ornamentais marinhos do Brasil: uma revisão bibliográfica e documental.  
 394 Boletim Técnico Científico do CEPENE, 13: 109 – 154.

395 Arellano-Martínez, M., Ceballos-Vásquez, B. P., Garcia-Dominquez, F. and Galván-  
 396 Magaña, F. 1999. Reproductive biology of the King Angelfish *Holacanthus passer*  
 397 Valenciennes 1846 in the Gulf of California, Mexico. Bulletin of Marine Science, 65:  
 398 677 - 685.

399 Arellano-Martínez, M. and Ceballos-Vásquez, B. P. 2001. Reproductive activity and  
 400 condition index of *Holacanthus passer* (Teleostei: Pomacanthidae) in the Gulf of  
 401 California, Mexico. Revista de Biología Tropical, 49: 939 – 943.

402 Arellano-Martínez, M., Ceballos-Vásquez, B. P., Olalde, L. H. and Galván-Magaña, F.  
 403 2006. Fecundidad del Ángel de Cortéz *Pomacanthus zonipectus* (Teleostei:  
 404 Pomacanthidae) en la isla Espíritu Santo, Golfo de California, Mexico. Ciencias  
 405 Marinas, 32: 65-71.

406 Bauer, J. A. and Bauer, S. E. 1981. Reproductive biology of pigmy angelfishes of the  
 407 genus *Centropyge* (Pomacanthidae). Bulletin of Marine Science, 31: 495 - 513.

408 Bruce, R. W. 1980. Protogynous hermaphroditism in two marine angelfishes. Copeia,  
 409 1980: 353 - 355.

410 Cervigón, F. 1993. Pomacanthidae. pp. 339 – 346. *In*: Los Peces Marinos de Venezuela.  
 411 Fundación Científica Los Roques, Caracas, 951pp.

412 Feitosa, C. V., Ferreira, B. P. and Araújo, M. E. 2008. A rapid new method for assessing  
 413 sustainability of ornamental fish by-catch from coral reefs. Marine and Freshwater Research,  
 414 59: 1092-1100.

415 Feitoza, B. M., Rocha, L. A., Luiz-Júnior, O. J., Floeter, S. R. and Gasparini, J. L. 2003. Reef  
 416 fishes of St. Paul's Rocks: new records and notes on biology and zoogeography. *Aqua, Journal*  
 417 *of Ichthyology and Aquatic Biology*. 7:61– 82.

418 García-Cagide, A., R. Claro, and B. V. Koshelev. 2001. Reproductive patterns of f ishes  
 419 of the Cuban shelf. *In: Ecology of the marine fishes of Cuba.* pp.73–114. Ed by R.  
 420 Claro, K. C. Lindeman and L. R. Parenti. Smithsonian Institution Press, Washington,  
 421 DC, 253pp.

422 Gaut, V. C. and Munro, J. L. 1983. The biology, ecology and bionomics of the grunts,  
 423 Pomadasydae. pp. 110 – 141. *In: Caribbean coral reef fisheries resource.* Ed by J. L.  
 424 Munro. *Iclarm studies and reviews* 7, Manila, Philippines, 283pp.

425 Hourigan, T. F. and Kelley, C. D. 1985. Histology of the gonads and observations on  
 426 the social behavior of the Caribbean angelfish *Holacanthus tricolor*. *Marine Biology*,  
 427 88: 311-322.

428 Hughes, J. M. and Stewart, J. 2006. Reproductive biology of three commercially  
 429 important Hemiramphid species in south-eastern Australia. *Environmental Biology of*  
 430 *Fishes*, 75: 237-256.

431 Hunter, J. R. and Goldberg, S. R. 1980. Spawning incidence and batch fecundity in  
 432 northern anchovy, *Engraulius mordax*. *Fishery bulletin*, 77: 641 - 652.

433 Hunter, J. R., Lo, N. H. and Leong, R. 1984. Batch fecundity in multiple spawning  
 434 fishes. *In: An egg production method for estimating spawning biomass of pelagic fish:*  
 435 *application to the northern anchovy, Engraulius mordax.* pp. 67 – 77. Edited by R.  
 436 Lasker. U.S. Department of Commerce, NOAA Technical Report NMFS 36.

437 Hunter, J. R. and Macewicz, B. J. 1985a. Measurement of spawning frequency in  
 438 multiple spawning fishes. *In: An egg production method for estimating spawning*  
 439 *biomass of pelagic fish: application to the northern anchovy, Engraulius mordax.* pp. 79



440 – 94. Edited by R. Lasker. U.S. Department of Commerce, NOAA Technical Report  
 441 NMFS 36.

442 Hunter, J. R. and Macewicz, B. J. 1985b. Rates of atresia in the ovary of captive and  
 443 wild northern anchovy, *Engraulius mordax*. Fishery Bulletin, 83: 119-136.

444 Jennings, S.; Kaiser, M. J.; Reynolds, J. D. 2001. Fishing effects on populations and  
 445 communities. pp. 239 – 257. *In*: Marine Fisheries Ecology. Blackwell Science, London,  
 446 417pp.

447 Jons, G. D. and Miranda, L. E. 1997. Ovarian weight as an index of fecundity, maturity  
 448 and spawning periodicity. Journal of Fish Biology, 50: 150-156.

449 Maugé, L. A. 1990 Pomacanthidae. p. 841. *In*: Ed by Quero, J.C., Hureau, J.C., Karrer,  
 450 C., Post, A. and Saldanha, L. Check-list of the fishes of the eastern tropical Atlantic  
 451 (CLOFETA). JNICT, Lisbon, SEI, Paris; and UNESCO, Paris. Vol. 2.

452 McBride, R. S. and Thurman, P. E. 2003. Reproductive biology of *Hemiramphus*  
 453 *brasiliensis* and *H. balao* (Hemiramphidae): maturation, spawning frequency, and  
 454 fecundity. Biological Bulletin, 204: 57-67.

455 Menezes, N. A., Backup, P. A., Figueiredo, J. L. and Moura, R. L. 2003. Catálogo das  
 456 espécies de peixes marinhos do Brasil. Museu de Zoologia USP, São Paulo.

457 Michael, S. W. 2004. Family Pomacanthidae/Angelfishes. pp. 296 – 323 *In*:  
 458 Angelfishes & Butterflyfishes. T.F.H. Publications, New Jersey, 344pp.

459 Monteiro Neto, C., Cunha, F. E. A., Nottingham, M. C, Araújo, M. E, Rosa, I. L. and  
 460 Barros, G. M. L. 2003. Analysis of the marine ornamental fish trade at Ceará State,  
 461 northeast Brazil. Biodiversity and Conservation, 12: 1287 – 1295.

462 Moyer, J. T. and Nazakomo, A. 1978. Population structure, reproductive behavior, and  
 463 protogynous hermaphroditism in the angelfish *Centropyge interruptus* at Miyake-jimi,  
 464 Japan. Japan Journal of Ichthyology, 25: 25-39.

465 Moyer, J. T.; Thresher, R. E. and Colin, P. L., 1983. Courtship, spawning and inferred  
 466 social organization of American angelfishes (Genera *Pomacanthus*, *Holacanthus* and  
 467 *Centropyge*; Pomacanthidae). Environmental Biology of Fishes, 9: 25-39.

468 Munro, J.L., Gaut, V.C., Thompson, R. and Reeson, P.H.. 1973. The spawning seasons  
 469 of Caribbean reef fishes. Journal of Fish Biology, 5: 69 - 84.

470 Murphy, M. D. and Taylor, R. G. 1990. Reproduction, growth and mortality of red  
 471 drum *Sciaenops ocellatus* in Florida waters. Fishery bulletin, 88: 531-542.

472 Nottingham, M. C.; Cunha, F. E. and Monteiro Neto, C. 2000. Captura de peixes  
 473 ornamentais marinhos no Ceará. Arquivos Ciências do Mar, 33: 113 – 118.

474 Nottingham, M.C.; Silva, J.R.F. and Araújo, M.E. 2003. Morphology and histology of  
 475 the testicles of Queen Angelfish *Holacanthus ciliaris* (Perciformes: Pomacanthidae).  
 476 Arquivos Ciências do Mar, 36: 89-94.

477 Peña, N.; Alheit, I.; Nakama, M. E. 1996. Fecundidad parcial de la caballa del Perú  
 478 (*Scomber japonicus peruanus*). Boletín del Instituto del Mar de Perú, 10: 93-104.

479 Randall, J. E. 1967. Food habits of reef fishes of the West Indies. Studies in Tropical  
 480 Oceanography Miami, 5: 665-847.

481 Ross, R.M. 1990. The evolution of sex-change mechanisms in fishes. Environmental  
 482 Biology of Fishes, 29: 81–93.

483 Sadovy, Y. 1996. Reproduction in reef fishery species. pp. 15 – 60. *In*: Reef fisheries  
 484 Edited by N. V. C. Polunin and C. M. Roberts. Chapman and Hall, London, 496pp.

485 Sadovy, Y., Shapiro, D. Y. 1987. Criteria for the diagnosis of hermaphroditism in  
 486 fishes. Copeia, 1987: 136-156

487 Sakai, Y. 1986. Fecundity of female angelfish, *Centropyge ferrugatus*, independent of  
 488 body size: field collection of spawned eggs. Ichthyological Research, 43: 186 - 189.

489 Sakai, Y. and Kohda, M. 1997. Harem structure of the protogynous angelfish,  
 490 *Centropyge ferrugatus* (Pomacanthidae). Environmental Biology of Fishes, 49: 333 -  
 491 339.

492 Thresher, R. E. 1980. Angelfish. pp. 37 – 46. *In*: Reef fish: behavior and ecology on the  
 493 reef and in the aquarium. The Palmetto Publishing Company, St. Petresburg, 171pp.

494 Thresher, R. E. 1982. Courtship and spawning in the emperor angelfish *Pomacanthus*  
 495 *imperator*, with comments on reproduction in other pomacanthid fishes. Marine  
 496 Biology, 70: 149-156.

497 Thresher, R. E. 1984. Angelfishes. pp. 244 – 261. *In*: Reproduction in reef fishes. TFH  
 498 Publications, New Jersey, 398pp.

499 Vazzoler, A. E. A. M. 1996. Fecundidade: Parte prática. pp. 142 – 151. *In*: Biologia da  
 500 reprodução de peixes teleósteos: Teoria e prática. EDUEM, Maringá, 169 pp.

501 Warner, R. R. 1982. Mating systems, sex change and sexual demography in the rainbow  
 502 wrasse, *Thalassoma lucasanum*. Copeia, 1982: 653-661.

503 West, G. 1990. Methods for assessing ovarian development in fishes: a review.  
 504 Australian Journal of Marine and Freshwater Research, 41: 199-222.

505 Wootton, R. J. 1979. Energy costs of egg production and environmental determinants  
 506 of fecundity in teleost fishes. pp. 133-159. *In*: Fish phenology: Anabolic adaptiveness in  
 507 teleosts. Edited by P. J. Miller. Academic Press, London, 449pp.

508 Yamaguchi, A.; Todoroki, T. and Kume, G. 2006. Reproductive cycle, sexual maturity  
 509 and diel-reproductive periodicity of white croaker, *Pennahia argentata* (Sciaenidae), in  
 510 Ariake Sound, Japan. Fisheries Research, 82: 95-100.

511

Capítulo 2 - **Idade e crescimento do Paru preto *Pomacanthus paru* (Teleostei: Pomacanthidae) no nordeste do Brasil.**

Chapter 2\* - **Age and growth of the French angelfish *Pomacanthus paru* (Teleostei: Pomacanthidae) in Northeastern Brazil.**

\* Capítulo em formato de artigo e padronizado de acordo com as normas da revista *Journal of Fish Biology*.

**Age and growth of the French angelfish *Pomacanthus paru* (Teleostei: Pomacanthidae) in Northeastern Brazil.**

Feitosa, C. V.<sup>a, b</sup>, Ferreira, B. P.<sup>a</sup>, Araújo, M. E.<sup>a</sup>

<sup>a</sup> - Departamento de Oceanografia, Centro de Tecnologia e Geociências, Universidade Federal de Pernambuco, 50670-901, Recife, Pernambuco, Brazil.

<sup>b</sup> - Corresponding author.

**Abstract**

The aim of the present study was to describe age and growth of the French angelfish *Pomacanthus paru*. Age was determined by sectioned otoliths alone. All treated otoliths with tetracycline were one year of age and revealed a clear fluorescent mark when observed under UV light. Based on age determination studies, increments observed in sagittae otoliths of tropical fish may be consistently interpreted as having annual periodicity. Otolith weight increased exponentially with standard length and linearly with age. This indicates that otolith growth continues with age and is independent of size. Age of the fish in the sample ranged from 1 to 27 years, with males exhibiting lower K and greater asymptotic length than females. *P. paru* attains maximal size slowly and has a long lifespan. Most linear growth is achieved by approximately 52% of lifespan. Besides being an important ornamental species, *P. paru* has been commonly caught as bycatch in trap fisheries for decades. These growth parameters should be used with the purpose of managing fisheries targeting this species before more meaningful limits can be imposed. In the case of aquarium trade management, it can be suggested that conservationist issues should be based on capture per area and the establishment of protected areas.

**Key words:** *Pomacanthus paru*, growth parameters, validation, aquarium trade.

Email: carol\_feitosa@hotmail.com. Tel.: + 55 – 81 – 21267223; Fax: + 55 – 81 – 32278227

## Introduction

One of the most commonly employed methods for age determination is the interpretation of rhythmic deposition in calcified tissues used as time markers (Casselman, 1987). Considered the most appropriate structure for age determination (Six & Horton, 1977), otoliths are calcium carbonate concretions in aragonite crystal form (Degens *et al.*, 1969) and are found in the membranous labyrinth of the inner ear of teleost fish (Lowestein, 1971). The use of otoliths for age determination was first developed for temperate fish populations (Brothers, 1980), which have seasonally controlled periods of growth and spawning. Due to the lack of seasonality in tropical waters, the use of the otolith method for age studies on tropical fish began as a complex process. However, in the late 1980s, Longhurst & Pauly (1987) stated that the growth of tropical fish follows expected seasonal patterns, which can be observed using length frequency data or the analysis of seasonal bands in otoliths.

Despite the importance of studies on age, growth, mortality and longevity, limited information is available on the population dynamics of tropical fish (Radtke, 1987). Age and growth studies are crucial to understanding the life history traits of fish species, including population structure, changes in population growth due to environmental variations and recruitment success (Dee & Radtke, 1989; Zekeria *et al.*, 2000). Such studies are required to explain demographic characteristics that are unique to tropical fish populations and determine environmental influences on life history events (Radtke, 1987). Furthermore, growth and ageing studies on species undergoing fishery pressure provides indispensable information for stock assessments and fishery management (Aliaume *et al.*, 2000).

Information on growth of coral reef fish is limited to a few families, such as acanthurids, haemulids, lutjanids, scarids, serranids, sparids (Choat & Axe, 1996; Choat

51 *et al.*, 1996). For the family Pomacanthidae, estimates from age-at-length data are only  
52 available for one species – *Pomacanthus imperator* (Bloch, 1787) (Chung & Woo,  
53 1999). Moreover, the method employed for ageing this species was the reading of  
54 scales, which is prone to error (Williams & Bedford, 1974; Erickson, 1983).

55 Fish from the family Pomacanthidae are among the most colorful and  
56 recognizable groups of reef fish (Thresher, 1982). This family comprises 88 species,  
57 consisting of eight genera and represents one of the most noticeable components in reef  
58 fish assemblages, with representatives in all tropical seas (Allen *et al.*, 1998). Although  
59 angelfish are analogous in overall appearance, there is a diverse range of ecological  
60 traits within the family, such as differences in body size, color pattern, reproductive  
61 system and diet (Bellwood *et al.*, 2004).

62 The French angelfish *Pomacanthus paru* (Bloch, 1787) occurs from Florida, the  
63 Bahamas and the northern Gulf of Mexico down to Brazil (Menezes *et al.*, 2003). It is  
64 also recorded for St. Peter and St. Paul's Rocks (Feitoza *et al.*, 2003) and Ascension  
65 Island (Maugé, 1990) in the Central Atlantic and the Gulf of Guinea. Maximal length  
66 reported for this species is 41 cm (Cervigón, 1993). It is distributed in waters from 5 to  
67 100 m, with juveniles and subadults being more frequent in shallow waters. This species  
68 is an omnivore and sponges are its main food item (Randall, 1967). It is a gonochoric  
69 species (Feitosa *et al.*, unpublished data) and it seems that adults generally form stable  
70 pairs, although in areas where the fish is more abundant, it is known to form small  
71 harems (Thresher, 1984; Michael, 2004).

72 Pomacanthid species are among the most harvested for the Brazilian aquarium  
73 trade and *P. paru* is one of the most exported species (Nottingham *et al.*, 2000; Araújo  
74 & Albuquerque-Filho, 2005; Monteiro-Neto *et al.*, 2003). Along with the fishing  
75 pressure by the Brazilian aquarium trade, this species is also caught as bycatch (Feitosa

76 *et al.*, 2008). Management measures for this species are restricted to a Federal Act  
77 (56/04) established in 2004 with the purpose of managing the aquarium trade through  
78 the establishment of export quotas.

79 In the present study, age and growth of *P. paru* were studied through the  
80 validation and examination of annual marks in sagittae otoliths.

## 81 **Materials and methods**

### 82 **Sampling design**

83 The fish were sampled from commercial boats from the Itamaracá Island Fishing  
84 Colony, located on Pilar beach (7°45'17.80''S – 34°49'26.46''W) (Itamaracá, state of  
85 Pernambuco, Brazil – Fig. 1). These fish were caught as bycatch during trap fishery  
86 activities targeting the spotted goatfish, *Pseudupeneus maculatus* (Bloch, 1793), and  
87 parrotfish, *Sparisoma* spp. In this fishery, traps are deployed unbaited next to reefs and  
88 hauled 6 hours later. Occasionally, depending on factors such as wind and currents,  
89 some traps are left for longer periods and only hauled after 48 hours. Sampling was  
90 performed weekly from March 2006 to February 2007. As individuals smaller than 15  
91 cm are not captured in this type of fishery, such individuals were bought from fish  
92 collectors between September 2006 and February 2007.





Figure 1: Location of sampled site.

#### Otolith ageing

Fish were measured for total length [TL (cm)] and standard length [SL (cm)], weighed (g) and sexed. The largest otoliths (*sagittae*) were removed for analysis and stored dry. Whenever possible, the left otolith was used for both whole and sectioned views. Rings were counted for whole otoliths by immersing the otoliths concave side up on a watch glass with a blackened bottom containing 100% ethanol and viewed through a binocular dissecting microscope. Opaque bands were counted on the distal surface from the nucleus to the dorsal side. Prior to sectioning, all pairs were measured and weighed to the nearest 0.01 g.

The otoliths were embedded in an epoxy resin prior to reading. Each otolith was sectioned transversely (approximately 0.3 mm thick) with a low-speed saw. Sections were mounted on microscope slides with Histomount mounting media and examined for opaque bands under a dissecting microscope using reflected light.

108 To assess the precision of the readings, two independent readers counted age  
109 marks on whole and sectioned otoliths with no knowledge of fish size. A second reader  
110 tested the precision of the first reader for a subsample of 50 otoliths sections (21%).  
111 With these results, the index of average percentage error (IAPE) between readers was  
112 calculated (Beamish & Fournier, 1981). If the readers diverged by more than 10% for a  
113 given otolith section, the readings were repeated. The data were included in the analysis  
114 only when the IAPE was below 10%.

115 A validation experiment was performed to determine the accuracy of the otolith  
116 readings. For this test 6 individuals were used. As individuals sampled for this test were  
117 smaller than 6 cm, the specimens were marked by immersion in a solution of  
118 tetracycline and seawater (Beamish & McFarlane, 1987), with an initial concentration  
119 of 100 mg l<sup>-1</sup>. Each subsequent four hours, the same concentration (100 mg l<sup>-1</sup>/h) was  
120 added to the solution until reaching the maximal concentration of 500 mg l<sup>-1</sup>. The fish  
121 were kept in this solution for 12 hours (Hernaman *et al.*, 2000). After this procedure,  
122 they remained in aquarium for 1 year, with a mean salinity and temperature of 30 ‰  
123 and 28°C, respectively.

124 The treated otoliths were sectioned in the same way as the unlabeled ones and  
125 analyzed under a microscope with an ultraviolet light source to detect the fluorescent  
126 mark. An external fiber-optic light was coupled to the microscope in order to observe  
127 the opaque bands and fluorescent marks simultaneously (Ferreira & Russ, 1992).

128 Growth was investigated by fitting the von Bertalanffy (1938) growth function  
129 to size-at-age data using the nonlinear optimization method on the Kaleidagraph 4.0  
130 software program. The model was fit for each sex separately. The growth function is  
131 defined as follows:

132 
$$L_t = L_{\infty} (1 - e^{-k(t - t_0)})$$

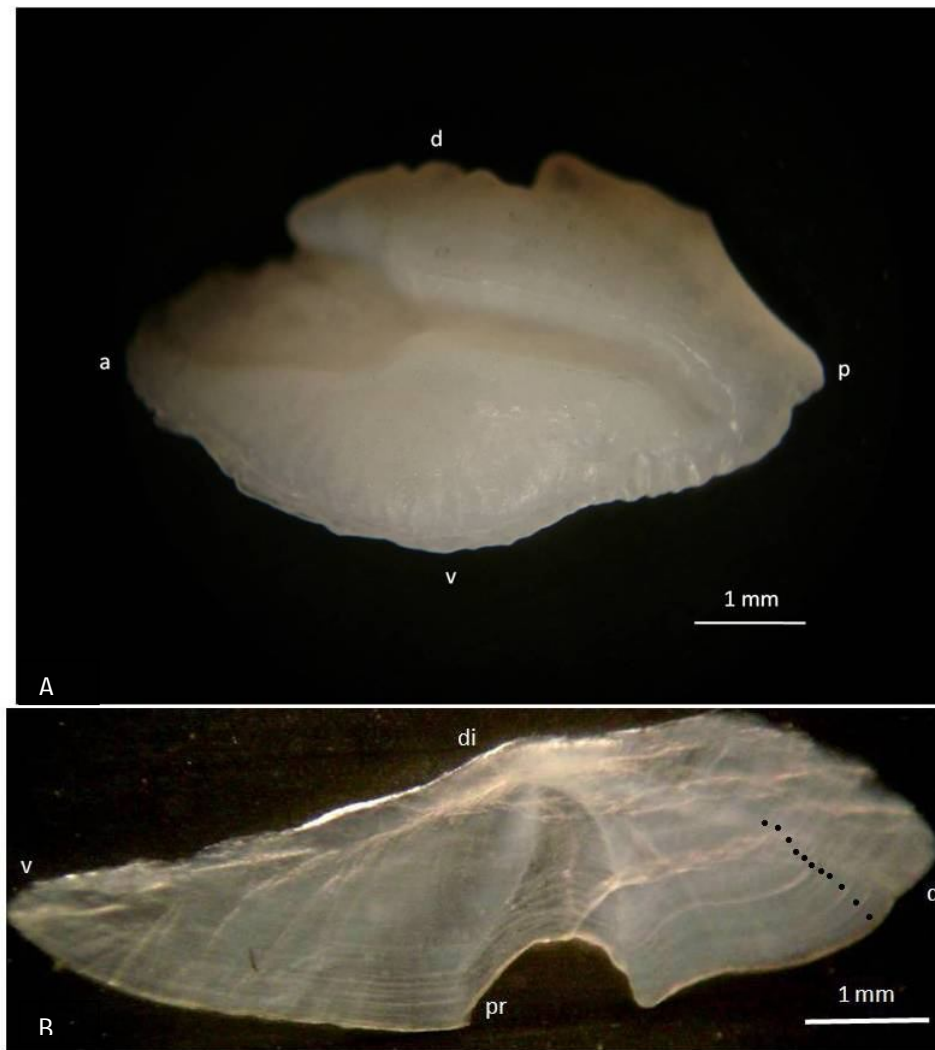
133 in which  $L_t$  is length at time  $t$ ,  $L_\infty$  is asymptotic length,  $k$  is the growth coefficient and  
134  $t_0$  is the hypothetical time at which length is equal zero. The rate of increase in size with  
135 age between sexes was compared using the Mann-Whitney test ( $\alpha = 0.05$ ). The paired  $t$ -  
136 test was used to compare the weight of left and right sagittae otolith ( $\alpha = 0.05$ ). The  
137 relationship between otolith weight, standard length and age was determined through  
138 simple linear regression analysis.

139 Parameters of the length-weight relationship were obtained by fitting the  
140 function  $W = aL_s^b$  to length and weight data, in which  $W$  is the total weight,  $a$  is the  
141 constant (determined empirically),  $L_s$  is the standard length and  $b$  is close to 3 for  
142 species with isometric growth.

143

## 144 **Results**

145 In French angelfish, the sagittae otolith is the largest and oval shaped, with a  
146 pointed rostrum. It is laterally compressed, with a heterosucoid sulcus acusticus (Fig. 2).  
147 A pattern of opaque and translucent bands can be recognized and counted on whole and  
148 sectioned otoliths. Nevertheless, counting opaque bands on whole otoliths is a complex  
149 task due to the overlapping of bands at the margins in larger fish and regular divergence  
150 in counts between different otolith regions, mainly in the dorsal area. Due to these  
151 complexities, the age of *P. paru* was determined only on sectioned otoliths (Fig. 2).



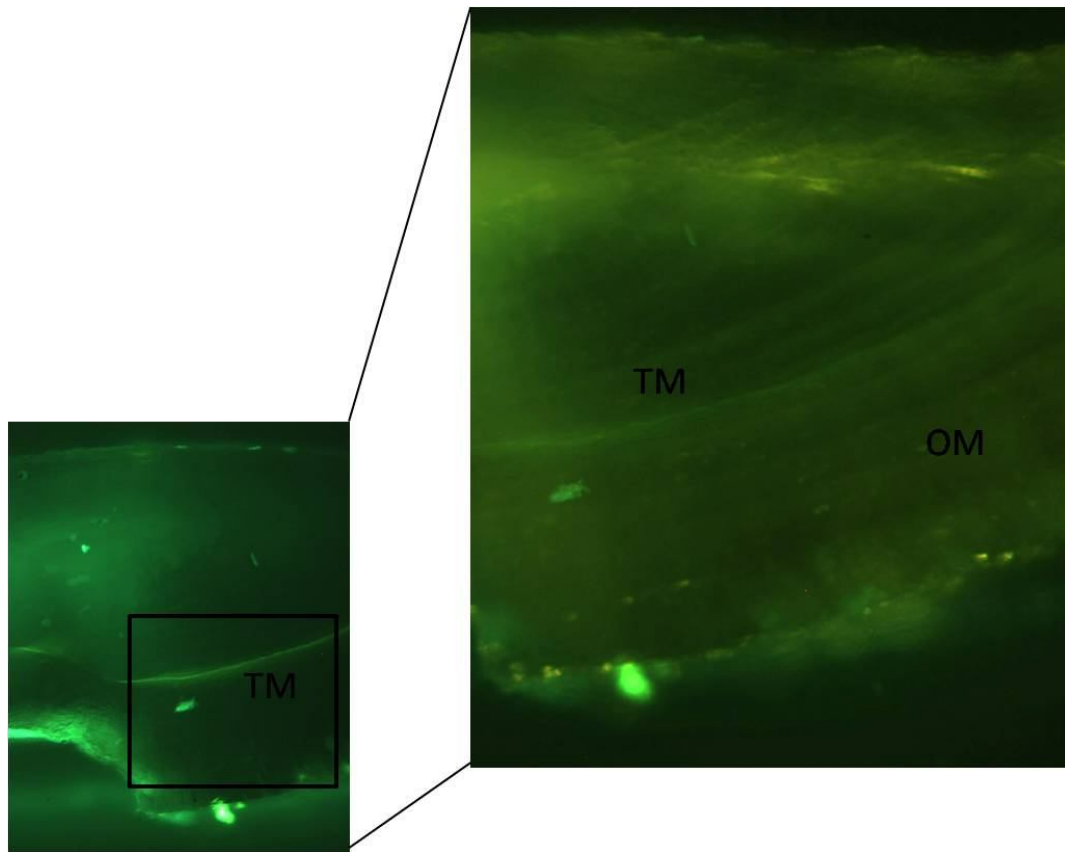
152

153

154 Fig. 2: Whole (A) and sectioned (B) *sagittae* otoliths of a 8 and a 11 years old  
 155 *Pomacanthus paru* (a = anterior, p = posterior, d = dorsal, v = ventral, di = distal, pr =  
 156 proximal).

157

158 The sections of all treated otoliths revealed an opaque band between a clear  
 159 fluorescent mark and the edge when observed under an external light source and UV  
 160 light (Fig 3). Observations from all of the sections support the hypothesis that the  
 161 annulus is formed once a year. All the treated French angelfish were one year old.



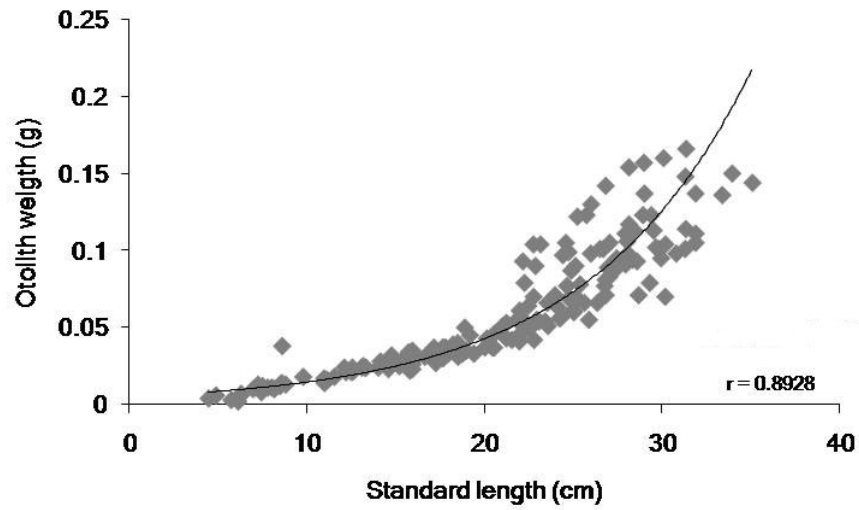
163

164 Fig. 3: Sectioned otolith treated with tetracycline observed under UV light (A) and  
 165 under an external light source and UV light (B). TM – tetracycline mark, OM – opaque  
 166 mark.

167

168 Otoliths ranged in weight from 0.002 (7.81 cm, 16 g, 4-year-old fish) to 0.169 g  
 169 (38.64 cm, 1.396 g, 26-year-old fish). There was no significant difference between the  
 170 weight of the left and right otolith (paired t-test,  $n = 153$ ,  $P > 0.05$ ). The exponential  
 171 relationship between otolith weight and standard length follows the equation  $OW =$   
 172  $0.0049e^{0.1082 SL}$  ( $r = 0.89$ ,  $n = 184$ ), in which OW is otolith weight (g) and SL is standard  
 173 length (cm) (Fig. 4). Otolith weight increased linearly with age and was described by  
 174 the equation  $OW = 0.0053 SL + 0.0036$  ( $r = 0.64$ ,  $n = 167$ ) (Fig. 5). The relationship of  
 175 total weight to standard length is  $W = 0.1192SL^{2.794}$  ( $r = 0.98$ ,  $n = 236$ ) (Fig. 6).

176

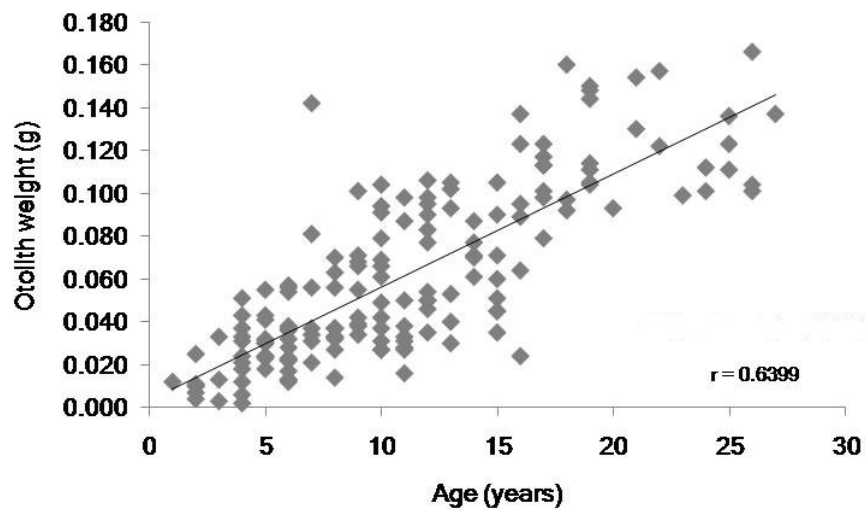


177

178 Figure 4: Relation between *sagittae* otolith weight and standard length for *Pomacanthus*

179 *paru*. SL = standard length (cm), OW = otolith weight

180

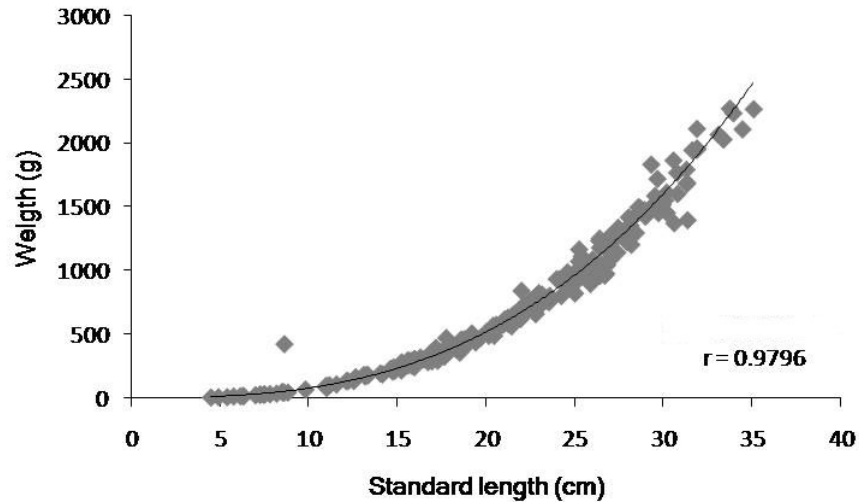


181

182 Figure 5: Relation between otolith weight and age for *Pomacanthus paru*. A = age

183 years, OW = otolith weight.

184



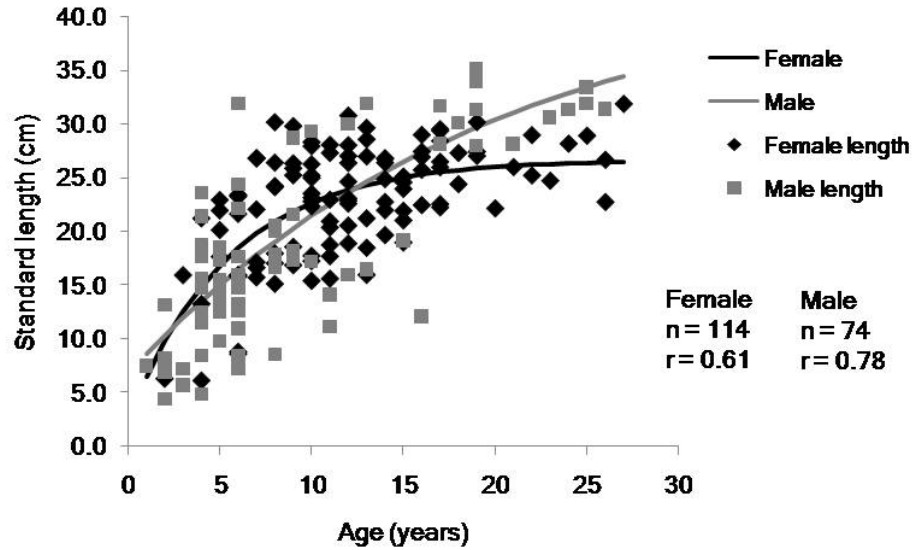
185

186 Figure 6: Relation of weight to length for *Pomacanthus paru*. SL = standard length  
187 (cm), W = total weight (g).

188

189 Age of the specimens in the sample ranged from 1 to 27 years (Fig. 7). Females  
190 age range was 2 to 27 years and males was 1 to 26. Statistically differences were  
191 recorded for size at age between sexes only for the age ranging from 16 to 20 ( $n = 27$ )  
192 and 21 to 27 years ( $n = 15$ ) (Mann-Whitney test,  $P < 0.05$ ). A total of 2.30% of the  
193 otolith sections were unreadable and were not included in the analysis. The average  
194 percentage error between readers was low (9.80%). French angelfish are long-lived (to  
195  $> 27$  years) and attain maximal size slowly, especially males. The plots were fitted best  
196 by the von Bertalanffy growth function for females:  $L_t = 26.66 (1 - e^{-0.18 (t + 0.56)})$  and  
197 males  $L_t = 44.11 (1 - e^{-0.05 (t + 3.34)})$ .

198



199

200 Figure 7: Von Bertalanffy growth curve fitted to length at age data of female and male  
201 of *Pomacanthus paru*.

202

## 203 Discussion

204 Despite being widely recognized as important coral reef inhabitants and very  
205 significant in marine aquarium trade, there are few studies on age and growth of  
206 representatives of the family Pomacanthidae (Chung & Woo, 1999). This may be  
207 attributed to the difficulty in sampling individuals in all length classes.

208 In the present study, sagittae otoliths – which are commonly the largest (Panella,  
209 1971) – from French angelfish exhibited a regular pattern of annual opaque and  
210 translucent bands, the readings of which were highly repeatable, with an average error  
211 of 9.80%. Based on age determination studies, increments observed in the sectioned  
212 sagittae otoliths of tropical fish may be reliably interpreted as having annual periodicity  
213 (Choat & Robertson, 2002).

214 The relationship between otolith weight and age was linear. This suggests that  
215 otolith growth is continuous for *P. paru*, while fish size (standard length) is asymptotic



216 at intermediate ages, indicating that this relationship is not continuous. Thus, otolith  
217 growth is continuous with age and is independent of fish size. It seems that this is a  
218 general rule of fish growth, as this pattern has been observed in several other species  
219 (Dee & Radtke, 1989; Hart & Russ, 1996; Schwamborn & Ferreira, 2000; Amezcua *et*  
220 *al.*, 2006; Araújo & Martins, 2006; Zekeria *et al.*, 2006). Bands were deposited annually  
221 and the weight of otoliths increased throughout the lifespan, thereby fulfilling the  
222 fundamental criteria required for ageing (Fowler & Doherty, 1992).

223         Growth data for the French angelfish fitted the von Bertallanfy model well.  
224 Longevity was up to 27 years and maximal asymptotic standard length was 44.11 cm,  
225 with males exhibiting lower K and greater  $L_{\infty}$ . However, the longevity was nearly the  
226 same between sexes. Females reach asymptotic length nearly at age 15 and in this age  
227 males continue to grow attaining greater sizes at the subsequent ages. The samples used  
228 in the present study came from trap fisheries and, consequently, were related to the  
229 selectivity of this fishing gear. For the angelfish *Holacanthus passer* sex ratio differed  
230 significantly over the size range, with females prevailing at small lengths and males at  
231 larger lengths (Arellano-Martínez *et al.* 1999). A similar pattern was observed in the  
232 present study, in which males were larger than females. It is possible that when females  
233 attain age at sexual maturity, they strongly reduce their growth.

234         Reinforcing the differences in growth patterns between sexes, Feitosa *et al.*  
235 (unpublished data - chapter 1) observed size dimorphism in *Pomacanthus paru*, with  
236 males attaining larger sizes. Studying *Pomacanthus paru* in the Caribbean (St Croix, US  
237 Virgin Islands), Aiken (1983) states that males tend to mature later than females. This  
238 difference in size at maturity was also recorded by Feitosa *et al.* (unpublished data -  
239 chapter 1). These authors affirm that individuals fit for reproduction appeared at 20 to  
240 25 cm for females and 25 to 30 cm TL for males, with  $L_{50}$  different between sexes,

241 being maturity reached at 30cm for females and 35 cm for males. These findings  
242 corroborate with the present results, in which males presented lower K and had greater  
243 asymptotic length when compared to females.

244         Using size distributions of trap catches of the Gray angelfish, *Pomacanthus*  
245 *arcuatus* (Linnaeus, 1758) based on moderate samples from the unexploited sites of  
246 Pedro Bank and the Porto Royal reefs, Aiken (1983) determined that 60 cm TL is the  
247 asymptotic length. Although *P. paru* and *P. arcuatus* are sympatric species, maximal  
248 length ( $L_{\infty}$ ) was divergent, even considering that the  $L_{\infty}$  established for *P. arcuatus*  
249 was based on total length. Aiken (1983) also examined scales from *P. paru* and found  
250 that rings or marks were present, but no interpretable pattern was discernible. The age  
251 and growth study of *Pomacanthus imperator* demonstrated that the  $L_{\infty}$ , based on  
252 standard length, for this specie is 41.19 cm (Chung & Woo, 1999). This value is very  
253 large when compared with the  $L_{\infty}$  recorded for *Pomacanthus paru* in this research.

254         Disparity between sets of growth parameters may be attributed to factors such as  
255 sample size, sample composition and the method employed to estimate growth  
256 parameters as well as divergences in population structure of different locations (Choat  
257 & Robertson, 2002; Araújo & Martins, 2006; Grandcourt *et al.*, 2006). Some  
258 hypotheses may explain these differences, such as the method used by Aiken (1983) and  
259 Chung & Woo (1999), which was length-frequency distribution in the former and the  
260 reading of scales in the latter. Another explanation is that *P. paru* occurs in the Atlantic,  
261 whereas *P. imperator* occurs in the Central and Indo-West Pacific (Allen *et al.*, 1998). It  
262 is likely that differences in environmental (such as different water temperatures) and  
263 genetic characteristics can explain the contrast in growth parameters (Grandcourt *et al.*,  
264 2006). Due to the issues mentioned above, Laurec & LeGuen (1981) state that growth

parameters are merely simple mathematical supports and have no value as biological characters *a priori*.

*P. paru* attains its maximal size slowly and has a long lifespan, with most linear growth achieved by approximately 52% of the lifespan. This growth pattern is shared by other reef fish (e.g. serranids, haemulids and lutjanids), which have K values ranging from 0.10 to 0.22 and maximal ages ranging from 9 to 16 years (Matheson *et al.*, 1986; Munro & Polunin, 1997). However, *P. paru* lives up to 10 years more and, in the case of males, the K value was smaller than that cited in the literature (Matheson *et al.*, *op. cit*; Munro & Polunin, *op. cit*). The low K value and long life among demersal fish suggests that these characteristics are associated to evolutionary success in the reef environment (Huntsman, 1981). According to Matheson *et al.* (1986), long life and relatively large body size maximize gamete and zygote production and allow reef fish to overcome the apparently low possibility of the pelagic offspring encountering a favorable habitat. Conversely, Cuban coastal reef fish, such as haemulids and lutjanids, have a short lifespan and a relatively high growth rate, with species reaching between 35 and 50% of maximal size in the first year of life, according to Claro & García-Arteaga (2001). The authors suggest that these characteristics are related to adaptive mechanisms for avoiding predation on juveniles. Studying the coral trout *Plectropomus leopardus* (Lacepède, 1802) from the Great Barrier Reef, Australia, Ferreira & Russ (1994) recorded a different survival mode in growth. This serranid is relatively short-lived. The growth rate becomes asymptotic from age six onwards and a substantial proportion of total growth is attained early in its life. This rapid growth is favored due to the availability of potential prey and avoidance of predators with the increase in body size. A similar pattern has been observed among scarids and acanthurids from the Great

289 Barrier Reef, for which growth patterns are highly deterministic and most linear growth  
290 is reached in the initial 10 to 20% of lifespan (Gust *et al.*, 2002).

291 *P. paru* and other pomacanthids are heavily exploited by the aquarium trade  
292 (Moenick, 1991; Wood, 2001a; Monteiro-Neto *et al.*, 2003). The family Pomacanthidae  
293 alone comprises 46% of exports by value (Pyle, 1993) and more than 21 species are  
294 harvested (Wood, 2001a). The French angelfish is a very important species in the  
295 Brazilian aquarium trade and is the second most exported species (Nottingham *et al.*,  
296 2000; Monteiro-Neto *et al.*, 2003; IBAMA, 2007). In the USA (state of Florida), this  
297 species is managed through size regulations (Wood, 2001b). In Brazil, a Federal Act  
298 (56/04) was established in 2004 with the purpose of managing fishery activities  
299 targeting this species through the establishment of export quotas. The extended growth  
300 and late sexual maturity ( $\approx 23$  cm TL (female) - Feitosa *et al.*, unpublished data;  $\approx 11$   
301 years old; 25 cm TL (male) - Feitosa *et al.*, unpublished data;  $\approx 15$  years old) of *P. paru*  
302 and other pomacanthids are angelfish characteristics that will render them vulnerable to  
303 extinction (Roberts & Hawkins, 1999). As large-body specimens require a long time to  
304 grow, the removal of these specimens through predation or human activities is least  
305 likely to be recovered quickly (Chung & Woo, 1999). Individuals of *P. paru* and other  
306 species collected for the aquarium trade are generally under 10 cm in standard length  
307 and rarely reach as much as 20 cm, which represents juveniles and young adults that  
308 have likely not reached sexual maturity (Wood, 2001b).

309 Chung & Woo (1999) affirm that it was fortunate that the commercial  
310 exploitation of *P. imperator* has only focused on the aquarium trade. As individuals  
311 longer than 25 cm (SL) are rarely exploited and specimens up to 29.6 cm (SL) are still  
312 reproductively active, these fish constitute an important breeding stock if younger  
313 individuals are overexploited. However, this is not the case with *P. paru*. Besides the

314 impact from the ornamental fish trade, this species has been commonly caught as  
315 bycatch in trap fisheries for decades (Aiken, 1983; Feitosa *et al.*, 2008). In this kind of  
316 fishery, more than 70% of individuals caught are fit for reproduction (Feitosa *et al.*,  
317 2008). Thus, it is possible that this species suffers fishing pressure at all length ranges  
318 and that this impact may threaten recruitment to the spawning stock. Besides, *P. paru*  
319 presents ecological importance. This species feeds on sponges and zoanthids (*Palythoa*)  
320 which compete with corals making their growth difficult.

321 In conclusion, the results demonstrate that it is possible to obtain estimates of  
322 growth for tropical species based on conventional age determination methods. These  
323 estimates should be used with the purpose of managing the fishery activity targeting this  
324 species. There are some management measures in place for the species throughout its  
325 range of distribution, which include size regulation (USA) and species-based  
326 exportation quotas (Brazil), all directed to the aquarium fish trade. While these  
327 regulations can clearly play an important role in helping to conserve stocks and prevent  
328 overexploitation, management measures should also include measures directed to other  
329 fisheries. In Brazil the species is commonly captured as bycatch in trap fisheries, an  
330 unregulated multi-specific fishery. Ornamental fish species are an important part of the  
331 bycatch (Feitosa *et al.*, 2008). Those species are caught and usually returned to the sea -  
332 many still alive, according to the fishermen. Survival of these individuals, however, is  
333 doubtful, as the traps are hauled quickly to surface and problems related to  
334 decompression have often been reported.

335 Monitoring and management of this fisheries, a growing activity with catches  
336 exported to the international market, is mandatory in order to assess the impact of this  
337 kind of fishery on both target and non-target species. This paper contributes to the  
338 knowledge on life history traits of *P. paru*, such as growth rates and age at sexual

339 maturity. Both are minimal requirements for effective fishery management regarding a  
340 given species. In the case of aquarium trade management, where most commercialized  
341 fish are juvenile and under than 10 cm in standard length, it can be suggested that  
342 conservationist issues should be based on quantitative data (capture) per area and the  
343 establishment of protected areas.

344

#### 345 **Acknowledgements**

346 This study was sponsored by the Brazilian Scientific Council - Conselho  
347 Nacional de Pesquisa e Desenvolvimento (CNPq) and Pernambuco State Scientific  
348 Council - Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco  
349 (FACEPE).

#### 350 **References**

- 351 Aiken, K. (1983). The biology, ecology and bionomics of the butterfly and angelfishes,  
352 Chaetodontidae. In Caribbean coral reef fisheries resource. pp. 155 – 165. (J. L. Munro  
353 ed.) Iclarm studies and reviews 7: Manila, Philippines.
- 354 Aliaume, C., Zerbi, A., Joyeux, J. C., Miller, J. M. (2000). Growth of juvenile  
355 *Centropomus undecimalis* in a tropical island. *Environmental Biology of Fishes* **59**, 299  
356 – 308.
- 357 Allen, G. R., Steene, R., Allen, M. (1998). Introducing angelfish. In A guide to  
358 angelfishes & butterflyfishes. pp. 8 – 12. Odyssey Publishing: USA/ Tropical Reef  
359 Research: Australia.
- 360 Allen, G. R., Steene, R., Allen, M. (1998). The Indo-West Pacific region. In A guide to  
361 angelfishes & butterflyfishes. pp. 86. Odyssey Publishing: USA/ Tropical Reef  
362 Research: Australia.

363 Araújo, J. N., Martins, A. S. (2006). Age and growth of coney (*Cephalopholis fulva*),  
 364 from the central coast of Brazil. *Journal of the Marine Biological Association of the*  
 365 *United Kingdom* **86**, 187 – 191.

366 Araújo, M. E. & Albuquerque Filho, A. C. (2005). Biologia das principais espécies de  
 367 peixes ornamentais marinhos do Brasil: uma revisão bibliográfica e documental.  
 368 *Boletim Técnico Científico do CEPENE* **13**, 109 – 154.

369 Arellano-Martínez, M., Ceballos-Vásquez, B. P., Garcia-Dominquez, F. and Galván-  
 370 Magaña, F. (1999). Reproductive biology of the King Angelfish *Holacanthus passer*  
 371 Valenciennes 1846 in the Gulf of California, Mexico. *Bulletin of Marine Science* **65**,  
 372 677 - 685.

373 Bannerot, S., Fox Jr., W. W. & Powers, J. E. (1987). Reproductive strategies and the  
 374 management of snappers and groupers in the Gulf of Mexico and Caribbean. In  
 375 *Tropical snappers and groupers: biology and fisheries management* (Polovina, J. J. &  
 376 Ralston, S eds.), pp. 561 – 603. Boulder: Westview Press.

377 Beamish, R. J. & Fournier, D. A. (1981). A method for comparing the precision of a set  
 378 of age determinations. *Canadian Journal of Aquatic Science* **38**, 982 – 983.

379 Beamish, R. J. & McFarlane, G. A. (1987). The forgotten requirements for age  
 380 validation in fisheries biology. *Transactions of the American Fisheries Society* **112**,  
 381 735-743.

382 Bellwood, D. R., van Herwerden, L. & Konow, N. (2004). Evolution and biogeography  
 383 of marine angelfishes (Pisces: Pomacanthidae). *Molecular Phylogenetics and Evolution*  
 384 **33**, 140-155.

385 Brothers, E. B. (1980). Age and growth studies in tropical fishes. In *Stock assessment*  
 386 *for tropical small-scale fisheries*, pp 119–136. University of Rhode Island: Kingston.

387 Casselman, J. M. (1987). Determination of Age and Growth. In The Biology of Fish  
388 Growth ( A.H. Weatherley and H. S. Gill Eds.), pp. pp. 209-242. London: Academic  
389 Press.

390 Cervigón, F. (1993). Pomacanthidae. pp. 339 – 346. In: Los Peces Marinos de  
391 Venezuela. Fundación Científica Los Roques, Caracas, 951pp.

392 Choat, J. H. & Axe, L. M. (1996). Growth and longevity in acanthurid fishes, an  
393 analysis of otolith increments. *Marine Ecology Progress Series* **134**, 15–26.

394 Choat, J. H., Axe, L. M. & Lou, D. C. (1996). Growth and longevity in acanthurid  
395 fishes of the family Scaridae. *Marine Ecology Progress Series* **145**, 33–41.

396 Choat, J. H. & Robertson, R. (2002). Age based studies. In *Coral reef fishes: dynamics*  
397 *and diversity in a complex ecosystem*. pp. 57-80. San Diego: Academic Press.

398 Chung, K. C., Woo, N.Y.S. (1999). Age and growth by scale analysis of pomacanthus  
399 imperator (Teleostei: Pomacanthidae) from Dongsha Islands, southern Chine.  
400 *Environmtal Biology of Fish* **55**, 399 – 412.

401 Claro, R. & García-Arteaga, J. P. (2001). Growth patterns of fishes of the Cuban shelf.  
402 In *Ecology of the marine fishes of Cuba* (R.Claro, K. C. Lindeman & L. R. Parenti,  
403 eds.), pp.149–178. Washington: Smithsonian Institution Press.

404 Dee, A. J. & Radtke, R. L. (1989). Age and growth of the bricke soldierfish, *Myripristis*  
405 *amaena* *Coral reefs* **8**, 79-85.

406 Degens, E. T., Deuser, W. G. & Haedrich, R. L. (1969). Molecular structure and  
407 composition of fish otoliths. *Marine Biology* **2**, 105-113.

408 Erickson, C. M. (1983). Age determination of Manitoban walleyes using otoliths, dorsal  
409 spines, and scales. *North American Journal of Fisheries Management* **3**, 176-181.



410 Feitosa, C. V., Ferreira, B. P. & Araújo, M. E. (2008). A rapid new method for assessing  
 411 sustainability of ornamental fish by-catch from coral reefs. *Marine and Freshwater Research*  
 412 **59**, 1092-1100.

413 Feitoza, B. M., Rocha, L. A., Luiz-Júnior, O. J., Floeter, S. R. & Gasparini, J. L. (2003).  
 414 Reef fishes of St. Paul's Rocks: new records and notes on biology and zoogeography.  
 415 *Aqua, Journal of Ichthyology and Aquatic Biology* **7**, 61– 82.

416 Ferreira, B. P., Russ, G. R. (1992). Age, growth and mortality of the inshore coral trout,  
 417 *Plectropomus maculatus* (Pisces: Serranidae) from the Central Great Barrier Reef,  
 418 Australia. *Australian Journal of Marine and Freshwater Research* **43**, 1301 – 1312.

419 Ferreira, B. P., Russ, G. R. (1994). Age validation and estimation of growth rate of the  
 420 coral trout, *Plectropomus leopardus*, (Lacépède 1802) from Lizard Island, Northern  
 421 Great Barrier Reef. *Fishery Bulletin* **92**, 46 – 57.

422 Fowler, A. J. & Doherty, P. J. (1992). Validation of annual growth increments in the  
 423 otoliths of two species of damselfish from southern Great Barrier Reef. *Australian*  
 424 *Journal of Marine and Freshwater Research* **43**, 1057-1068.

425 Grandcourt, E. M., Al Abdessalaam, T. Z. & Francis, F. (2006). Age, growth, mortality  
 426 and reproduction of the blackspot snapper, *Lutjanus fulviflamma* (Forsskal, 1775), in the  
 427 southern Arabian Gulf. *Fisheries Research* **78**, 203 – 210.

428 Gust, N., Choat, J. H., Ackerman, J. L. (2002). Demographic plasticity in tropical reef  
 429 fishes. *Marine Biology* **140**, 1039 – 1051.

430 Hart, A. M. & Russ, G. R. (1996). Response of herbivorous fish to crown of thorns  
 431 starfish *Acanthaster planci* outbreaks. III. Age, growth, mortality and maturity indices  
 432 of *Acanthurus nigrofusus*. *Marine Ecology Progress Series* **136**, 25 – 35.

433 Hernaman, V., Munday, P. L. & Schläppy, M. L. (2000). Validation of otolith growth-  
 434 increment periodicity in tropical gobies. *Marine Biology* **137**, 715 - 726.

435 Huntsman, G. R. (1981). Ecological considerations influencing the management of reef  
 436 fishes. In *Artificial reefs: Conference proceedings*. pp. 167 – 175. Sea Grant Report **41**.  
 437 IBAMA. (2007). [http://www.ibama.gov.br/recursos-pesqueiros/wp-](http://www.ibama.gov.br/recursos-pesqueiros/wp-content/files/quantidades_exportadas_2007_marinho.pdf)  
 438 [content/files/quantidades\\_exportadas\\_2007\\_marinho.pdf](http://www.ibama.gov.br/recursos-pesqueiros/wp-content/files/quantidades_exportadas_2007_marinho.pdf)

439 Laurec, A. & LeGuen, J. C. (1981). Dynamique des populations marines exploitées. In  
 440 *Rapports Scientifiques et Techniques*, 45. Centre National pour l'exploitation des  
 441 Océans.

442 Longhurst, A. R. & Pauly, D. (1987). Dynamics of Tropical Fish Populations. In  
 443 *Ecology of tropical oceans*. pp. 300 - 306. Orlando: Academic Press

444 Lowenstein, O. (1971). The labyrinth. In *Fish physiology* (Hoar, W. S. & Randall, D. J  
 445 eds.) pp. 207 - 240. Academic Press: New York.

446 Matheson, R. H., Huntsman, G. R. & Manooch, C. S. (1986). Age, growth, mortality,  
 447 food and reproduction of the Scamp, *Mycteroperca phenax*, collected off North  
 448 Carolina and South Carolina. *Bulletin of Marine Science* **38**, 300 – 312.

449 Maugé, L. A. (1990) Pomacanthidae. p. 841. In Check-list of the fishes of the eastern  
 450 tropical Atlantic (CLOFETA). (Quero, J.C., Hureau, J.C., Karrer, C., Post, A. &  
 451 Saldanha, L. eds.) JNICT, Lisbon, SEI, Paris; and UNESCO, Paris. Vol. 2.

452 Menezes, N. A., Backup, P. A., Figueiredo, J. L. & Moura, R. L. (2003). Catálogo das  
 453 espécies de peixes marinhos do Brasil. Museu de Zoologia USP, São Paulo.

454 Michael, S. W. (2004). Family Pomacanthidae/Angelfishes. In *Angelfishes &*  
 455 *Butterflyfishes*. pp. 296 – 323. T.F.H. Publications: New Jersey.

456 Moenich, D. R. (1991). Marine angels – *Pomacanthus*. T. F. H. Magazine **39**, 68-81.

457 Monteiro Neto, C., Cunha, F. E. A., Nottingham, M. C, Araújo, M. E, Rosa, I. L. &  
 458 Barros, G. M. L. (2003). Analysis of the marine ornamental fish trade at Ceará State,  
 459 northeast Brazil. *Biodiversity and Conservation* **12**, 1287 – 1295.

460 Munro, J. L. & Polunin, N. C. V. (1987). A decade of progress in coral reef fisheries  
 461 research: 1986-1995. *Proceedings of the 8<sup>th</sup> International Coral Reef Symposium* **2**,  
 462 2003-2008.

463 Nottingham, M. C., Cunha, F. E. & Monteiro Neto, C. (2000). Captura de peixes  
 464 ornamentais marinhos no Ceará. *Arquivos Ciências do Mar* **33**, 113 – 118.

465 Panella, G. (1971). Fish otoliths: daily growth layers and periodical patterns. *Science*,  
 466 137: 1124 – 1127.

467 Pyle, R. L. (1993). Marine aquarium fish. In *Nearshore marine resources of the South*  
 468 *Pacific. Information for fisheries development and management* (Wright, A. & Hill, L.  
 469 eds). pp. 135-176. Institute of Pacific Studies: Suva and Forum Fisheries Agency:  
 470 Honiara.

471 Radtke, R. L. (1987). Age and growth information available from the otoliths of the  
 472 Hawaiian snapper, *Pristipomoides filamentosus*. *Coral reefs* **6**, 19-25.

473 Randall, J. E. 1967. Food habits of reef fishes of the West Indies. *Studies in Tropical*  
 474 *Oceanography Miami* **5**, 665-847.

475 Roberts, C. M. & Hawkins, J. P. (1999). Extinction risk in the sea. *Trends in Ecology*  
 476 *and Evolution* **14**, 241-246.

477 Schwaborn, S. H. L. & Ferreira, B. P. (2002). Age structure and growth of the dusky  
 478 damselfish, *Stegastes fuscus*, from Tamandaré reefs, Pernambuco, Brazil. *Environmental*  
 479 *Biology of Fishes* **63**, 79 - 88.

480 Six, L. D. & Horton, H. F. (1977). Analysis of age determination methods for yellowtail  
 481 rockfish, canary rockfish, and black rockfish off Oregon. *Fishery Bulletin* **75**, 405 - 414.

482 Thresher, R. E. (1982). Courtship and spawning in the Emperor Angelfish *Pomacanthus*  
 483 *imperator*, with comments on reproduction by other pomacanthids fishes. *Marine*  
 484 *biology* **70**, 149 - 156.

485 Thresher, R. E. (1984). Angelfishes. In *Reproduction in reef fishes*. pp. 244 – 261. TFH  
 486 Publications: New Jersey.

487 Wood, E. M. (2001a). Collection of coral reef fish for aquaria: global trade,  
 488 conservation issues and management strategies. Marine Conservation Society, Ross-on-  
 489 Wye: UK.

490 Wood, E. M. (2001b). Global advances in conservation and management of marine  
 491 ornamental resources. *Aquarium Sciences and Conservation* **3**, 65 – 77.

492 Williams, T. & Bedford, B. C. (1974). The use of otoliths for age determination. In  
 493 *Ageing of fish* (Bagenal, T. B ed.). pp.. 114-123., Old Woking: Gresham Press.

494 Zekeria, Z. A., Weertman, S., Samuel, B., Kale-ab, T. & Videler, J. J. (2006). Growth  
 495 of *Chaetodon larvatus* (Chaetodontidae: Pisces) in the southern Red Sea. *Marine*  
 496 *Biology* **148**, 1113 - 1122.

Capítulo 3 - Variabilidade do ADN mitocondrial em populações simpátricas das espécies ornamentais recifais do gênero *Pomacanthus* (Actinopyterigii: Pomacanthidae) no Atlântico Ocidental.

Chapter 3\* - Mitochondrial DNA variability in sympatric populations of the ornamental reef fish *Pomacanthus* (Actinopyterigii: Pomacanthidae) in the southwestern Atlantic.

\* Capítulo em formato de artigo e padronizado de acordo com as normas da revista *Journal of Heredity*.

**Mitochondrial DNA variability in sympatric populations of the ornamental reef fish  
*Pomacanthus* (Actinopyterigii: Pomacanthidae) in the southwestern Atlantic**

Maria Elisabeth de Araújo<sup>a\*</sup>, Martin Lindsey Christoffersen<sup>b</sup> Caroline Vieira Feitosa<sup>a</sup>, Miguel  
Ângelo Martins Moreira<sup>c</sup>

<sup>a</sup> Departamento de Oceanografia, Centro de Tecnologia e Geociências, Universidade Federal  
de Pernambuco, 50670-901, Recife, Pernambuco, Brazil, elisabeth.araujo@ufpe.br,  
carol\_feitosa@hotmail.com

<sup>b</sup> Universidade Federal da Paraíba, Departamento de Sistemática e Ecologia, 58059-900, João  
Pessoa, Paraíba, Brazil, mlchrist@dse.ufpb.br

<sup>c</sup> Instituto Nacional do Câncer, Divisão de Genética, Rua André Cavalcanti 37, 20231-050, Rio de  
Janeiro, RJ, Brazil, miguelm@inca.gov.br

Running title: Mitochondrial DNA in populations of *Pomacanthus*

\* Corresponding author: M.E.A.

## Abstract

A number of studies have shown that reef fish populations have distinct color patterns, but these patterns are not associated with mt-DNA haplotypes. The aim of this study was to analyze the genetic diversity of *Pomacanthus paru* and *P. arcuatus* among populations of each species collected along the coast of northeastern Brazil. Thirty individuals were collected from coasts of Ceará, Pernambuco and Alagoas states. A region of the 16S rDNA was amplified. For phylogenetic analyses additional sequences from four species of the genus *Pomacanthus* were obtained in GenBank. Six mitochondrial DNA haplotypes were found for *P. paru* and a single haplotype for *P. arcuatus*. Topologies indicate that *P. paru* haplotypes are monophyletic and *Pomacanthus* species constitute a monophyletic group. The single haplotype of *P. arcuatus* was shared by individuals sampled in Pernambuco, Ceará (northeastern coast of Brazil) and the Caribbean. The considerations presented here represent a first step toward understanding the population structures of the sister species *Pomacanthus paru* and *P. arcuatus*. Complementary studies on larval settlement, physiological characteristics and anthropogenic actions directed at these populations are needed for a greater understanding of the dynamics of these species.

**Keywords:** mt-DNA, haplotypes, diversity, *Pomacanthus paru*, *Pomacanthus arcuatus*

## **Introduction**

Biodiversity based on morphology alone may considerably underestimate evolutionary lineages, particularly when morphological stasis persists after speciation events. This results in species that continue to diverge genetically in the absence of morphological differentiation, producing cryptic or sibling species (Knowlton 1986). Our inability to recognize cryptic species with traditional morphological characters has hindered our understanding of ecological and evolutionary processes. The negative consequences include underestimation of true levels of species-richness, overestimation of potential for long-distance dispersal in supposedly cosmopolitan species, failure to recognize cryptic biological invasions, and misinterpretation of ecological data (Knowlton et al 1992; Geller et al 1997; Rocha-Olivares et al 2001). Even when changes in allele frequencies and differences in sequence data among populations are not considered sufficient for attributing species-level status, organism and population genetic changes must represent the first steps linking micro-phylogeny with macro-phylogeny, because population genetics and phylogenetic history must in truth be parts of a temporal continuum (Avice 2009).

Most marine habitats have traditionally been thought to be highly interconnected by gene flow, because of the absence of obvious barriers to migration and because many marine taxa have pelagic larval stages that have been considered passive dispersers (Palumbi 1994). In contrast, high levels of biodiversity in marine systems suggest that genetic divergence and eventual speciation may be common and rapid in marine systems, indicating that marine biodiversity is actually much higher than earlier estimates (Mathews 2006).

Molecular genetic technologies have changed the way we describe and catalogue biological diversity. Mitochondrial sequence data has sometimes indicated no genetic structure within species (McMillen-Jackson and Bert, 2003), but usually points to an



increasing diversity of lineages, from two- to nineteen and perhaps more genetic or specific lineages (e.g., Rocha-Olivares et al 1999; Bichain et al 2007; Rocha et al 2008; Mathews and Anker 2009). When mitochondrial DNA differences become correlated with nuclear DNA sequence patterns, differences in geographical distribution, color distinctions, ecological and behavioral data, or even slight but constant morphological differences, such congruent evidence has been used to establish the presence of distinct evolutionary lineages (Mathews et al 2002; Sandoval-Castillo et al 2004; Thompson et al 2005).

Phylogeography, the geographical study of population genetics, is now a burgeoning discipline that is establishing a conceptual bridge between microevolutionary population genetics and macroevolutionary phylogenetics and historical biogeography (Avice 2009). These studies have the power to yield insights into the past as well as present evolutionary processes, having been used to uncover information about the roles of geographical history and ecology in generating marine species diversity (e.g., Williams and Reid 2004).

Phylogeography has triggered a renewed interest in speciation processes affecting Neotropical coral reef fish (Taylor and Hellberg 2005; Rocha and Bowen 2008). Reef fish are characterized by their bright and diversified coloration (Sale 1991). Many polytypic species are the result of fragile diagnoses and may include subspecies (Wiley 1981).

Comparing Brazilian populations of two species of reef fish that inhabit the Rocas atoll and the coast of the state of Ceará, Freitas et al (2003) differentiated fish stocks in *Acanthurus chirurgus*, but found unrestricted gene flow among the populations of *Cephalopholis fulva*. The majority of marine organisms are dispersed in the form of pelagic larvae that travel long distances by way of ocean currents. Upon reaching adulthood, many reef fish are benthic, territorial and remain associated with reef substrates (e.g. Sale 1980; Doherty and Williams 1988).

Species of the genus *Pomacanthus* occur in tropical and subtropical waters around the world (Allen et al 1998). Among the thirteen species of this genus, only *P. paru* and *P. arcuatus* occur in the Western Atlantic (Allen 1979; Allen et al 1998; Menezes et al 2003). There is agreement that these species generally share the same habitat and their home ranges overlap (Moyer et al 1983; Hourigan et al 1989). However, authors disagree with regard to the behavior between the two species. Moyer et al (1983) have recorded aggressive postures, whereas Hourigan et al (1989) found no agonistic behavior between the two species. Sympatric sister species such as *P. paru* and *P. arcuatus* can form hybrids and, although there is no record of a wild hybrid between these species (Deloach 1999), some have been produced in captivity (Moe 1976).

The aims of the present study were to analyze the mitochondrial DNA genetic diversity of *P. paru* and *P. arcuatus* and to determine whether there is geographic structuring of this diversity among the populations of each species collected at three sites along the coast of Brazil.

## **Materials and Methods**

Thirty specimens of *P. paru* and *P. arcuatus* were analyzed in the present study. All specimens were collected on the coast of the states of Ceará, Pernambuco and Alagoas (Fig. 1). Thirteen were collected from a single site in Ceará (seven *P. paru* and six *P. arcuatus*). Among these *P. paru* specimens, there were two individuals (belonging to Haplotype Hp1) with a different color pattern (more grayish).

Fourteen of the specimens (12 *P. paru* and two *P. arcuatus*) were acquired from the Itamaracá Island Fishing Colony in the state of Pernambuco (7°45'17.80''S–

34°49'26.46''W). The remaining three specimens were *P. paru* from the state of Alagoas. The specimens from Alagoas and Ceará were donated by the proprietors of the aquarium trade companies D'água and H&K Ornamental Fishes, respectively, who offered no information on the coordinates of the collection sites (Fig. 1).

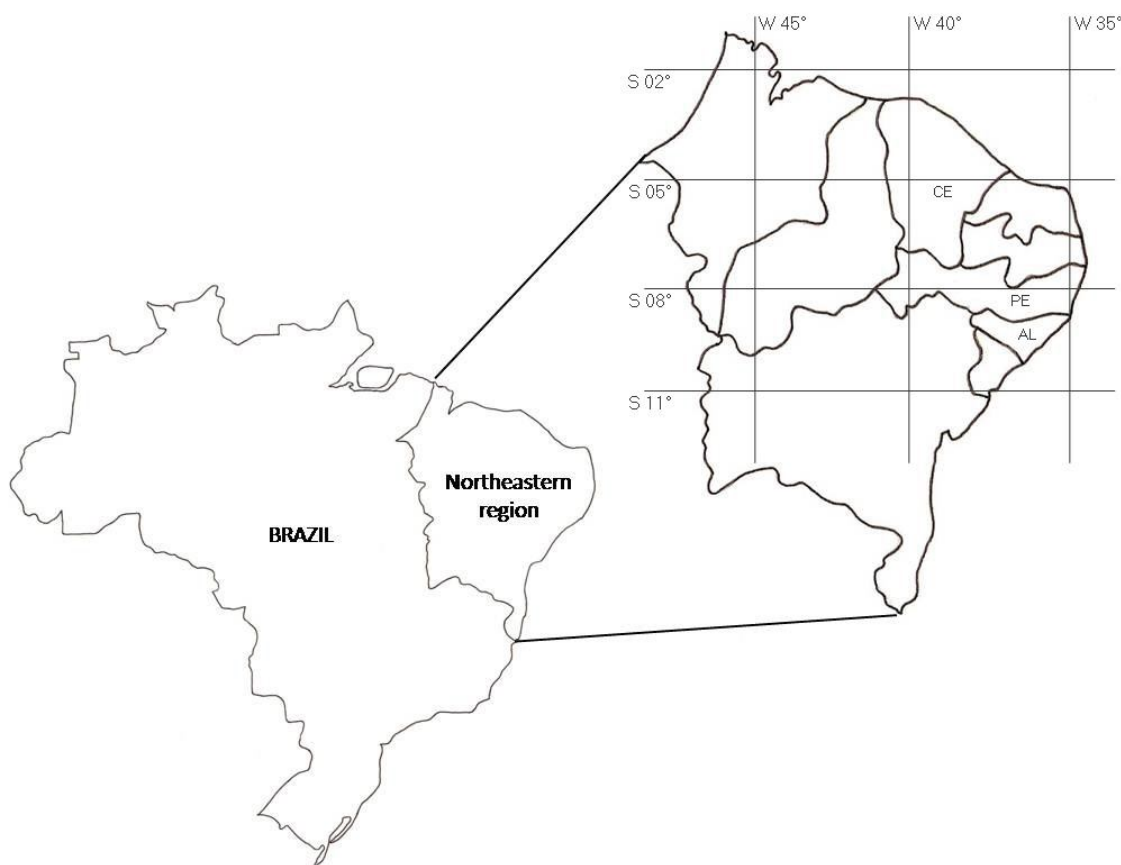


Figure 1: Location of sampled area. CE - Ceará state, PE - Pernambuco state and AL - Alagoas state.

Muscle and liver samples from each individual were stored in liquid nitrogen at the Laboratory of Applied Genetics of the Fishery Department of the Universidade Federal Rural de Pernambuco (UFRPE). The DNA extraction procedure followed the standard protocol described by Sambrook et al. (1989). Besides the collected specimens, sequences from the

GenBank were used for the specimens of *P. arcuatus* (AY530868 from the Caribbean) and *P. paru* (AY530852 from Belize) (Bellwood et al 2004).

A region of the 16S rDNA was amplified with the primers described by Simon et al. (1994): LR-J-12887 5'-CCG GTC TGA ACT CAG ATC ACG T-3' and LR-N-13398 5' CGC CTG TTT ACC AAA AAC AT 3'. PCR reactions were carried out as described in Bellwood et al. (2004). PCR products were purified using the GFX PCR DNA & Gel Band Purification Kit (GE-Healthcare) and sequenced in both directions using the DYEnamic ET Terminator Sequencing kit (GE-Healthcare) in an ABI-377 automated sequencer.

For the phylogenetic analyses, additional 16S rDNA sequences from four species of the genus *Pomacanthus* were obtained from the GenBank: *P. asfur* (AY530854), *P. semicirculatus* (AY530844), *P. sexstriatus* (AY530858) and *P. zonipectus* (AY530874). Sequences from *Chaetodontoplus duboulayi* (AY530846) and *Centropyge flavissimus* (AY530841) were used as outgroups. Nucleotide alignment was carried out by eye. A distance tree was constructed by neighbor-joining, with Kimura 2-parameter distance estimates, using the Mega 4.0 program (Tamura et al 2007). Parsimony topology was obtained using PAUP 4.0 (Swofford 1998). The bootstrap test (1000 replicates) was used in both analyses to evaluate node robustness. Network analysis was carried out for *P. paru* and *P. arcuatus* haplotypes, using the TCS software Phylogenetic Network Estimation Using Statistical Parsimony (Clement et al 2000). The choice of the ancestral haplotype was based on the central position in the network analysis as well as on frequency (Crandall and Templeton 1996). Haplotype diversity, nucleotide diversity indexes (Nei 1987), and Fu's  $F_s$  test of selective neutrality were estimated and carried out with the Arlequin 3.01 program (Excoffier et al 2005).

## Results

Six mitochondrial DNA haplotypes were found for *P. paru* (Haplotype Diversity,  $h = 0.67$ ; Nucleotide Diversity,  $\pi = 0.0018$ ) and a single haplotype was found for *P. arcuatus* (Table 1). Neighbor-joining and parsimony topologies (tree length = 154 steps; Consistency Index = 0.81) indicate that *P. paru* haplotypes are monophyletic and *P. paru* and *P. arcuatus* are sister species. These topologies had a similar arrangement (Fig. 2), revealing *Pomacanthus* species to be in a monophyletic group and *P. zonipectus* grouped with *P. paru* and *P. arcuatus*.

Table 1: Variable sites found in haplotypes of *P. paru* (Hp 1-6) and *P. arcuatus* (Ha 1) and number of individuals sharing a same haplotype per locality. Sequences from the Belize specimen belonging to Hp 4 haplotype and the specimen from *P. arcuatus* from the Caribbean were obtained from GenBank.

Haplotype	Variable position	Specimens / Locality			
	111111222233344 2016667366826634 6901679669235766	Ceará (CE)	Pernambuco (PE)	Alagoas (AL)	Caribbean
<i>P. paru</i>					
Hp 1 (11)	CTACGTA-GACAT-GC	4	5	2	
Hp 2 (1)	..T....-.....-		1		
Hp 3 (1)	.....-.....C-..			1	
Hp 4 (1)	.....-.....T..				1
Hp 5 (8)	.....-..G...-..	2	6		
Hp 6 (1)	T.....-..G...-..	1			
<i>P. arcuatus</i>					
Ha 1 (9)	..-..TTCGAT..GT..-AT	6	2		1

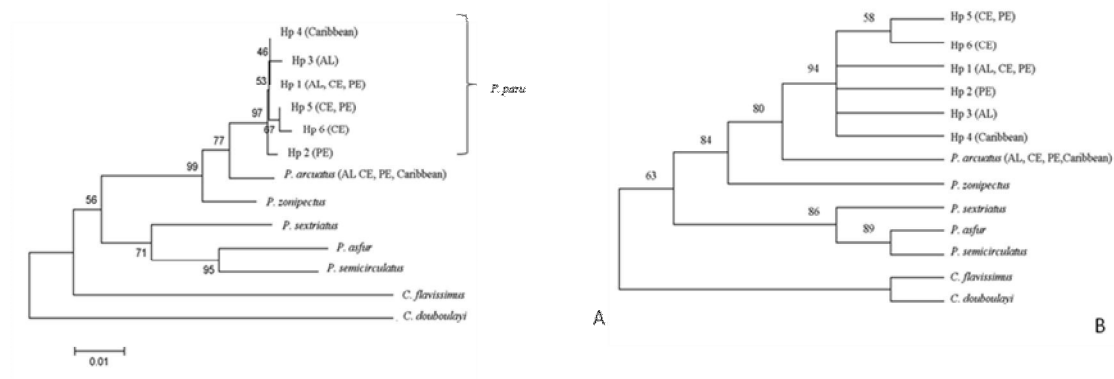


Figure 2: Neighbor-Joining (A) and Parsimony (B) topologies based on 16S rDNA mitochondrial sequences for the haplotypes of *Pomacanthus paru* and *Pomacanthus arcuatus*, and other *Pomacanthus* species. *C. flavissimus* - *Centropyge flavissimus* and *C. dubolayi* - *Chaetodontoplus duboulayi* were used as outgroups. Localities where the haplotypes were found are indicated between parenthesis: AL – Alagoas state, Brazil; CE – Ceará state, Brazil; PE – Pernambuco state, Brazil; and Caribbean. DC indicates specimens of *P. paru* with grayish coloration.

Concerning the six *P. paru* haplotypes, two had a greater frequency (Hp1 and Hp5, Table 1), with Hp1 present at all sample sites (Ceará, Pernambuco, and Alagoas). The network analysis (Fig. 3) suggests that Hp1 is the ancestral haplotype. The single haplotype of *P. arcuatus* (Ha1) was shared by individuals captured at two sites along the northeastern coast of Brazil (Pernambuco and Ceará) and is the same haplotype found in a Caribbean specimen.

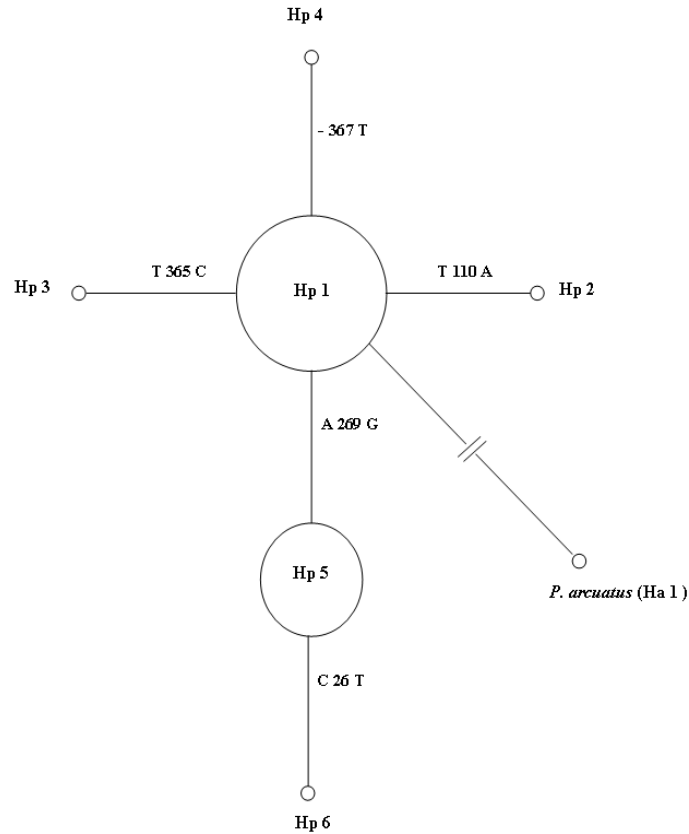


Figure 3: Parsimony Network of the mitochondrial haplotypes from *Pomacanthus paru* and *P. arcuatus*. The circles represent the haplotypes and the diameter is proportional to the number of specimens carrying each haplotype, except for *P. arcuatus* (Ha 1). Nucleotide substitutions between haplotypes were indicated.

The Fu's  $F_s$  test of selective neutrality for *P. paru* was significant marginally:  $F_s$  test = -2.35 and  $P = 0.022$  ( $F_s$  is significant when  $P < 0.02$ ) (Fu, 1997).

## Discussion

### Phylogeny

The molecular results obtained here indicate that *P. arcuatus* is a well-defined group, with only one haplotype identified, whereas the *P. paru* individuals form an independent

clade with at least six haplotypes for the tropical southwestern Atlantic. The negative  $F_s$  value and the star-like network indicate a demographic expansion for *P. paru* populations. This hypothesis can also be extended to *P. arcuatus* with only one haplotype.

#### *Genetic structure of species*

In the tropical southwestern Atlantic the *Pomacanthus* species complex shows strong genetic heterogeneity in the populations belonging to oceanic islands, presently identified as *P. paru*, and genetic homogeneity in coastal populations classified as *P. arcuatus*. Coastal populations of *Pomacanthus* thus appear to be genetically less diverse than offshore populations of this species, paralleling results recently obtained for dolphins in the Gulf of California (Segura et al 2006). These fish species may be the result of range shifts and secondary contact after allopatric speciation, or may be indicative of parapatric ecological speciation. On the whole, *P. paru* appears to have undergone some genetic differentiation into distinct haplotypes, although the occurrence of a same haplotype in different geographical localities indicates an incomplete differentiation into distinct populations.

In a single species, the number of haplotypes may vary considerably. The calculation proposed by Nei (1987) for haplotype diversity ( $h$ ) demonstrates that the greater it is, the greater the selective pressure on a given population will be and the more it will grow (Bowen et al., 2006). These authors consider the haplotype diversity among species of the genus *Centropyge* to be high ( $h = 0.991-1.0$ ) (14 in *C. argi*, 15 in *C. resplendens* and 17 in *C. aurantonotus*). As the authors analyzed a region of mt-DNA (d-loop) that is more variable than that analyzed in the present study (16S rDNA), the value obtained for *P. paru* ( $h = 0.6719$ ) could be higher if the d-loop were the molecular segment analyzed.

The present paper recognizes 6 previously unidentified haplotypes within the species *P. paru*. These evolutionary units are presently suggested by genetic divergence in the



mitochondrial DNA (shifts in allele frequencies). Independent lines of research are needed in order to determine if some of these haplotypes may indicate population and species level separations (for example, constant color pattern differences, morphological differences, or reproductive isolation data). Our data are still inconclusive in this respect. Genetic data from samples collected along the entire ranges of the species are still needed in order to establish such ecological and geographical patterns more precisely. The identification of discrete genetic lineages within a widely distributed species is often followed by the discovery of previously unobserved morphological differentiation (genealogical concordance, and subsequent recognition of the lineages as valid species (Rocha et al 2007). Our data increase our awareness of the net biodiversity in the marine environment of the tropical southwestern Atlantic, although we have so far been unable to associate color differences with population genetic differences within *P. paru*.

The genetic pattern observed in *P. paru* may not correspond to the species evolutionary history, but merely to the history of the mitochondria (Ballard and Whittock 2004). How differentiated conspecific lineages should be before they are elevated to species level is an unresolved question, and taxonomic decisions should thus be based on the congruence of independent data sets (e.g., including sequence data from nuclear genomes, color and morphological differences, data from ecological partitions and geographical separations), rather than on absolute levels of genetic divergence.

### *Color Patterns*

Mutations in genes that control morphological features, such as color, are commonly used to differentiate populations of marine animals (Knowlton and Mills 1992; Losos et al 1998, Endler et al 2005). The two individuals of *P. paru* sampled along the littoral of the state of Ceará with a distinct more grayish color pattern represent only a small fraction of our Hp1

haplotype. This could indicate that (1) there is no correlation between haplotype and color pattern, or (2) that there is further hierarchical structure undetected with molecular markers within *P. paru*.

A number of studies have shown that many fish populations have distinct color patterns, but these patterns are not associated with mt-DNA haplotypes (e.g., Rocha et al 2005; 2007; Schultz et al 2007). As far as our data indicate, this seems to be the case also for *Pomacanthus*. According to Rocha et al. (2007), there are two alternative explanations for this: 1) disruptive natural selection for color pattern (McCartney et al 2003; Ramon et al 2003); and 2) hybridization among color morphotypes (Puebla et al 2007). Although many species of reef fish are described based on their color patterns, morphological and genetic studies are essential for their diagnosis (Rocha 2004; Lima et al 2005).

#### *Distribution and Ecology*

Although the geographic distribution of these species coincides, *P. arcuatus* is recorded at greater latitudes in the northern hemisphere (New York), whereas *P. paru* (from Florida) reaches greater latitudes in the southern hemisphere. In Brazil, the latter species occurs as far as the state of Santa Catarina (26°58'01.67''S and 49°22'23.45''W), whereas *P. arcuatus* only occurs as far south as the state of Espírito Santo (19°11'22.84''S and 40°18'33.50''W) (Menezes et al 2003). *Pomacanthus paru* is the only species in this genus to occur in Brazilian oceanic islands, such as the Rocas atoll (Rosa and Moura 1997) and St. Peter and St. Paul's Rocks (Feitoza et al 2003). The two species differ in abundance. *P. paru* is more common along the Brazilian coast (Carvalho Filho 1999, Ferreira et al 2004) and less common in the Caribbean (Smith 1997) in comparison to *P. arcuatus*. This observation was confirmed by Hourigan et al. (1989), who carried out a transect census in the Caribbean (St. Croix, US Virgin Islands) and obtained a greater average density for *P. arcuatus*

(0.56/1000m<sup>2</sup>) in comparison to *P. paru* (0.31/1000m<sup>2</sup>). Based on the fact that *P. arcuatus* is more frequent in the Caribbean, it is possible that *P. paru* is more apt to occupy more recent habitats, such as those found along the Brazilian coast.

For a definitive picture of management units within *P. paru*, it will be necessary to survey the entire geographical and ecological range of the species, in order to fully understand the phylogeography, or geographical distribution of genetic lineages. For cases in which sibling species complexes occur in sympatry and show no clear differences in ecological specialization, Gittenberger (1991) has used the term nonadaptive radiation.

The partial overlap between the geographical ranges of *P. arcuatus* and *P. paru*, and also among the haplotypes of the latter species, indicates that conventional allopatric speciation by biogeographical barriers (e.g., Bernardi et al 2003) is not the only explanation for the speciation of tropical reef fishes. Parapatric ecological speciation by habitat partitioning at a local scale represents a mechanism that may explain the long-standing enigma of high biodiversity in coral reef faunas (Rocha et al 2005).

## Conclusions

Our data support the phylogenetic proximity between *Pomacanthus paru* and *P. arcuatus* and corroborate morphological, ethological (Pyle and Randall 1994), enzymatic (Chung and Woo 1998) and chromosomal (Galetti et al 2006) patterns. The molecular phylogeographical results of the present study confirm the taxonomic status of these sister species and suggest that, although they are sympatric (frequently syntopic), they have different evolutionary histories.

The considerations presented here represent a first step toward understanding the population structures of the sister species *Pomacanthus paru* and *P. arcuatus*, which have

economic value as ornamental fish. Complementary studies on larval settlement, physiological characteristics and anthropogenic actions directed at these populations are needed for a greater understanding of the dynamics of these species. The knowledge obtained herein on genetic distinctions into distinct haplotypes of *P. paru* has important conservation implications, considering that population diversity of this commercial fish may be larger than previously thought.

Although our paper deals primarily with the intraspecific genetic diversity of a commercial group of reef fish, stretches of mtDNA have nevertheless been shown to have tree-like histories within a species, and the branching structures of these gene genealogies may bear considerable analogy to phylogenetic trees summarizing historical relationships among reproductively isolated taxa and higher taxa, albeit on a different time scale (Avice 2009).

## References

- Allen GR, 1979. Butterfly and angelfishes of the world. Melle: Mergus Publishers, Melle.
- Allen GR, Steene R, Allen M, 1998. A guide to angelfishes & butterflyfishes. USA: Odyssey Publishing/Australia:Tropical Reef Research.
- Avice JC, 2009. Phylogeography: Retrospect and Prospect. J Biog. 36: 3–15.
- Ballard JW, Whitlock MC, 2004. The incomplete natural history of mitochondria. Mol Ecol. 13:729–744.
- Bellwood DR, van Herwerden L, Konow N, 2004. Evolution and biogeography of marine angelfish (Pisces: Pomacanthidae). Mol Phylogen Evol. 33:140–155.
- Bernardi G, Findley L, Rocha-Olivares A, 2003. Vicariance and dispersal across Baja California in disjunct marine fish populations. Evolution 57:1599–1609.

- Bichain JM, Boisselier-Dubayle MC, Bouchet P, Samadi S, 2007. Species delimitation in the genus *Bythinella* (Mollusca : Caenogastropoda : Rissooidea): a first attempt combining molecular and morphometrical data. *Malacologia* 49: 293–311.
- Bowen BW, Muss A, Rocha LA, Grant WS, 2006. Shallow mtDNA coalescence in Atlantic pygmy angelfishes (Genus *Centropyge*) indicates a recent invasion from the Indian Ocean *J Hered.* 97:1–12.
- Carvalho-Filho A, 1999. As espécies e suas famílias. In: *Peixes: Costa Brasileira*. São Paulo: Editora Melro. p. 40—83.
- Chung KC, Woo NYS, 1998. Phylogenetic relationships of the Pomacanthidae (Pisces: Teleostei) inferred from allozyme variation. *J Zool.* 246:215-231.
- Clement M, Posada D, Crandall K A, 2000. TCS: a computer program to estimate gene 6 genealogies. *Mol Ecol.* 9:1657–1659.
- Crandall KA, Templeton AR, 1996. *New uses for new phylogenies*. UK: Oxford University Press
- Deloach N, 1999. *Reef fish behavior: Florida, Caribbean, Bahamas*. Florida (USA): New World Publication.
- Doherty PJ, Williams DMB, 1988. The replenishment of coral reef fish populations. *Annu Rev Oceanogr Mar Biol.* 26:487–551.
- Endler JA, Wescotte DA, Madden JR, Robson T, 2005. Animal visual systems and the evolution of color patterns: sensory processing illuminates signal evolution. *Evolution* 59:1795–1818.
- Excoffier L, Laval G, Schneider S, 2005. Arlequin ver. 3.0: An integrated software package for population genetics data analysis. *Evol Bioinf Online* 1:47–50.

- Feitoza BM, Rocha, LA, Luiz-Júnior OJ, Floeter SR, Gasparini JL, 2003. Reef fishes of St. Paul's Rocks: new records and notes on biology and zoogeography. *Aqua, J Ichth Aquat Biol.* 7:61– 82.
- Ferreira CEL, Floeter SR, Gasparini JL, Joyeux JC, Ferreira BP, 2004. Trophic structure patterns of Brazilian reef fishes: a latitudinal comparison. *J Biogeogr.* 31:1093–1106.
- Freitas JEP, Araújo ME, Solé-Cava AM, 2003. Estruturação genética das populações de duas espécies de peixes recifais do Atol das Rocas e da costa do Ceará. *Trop Oceanogr.* 31:193–201.
- Fu YX, 1997. Statistical tests of neutrality of mutations against population growth, hitchhiking, and background selection. *Genetics* 147:915–925.
- Galetti Jr PM, Molina WF, Affonso PRAM, Aguilar CT, 2006. Assessing genetic diversity of Brazilian reef fishes by chromosomal and DNA markers. *Genetica* 126:161–177.
- Geller JB, Walton ED, Grosholz ED, Ruiz GM, 1997. Cryptic invasions of the crab *Carcinus* detected by molecular phylogeography. *Mol Ecol.* 6:901–906.
- Gittenberger E, 1991. What about nonadaptive radiation ? *Biol J Linn Soc.* 43:263–272.
- Hourigan TF, Stanton FG, Motta PJ, Kelley CD, Carlson B, 1989. The feeding ecology of three species of Caribbean angelfishes (family Pomacanthidae). *Env Biol Fish* 24:105–116.
- Knowlton N, 1986. Cryptic and sibling species among the decapod Crustacea. *J Crust Biol.* 6:356–363.
- Knowlton N, Mills EEK, 1992. The systematic importance of color and color pattern: Evidence for complexes of sibling species of snapping shrimp (Caridea: Alpheidae: *Alpheus*) from the Caribbean and Pacific coasts of Panama. *Proc San Diego Soc Nat Hist* 18:1–5.
- Knowlton N, Weil E, Weigt LA, Guzman HM, 1992. Sibling species in *Montastrea annularis*, coral bleaching, and the coral climate record. *Science* 255:330–333.

- Lima D, Freitas JE, Araujo ME, Solé-Cava AM, 2005. Genetic detection of cryptic species in the frillfin goby *Bathygobius soporator*. J Exp Mar Biol Ecol. 320:211–223.
- Losos JB, Jackman TR, Larson A, de Queiroz K, Rodriguez-Schettino L, 1998. Contingency and determinism in replicated adaptive radiations of Island lizards. Science 279:2115–2118.
- Mathews LM, 2006. Cryptic biodiversity and phylogeographical patterns in a snapping shrimp species complex. Mol Ecol. 15:4049–4063.
- Mathews LM, Anker A, 2009. Molecular phylogeny reveals extensive ancient and ongoing radiations in a snapping shrimp species complex (Crustacea, Alpheidae, *Alpheus armillatus*). Mol Phylog Evol. 50: 268–281.
- Mathews LM, Schubart CD, Neigel JE, Felder DL, 2002. Genetic, ecological, and behavioural divergence between two sibling snapping shrimp species (Crustacea, Decapoda: *Alpheus*). Mol Ecol. 11:1427-1437.
- McCartney MA, Acevedo J, Heredia C, Rico C, Quenouille B, Bermingham E, McMillan WO, 2003. Genetic mosaic in a marine species flock. Mol Ecol. 12:2963–2973.
- Mcmillen-Jackson AL, Bert TM, 2003. Disparate patterns of population genetic structure and population history in two sympatric penaeid shrimp species (*Farfantepenaeus aztecus* and *Litopenaeus setiferus*) in the eastern United States. Mol Ecol. 12:2895–2905.
- Menezes NA, Buckup PA, Figueiredo JL, Moura RL, 2003. Catálogo das espécies de peixes marinhos do Brasil. Museu de Zoologia USP, São Paulo.
- Moe MA, 1976. Rearing Atlantic angelfish. Mar Aquar. 7: 17-26.
- Moyer JT, Thresher RE, Colin PL, 1983. Courtship, spawning and inferred social organization of American angelfishes (genera *Pomacanthus*, *Holacanthus* and *Centropyge*; Pomacanthidae). Env Biol Fish 9:25–39.
- Nei M, 1987. Molecular evolutionary genetics. New York: Columbia University Press.

- Palumbi SR, 1994. Genetic divergence, reproductive isolation, and marine speciation. *Ann Rev Ecol System.* 25:547–572.
- Puebla O, Bermingham E, Guichard F, Whiteman E, 2007. Colour pattern as a single trait driving speciation in *Hypoplectrus* coral reef fishes? *Proc. R. Soc. Lond. B.* 274:1265–1271.
- Pyle RL, Randall JE, 1994. A review of hybridization in marine angelfishes (Perciformes: Pomacanthidae). *Env Biol Fish* 41:127-145.
- Ramon ML, Lobel PS, Sorenson MD, 2003. Lack of mitochondrial genetic structure in hamlets (*Hypoplectrus* spp.): recent speciation or ongoing hybridization? *Mol Ecol.* 12:2975–2980.
- Rocha LA, 2004. Mitochondrial DNA and color pattern variation in three western Atlantic *Halichoeres* (Labridae), with the revalidation of two species. *Copeia* 2004:770-782.
- Rocha LA, Bowen BW, 2008. Speciation in coral-reef fishes. *J Fish Biol.* 72: 1101–1121.
- Rocha LA, Craig MT, Bowen BW, 2007. Phylogeography and the conservation of coral reef fishes. *Coral Reefs*, 26:501-512.
- Rocha LA, Robertson RD, Roman J, Bowen BW, 2005. Ecological speciation in tropical reef fishes. *Proc R Soc B.* 272:573–579.
- Rocha LA, Rocha CR, Robertson DR, Bowen BW, 2008. Comparative phylogeography of Atlantic reef fishes indicates both origin and accumulation of diversity in the Caribbean. *Bmc Evolutionary Biology* 8: Art. 157. [cited 2009 March 5] Available from: <http://creativecommons.org/licenses/by/2.0>.
- Rocha-Olivares A, Fleeger JW, Folz DW, 2001. Decoupling of molecular and morphological evolution in deep lineages of a meiobenthic harpacticoid copepod. *Mol Biol Evol.* 18: 1088–1102.



- Rocha-Olivares A, Rosenblatt RH, Vetter RD, 1999. Cryptic species of rockfishes (Sebastes : Scorpaenidae) in the southern hemisphere inferred from mitochondrial lineages. J Hered. 90: 404–411.
- Rosa RS, Moura RL, 1997. Visual assessment of reef fish community structure in the Atol das Rocas Biological Reserve, off Northeastern Brazil. Proc 8<sup>th</sup> Internat Coral Reef Symp 1:983 - 986.
- Sale PF, 1980. The ecology of fishes on coral reefs. Annu Rev Oceanogr Mar Biol. 18:367-421.
- Sale PF, 1991. The Ecology of Fishes on Coral Reefs. San Diego: Academic Press.
- Sambrook J, Fritsch EF, Maniatis T, 1989. Molecular cloning: A Laboratory Manual. New York: Cold Spring Harbor lab.
- Sandoval-Castillo J, Rocha-Olivares A, Villavicencio-Garayzar C, Balart E, 2004. Cryptic isolation of Gulf of California shovelnose guitarfish evidenced by mitochondrial DNA. Mar Biol. 145: 983–988.
- Schultz JK, Pyle RL, DeMartini E, Bowen BW, 2007. Genetic connectivity among color morphs and Pacific archipelagos for the flame angelfish, *Centropyge loriculus*. Mar Biol. 151:167–175.
- Segura I, Rocha-Olivares A, Flores-Ramirez S, Rojas-Bracho L, 2006. Conservation implications of the genetic and ecological distinction of *Tursiops truncatus* ecotypes in the Gulf of California. Biol Cons 133:336–346.
- Smith CL, 1997. National Audubon Society field guide to tropical marine fishes of the Caribbean, the Gulf of Mexico, Florida, the Bahamas, and Bermuda. New York: Alfred A. Knopf, Inc.
- Swofford DL, 1998. PAUP\*: Phylogenetic analysis using parsimony (and other methods). Sunderland, MA: Sinauer Associates.

- Tamura K, Dudley J, Nei M, Kumar S, 2007. MEGA4: Molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol. Biol Evol.* 24:1596-1599.
- Taylor MS, Hellberg ME, 2005. Marine radiations at small geographic scales: speciation in Neotropical reef gobies (Elacatinus). *Evolution* 59:374–385.
- Thompson AR, Thacker CE, Shaw EY, 2005. Phylogeography of marine mutualists: parallel patterns of genetic structure between obligatory goby and shrimp partners. *Mol Ecol.* 14: 3557–3572.
- Wiley EO, 1981. *Phylogenetics: the theory and practice of phylogenetic systematics*. USA: John Wiley & Sons, Inc.
- Williams ST, Reid DG, 2004. Speciation and diversity on tropical rocky shores: a global phylogeny of snails of the genus *Echinolittorina*. *Evolution* 58:2227–2251.

Capítulo 4 - **Um método novo e rápido para avaliar a sustentabilidade do bycatch de peixes ornamentais dos recifes de corais.**

Chapter 4\*† - **A rapid new method for assessing sustainability of ornamental fish bycatch from coral reefs.**

\* Capítulo em formato de artigo e padronizado de acordo com as normas da revista *Marine and Freshwater Research*.

† Artigo publicado na revista *Marine and Freshwater Research*.

**A rapid new method for assessing sustainability of ornamental fish bycatch from coral reefs.**

Caroline Vieira Feitosa<sup>A,B</sup>, Beatrice Padovani Ferreira<sup>A</sup>, Maria Elisabeth de Araújo<sup>A</sup>

A - Departamento de Oceanografia, Centro de Tecnologia e Geociências, Universidade  
5 Federal de Pernambuco, 50670-901, Recife, Pernambuco, Brazil.

B - Corresponding author. Email: carol\_feitosa@hotmail.com

**Abstract**

The ornamental fish trade has a considerable potential for damaging reef environments.

10 Nearly 120 species are currently harvested in the Brazilian trade and little attention has been paid to the effects of the accidental catch of these species. Records of ornamental fish bycatch and the susceptibility and resilience of species from the major Brazilian families in the aquarium trade were examined to test a method and determine the sustainability of these catches. Each species was assessed based on two groups of  
15 criteria: (1) relative vulnerability to capture; and (2) population resilience regarding mortality due to fishing efforts. Each species was also ranked (1-3) based on individual criteria regarding ecology, biology, catchability, numbers of individuals exported by aquarium companies and mortality index. These criteria were then weighted and each species rank in the two groups of criteria was plotted. Nineteen species from 10 families  
20 were recorded. The species least likely to have sustainable catch rates were *Holacanthus tricolor*, *H. ciliaris*, *Chaetodon striatus* and *Pomacanthus paru*. This approach is a useful alternative for management efforts and the assessment of catch sustainability in newly-developed multi-species fisheries with limited available data, which occur in areas of high species richness such as reefs.

25 **Additional keywords:** aquarium, trap, fishery, susceptibility, resilience, Pomacanthidae, Chaetodontidae.

## Introduction

Species classified as bycatch (Eayrs 2007) are caught in fisheries that target other groups and are then either used or discarded. It is estimated that 7.3 million tonnes of fish were discarded globally in commercial fisheries in the period of 1995-2005 (Kelleher 2005). This is equivalent to 8% of worldwide marine fish production (Eayrs 2007). The global effect of fisheries on bycatch species is poorly known, although some studies have documented ecological changes on the species level (Casey and Myers 1998; Pope *et al.* 2000). Even less attention has been paid to accidental catches affecting reef fish, including ornamental species.

Beauty and perfect patterns on ornamental reef fish are requirements imposed by the international aquarium trade (Araújo and Albuquerque Filho 2005), which is one of the world's largest industries. The import value worldwide of marine fish and invertebrates is estimated to be between US\$ 24 and 40 million annually (Wood 2001a). Nearly 120 species are currently harvested in the Brazilian aquarium trade (Monteiro Neto *et al.* 2003). Most belong to the Pomacanthidae family (Araújo and Albuquerque Filho 2005), while the Chaetodontidae are among the most exported (Monteiro Neto *et al.* 2003; Nottingham *et al.* 2000). Although other fisheries that use traps have an effect on ornamental marine fish stocks (through by-catch), these stocks have only been managed through a Federal Act (56/04, established in 2004 by the Brazilian environmental protection agency – Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis – IBAMA) that monitors ornamental fish trade through export quotas. The purpose of this Federal Act is to administer this activity through the establishment of export quotas.

Traps are the predominant gear employed to capture reef fish in the Caribbean (Swingle *et al.* 1970; Garrison *et al.* 1998) as well as other places in the world (e.g. Arabian Gulf, Seychelles, Papua New Guinea) (Dalzell 1996) and are responsible for the decline of many reef species, which are fishery targets (Wolff *et al.* 1999). Among the organisms caught by these devices, the most frequently caught ornamental fish belong to the following genera: *Acanthurus*, *Chaetodon*, *Bodianus*, *Holacanthus* and *Pomacanthus* (Garrison *et al.* 1994; Wolff *et al.* 1999). In Brazil, trap fisheries targeting reef fish have been on the rise in the last ten years (Ribeiro 2004). The target

65 species of these fisheries belong to the genera *Acanthurus*, *Cephalopholis*, *Lutjanus*,  
*Pseudupeneus* and *Sparisoma* (Ribeiro 2004). Because by-catch, including ornamental  
species, is returned to the sea and not recorded, there are no data or regulations available  
about the actual catch of ornamental species.

Due to the increasing pressure to manage the impact of fisheries on non-target  
70 species and the uncertainty of how this is to be done, Stobutzki *et al.* (2001) has  
developed a broad method using qualitative measures of the biological and ecological  
characteristics of species to rank vulnerability to a known threat. These authors examine  
the impact of trawling on the catch sustainability of teleost bycatch species, identifying  
the species with the least likelihood of catch sustainability in bycatch. Such species  
75 should then be the focus of research and management. The authors have recorded at  
least 411 species from 99 families as bycatch, identifying 15 species with the least  
likelihood of catch sustainability as well as the 13 with the most likelihood of catch  
sustainability. This approach was designed to overcome problems such as the high  
diversity of bycatch in tropical fisheries and the lack of information on individual  
80 species. The method is versatile and can also be applied to other fisheries, although the  
criteria employed may vary.

The present study describes a rapid, innovative method, based on the method  
developed by Stobutzki *et al.* (2001), for evaluating the impact of trap fisheries on  
ornamental species using biological data and the portion of the population caught. The  
85 intention is to answer the following two questions. (1) Is the method appropriate for  
ornamental reef fish? (2) Is the capture of ornamental reef fish as by-catch sustainable?  
The present study records the occurrence of ornamental fish caught as by-catch and  
assesses the susceptibility and resilience of fish from the Pomacanthidae and  
Chaetodontidae families.

## 90 **Materials and Methods**

### *Sampling*

The fish recorded here were acquired from the fishing colony of Itamaracá  
island located on Pilar beach (7°45'17.80'' S – 34°49'26.46''W) (Itamaracá, state of  
Pernambuco, Brazil – Fig. 1) and were caught as bycatch while trap fishing for the  
95 spotted goatfish, *Pseudupeneus maculatus* and parrotfish, *Sparisoma* spp. This kind of  
fishing occurs throughout the year and is carried out on alternating days. The boats

leave Pilar beach at midnight and arrive at Tambaba beach, 46 km away, ( $7^{\circ}22'2.93''S$  –  $34^{\circ}47'48.51''W$ ) (state of Paraíba) in the early morning. Four boats are employed in this fishery. Each has a fishing effort of 15 to 20 traps and trap mesh varying between 1 and 7 cm. The traps are deployed unbaited next to reefs and hauled 6 hours later. Occasionally, depending on factors such as wind and currents, some traps are left for longer periods and only hauled after 48 hours. The fish are stored for approximately 7 hours in 100-L containers. For the present study, weekly samplings were performed from April to June 2006. The fishermen were asked to bring back all of specimens with ornamental characteristics, which were then bought. In the case of *H. ciliaris* and *P. paru*, this procedure was carried out over a longer period (March 2006 to February 2007), as these two species were the focus of another study.

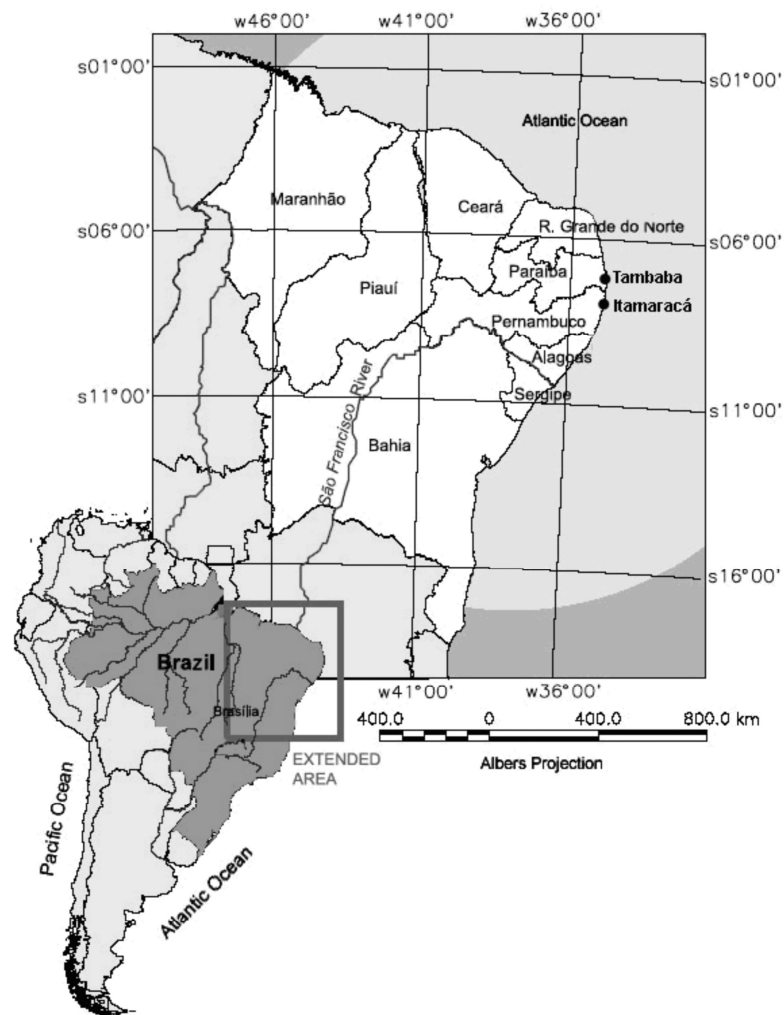


Figure 1: Location of sampled area.

110 The sampled fish were placed in polystyrene chests with ice and taken to the Nekton and Aquaculture Laboratory of the Oceanography Department of the Universidade Federal de Pernambuco, where they were identified, measured and photographed.

*Analysis of catch sustainability of ornamental reef fish bycatch*

115 Only the most abundant species were analysed, i.e., species from the Pomacanthidae and Chaetodontidae families. Rare species for Brazilian northeastern coast, such as *C. ocellatus* and *P. arcuatus* (Thresher 1980; Carvalho Filho 1999) were not surveyed due to the small sample size. Information on the biological characteristics of the selected species was taken from specific publications, such as Robins and Ray  
120 (1986); Thresher (1980, 1984); Allen (1998), Hourigan *et al.* (1989) and Froese and Pauly (2007).

The method proposed to access the sustainability of selected bycatch species was adapted from Stobutzki *et al.* (2001) and applied based on two scored sets of criteria: (1) relative vulnerability to capture by traps and mortality due to trap fishery; and (2)  
125 the recovery capacity of a species after the population is depleted. The criteria were originally developed during a study on the sustainability of all North Australian Prawn Fishery (NPF) prawn-trawl vertebrate bycatch (Stobutzki *et al.* 2000). These criteria were modified and assigned relative importance weights by the NPF Assessment Group (Milton, 2001). Values ranging 1 to 3 were attributed to each sustainability indicator,  
130 for which 1 denotes the highest value for the susceptibility criteria and the lowest for resilience. Depending on the criterion, these values were based on either categorical or continuous data. When no information was available to assign the divisions between values, the range of the data was divided into thirds in order to create categories (Table 1). The rank matrix constructed from these values for each set of criteria are plotted as  
135 axes (resilience *versus* susceptibility) on a graph indicating the species with the greatest catch sustainability. This graph is recommended only if the different species have different values on each axis. According to Stobutzki *et al.* (2001), when species-specific information is unavailable, a species is given the same value as other species within the same family. With criteria for which family members are not similar or for  
140 which no family information is available, the rank of 1 is given as a precautionary measure.



The criteria adopted to analyse species susceptibility (axis 1) to capture and possible mortality due to fishery efforts were adapted from Stobutzki *et al.* (2001) and are summarised as follows: (1.1) water column position – as traps are deployed on the substrate (Mahon and Hunte, 2001; Garrisson *et al.* 2004), benthic and demersal species are more susceptible to capture than the pelagic species; (1.2) preferred habitat – species are ranked according to their preference for rocky habitats. This criterion addresses the likelihood that a species habitat overlaps the area in which trap fishery occurs. Coral reef habitats have been historically targeted by trap fisheries (Wolff *et al.* 1999). Thus, species that live near shore and depend on a specific habitat, such as coral reefs, are highly vulnerable to a possible risk of extinction (Dye *et al.* 1994; Roberts and Hawkins, 1999); (1.3) day/night catchability – as trap fishery is a daytime operation, species with diurnal habits are more susceptible to capture by traps; (1.4) diet – addressing whether eating habits attract species to trap fishery areas. Thus, species that generally feed on benthic or demersal organisms found on reefs are more vulnerable to capture. Invertebrate feeders (mobile and sessile) are the dominant trophic guild in traps deployed in coral reef habitats (Garrison *et al.* 2004); (1.5) depth range – this trap fishery occurs in areas between 30 and 40 m. According to Wolff *et al.* (1999), trap fisheries in adjacent coral reef and gorgonian habitats range from 3 to 38 m. Overlap between the depth range of the trap fishery area and that of the preferred habitat will affect the vulnerability of a species to capture more than that of others that live in either deeper or shallower areas. Fish that have a narrow vertical depth range are highly vulnerable to extinction (Dye *et al.* 1994; Roberts and Hawkins, 1999) (Table 1).

Six criteria adapted from Stobutzki *et al.* (2001) were employed to analyse species resilience: (2.1) the maturity criterion – if most individuals caught are larger than their size at first maturity, the species has a greater probability of recovering from a declining population; (2.2) maximum size – species that attain large sizes tend to live longer and therefore the resilience of the population is lower than species with smaller body sizes. A slow life history (e.g. late maturity, long reproductive span and large body size) is related to low maximum population growth rates and, consequently, such species are more vulnerable and have less capacity to recover from exploitation (Hutchings 2001; Reynolds *et al.*, 2001, 2002); (2.3) removal rate – species-based quotas established by the Brazilian environmental protection agency (IBAMA) to

175 ensure that harvests are maintained at a sustainable level. The ornamental species  
fishery is highly selective and certain species such as chaetodontids and pomacanthids  
are preferred targets. Among those, the species which are rare may be vulnerable to  
over-exploitation (Wood 2001b); (2.4) reproductive strategy – species capable of  
dispersion over long distances are less vulnerable to extinction (Dye *et al.* 1994;  
Roberts and Hawkins, 1999). Thus, the resilience of R-strategist species is generally  
180 higher than that of K-strategist species; (2.5) hermaphroditism – hermaphroditic  
species, especially protandric species, may be at great risk of extinction, as fisheries  
select individuals of certain sizes, which has a negative effect on reproduction (the  
fishing effort is directed at only one sex) (Dye *et al.* 1994; Roberts and Hawkins, 1999;  
Hawkins and Roberts, 2003); (2.6) mortality index – this index was proposed by Sparre  
185 and Venema (1992) and Stobutzki *et al.* (2001). This index can be derived from the  
length frequency of a species and von Bertalanffy growth parameters. However, such  
parameters are not available for most bycatch species. Thus, the index can be calculated  
as follows:

Mortality index (Z):  $(L_{\max} - L_{\text{mean}}) / (L_{\text{mean}} - L_{\min})$

190 in which  $L_{\max}$  is the maximum length recorded in the literature;  $L_{\text{mean}}$  is the mean  
length of total number of individuals caught;  $L_{\min}$  is the minimum length of the  
individuals in the sample. In the present study, the mortality index range was calculated  
(0.13 – 0.74) and divided into thirds for the division between ranks (Table 1).

## Catch sustainability of ornamental fish bycatch...

Table 1: The criteria used to assess the susceptibility of bycatch species to capture and mortality due to traps and their recovery. For each criterion the weighting score and definition of the three ranks is given.

Criteria	Weighting score	1	2	3
<b>Susceptibility</b>				
Water column position	3	Demersal or benthic	Benthopelagic	Pelagic
Preferred habitat	3	Rocky habitats with gravel and sand	Habitats with soft or muddy bottoms	No fishing habitats
Day/night catchability	2	Higher catch rate at day	No difference between day and night	Higher catch rate at night
Diet	3	Feed on demersal or bent. organisms	Feed on benthopelagic organisms	Feed on pelagic organisms
Depth range	1	Less than 40m	-	Deeper than 40m
<b>Resilience</b>				
Maturity	3	Mature individuals < 50%		Mature individuals > 50%
Maximum size	3	Max. size $\geq$ 50 cm	16 cm < Max. size < 50 cm	Maximum size $\leq$ 16 cm
Removal rate	3	Catch $\geq$ 3500 inds/comp/yr	1000 < catch < 3500 inds/comp/yr	Catch $\leq$ 1000 inds/comp/yr
Reproductive strategy	2	Bear live young or brood young	Guard eggs and/or young	Broadcast spawners
Hermaphroditism	1	Protandric hermaphrodites	Protoginic hermaphrodites	Dioecious
Mortality index	1	Mortality index $\geq$ 3.19	0.96 < mortality index < 3.19	Mortality index $\leq$ 0.96

Caption: bent – benthic; Max – Maximum; inds/comp/yr – individuals/company/year

*Analysis of criteria*

200 The overall ranks of susceptibility or resilience were calculated according to the equation below:

$$S_i = \sum_{j=1}^n w_j R_i / \sum_{j=1}^n w_j$$

205

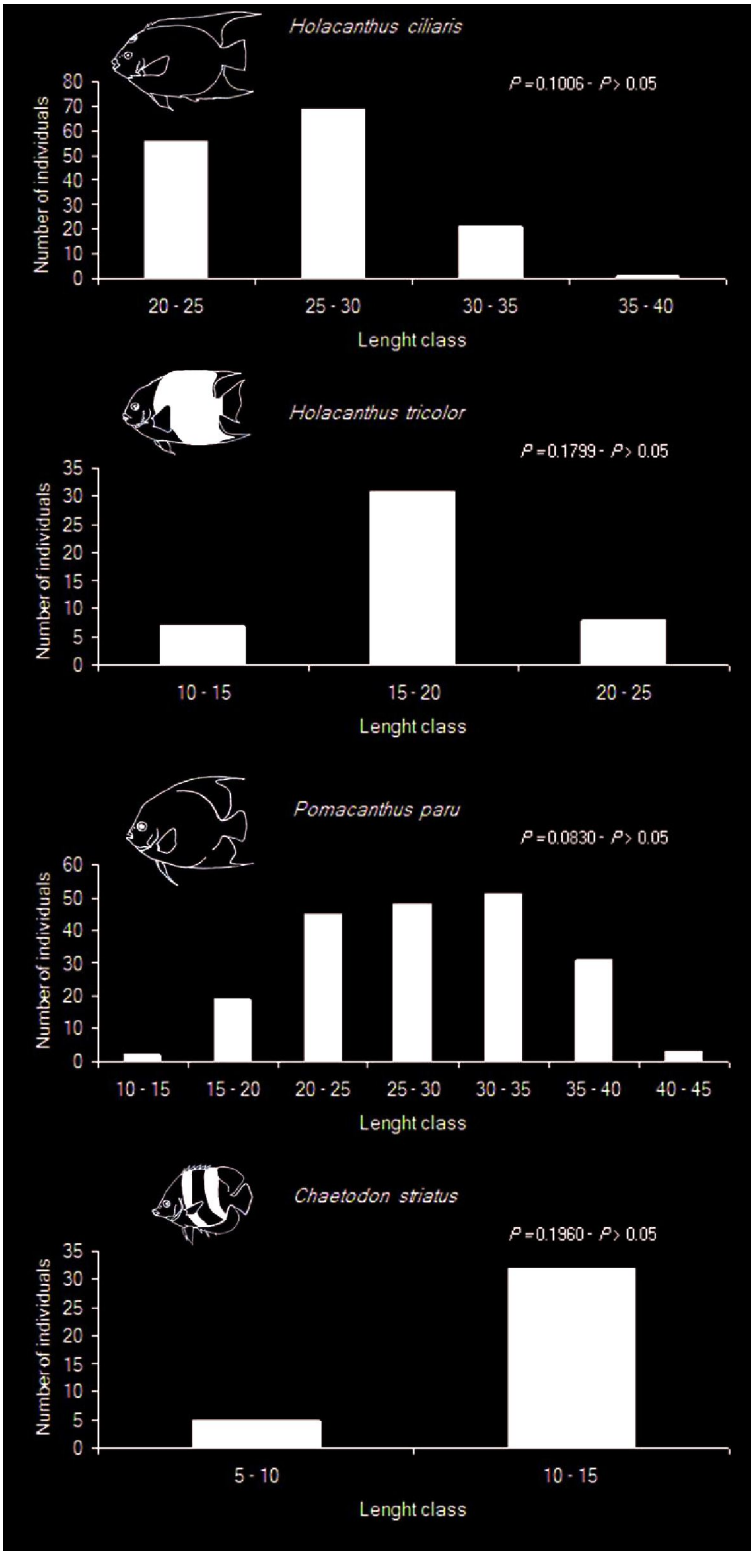
in which  $S_i$  is the total susceptibility or resilience rank for species  $i$ ;  $w_j$  is the weight for criterion  $j$ ;  $R_i$  is the rank of species  $i$  for criterion  $j$ ; and  $n$  is the number of criteria for each axis.

*Statistical analysis*

210 Data normality was evaluated using the Kolmogorov-Smirnov test (BioEstat 5.0 software – Instituto de Desenvolvimento Sustentável Mamirauá). The parametric Z test (BioEstat 5.0 software) was used to determine the probability of a captured individual being smaller than size at first maturity and the probability of a captured individual having bred at least once. Correspondence analysis (Multivariate Statistical Package –  
215 MVSP 3.1 software – Kovach Computing Services) was used to determine and describe the correspondence between species and criteria on a same graph. This multivariate analysis was used only for criteria related to resilience, as values regarding susceptibility did not vary among species. Since the variance of the reproductive strategy criterion was equal to zero, this criterion was not applied in the present  
220 analysis.

**Results**

Among the fish caught, 19 were recorded as bycatch species and belong to 14 genera and 10 families (Table 2). Of these, 100% come from reef environments and are used in the aquarium trade and only one species (*Equetus lanceolatus*) was not  
225 authorised for export. A total of 430 individuals were sampled: 147 *H. ciliaris*, 46 *H. tricolor*, 200 *P. paru* and 37 *C. striatus*. Length frequency distribution by species (Fig. 2) revealed a broad length range among the individuals sampled.



230

Figure 2: Length distributions for the four analyzed species. P values refers to normality.

Table 2: Species registered as bycatch, with the respective common names.

Family	Species	Common name	n
Syngnathidae	<i>Hippocampus reidi</i> Ginsburg, 1933	Longsnout seahorse	2
Synodontidae	<i>Synodus foetens</i> (Linnaeus, 1766)	Inshore lizardfish	2
Aulostomidae	<i>Aulostomus maculatus</i> Wheeler, 1955	Trumpetfish	1
Sciaenidae	<i>Equetus lanceolatus</i> (Linnaeus, 1758)	Jack-knife fish	2
Sciaenidae	<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)	Highhat	1
Chaetodontidae	<i>Chaetodon ocellatus</i> Bloch 1787	Spotfin butterflyfish	2
Chaetodontidae	<i>Chaetodon striatus</i> Linnaeus, 1758	Banded butterflyfish	37
Pomacanthidae	<i>Holacanthus ciliaris</i> (Linnaeus, 1758)	Queen angelfish	147
Pomacanthidae	<i>Holacanthus tricolor</i> (Bloch, 1795)	Rock beauty	46
Pomacanthidae	<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	Gray angelfish	3
Pomacanthidae	<i>Pomacanthus paru</i> (Bloch, 1787)	French angelfish	200
Labridae	<i>Bodianus rufus</i> (Linnaeus, 1758)	Spanish hogfish	23
Labridae	<i>Bodianus pulchellus</i> (Poey, 1860)	Spotfin hogfish	3
Labridae	<i>Halichoeres dimidiatus</i> (Agassiz, 1831)	Brazilian yellowcheek wrasse	31
Labridae	<i>Halichoeres brasiliensis</i> (Bloch, 1791)	Brazilian wrasse	10
Monacanthidae	<i>Cantherhines macrocerus</i> (Hollard, 1855)	Whitespotted filefish	1
Monacanthidae	<i>Stephanolepis hispidus</i> (Linnaeus, 1766)	Planehead filefish	1
Ostracidae	<i>Lactophrys trigonus</i> (Linnaeus, 1758)	Buffalo trunkfish	9
Tetraodontidae	<i>Canthigaster figueiredoi</i> Moura & Castro (2002)	Southern Atlantic sharpnose-puffer	1

235

All information on susceptibility criteria is species-specific and the four species evaluated in the sustainability analysis obtained the same value (1) for this criterion. Rank resilience ranged from 1.92 to 2.54; *H. tricolor* and *P. paru* obtained the smallest and the largest values, respectively (Table 3). Based on these results, it can be affirmed that *H. tricolor* has the least catch sustainability, followed by *H. ciliaris* and *C. striatus*. *P. paru* is the most capable of recovering from a population decline.

240

Mean and smallest lengths for *H. tricolor* and *C. striatus* individuals were under size at first maturity. There was a 71% probability of *H. tricolor* individuals and a 92% probability of *C. striatus* being caught at sizes smaller than first maturity. Contrarily, the probability of captured individuals being larger than maturity size was 76% for *H. ciliaris* and 70% for *P. paru*. *H. ciliaris* had the highest mortality index ( $Z=3.19$ ) and *P. paru* had the lowest ( $Z=0.96$ ) (Table 4).

245

# Catch sustainability of ornamental fish bycatch...

Table 3: The ranking of the species on the criteria on the resilience axis.

Species/Weight	Resilience						Rank
	3	3	3	2	1	1	
	Maturity	Maximum size	Removal rate	Reproductive strategy	Hermaphroditism	Mortality index	
<i>Holacanthus ciliaris</i>	3	2	1	3	2	1	2.08
<i>Holacanthus tricolor</i>	1	2	2	3	2	2	1.92
<i>Pomacanthus paru</i>	3	2	2	3	3	3	2.54
<i>Chaetodon striatus</i>	1	3	3	3	3	2	2.46

Table 4: The length at first capture, length at maturity, probability of individuals caught are below of the length at maturity, probability of breeding, mean and maximum length (Max. length) of the species and the mortality index (Z) of each.

Species	n	L' (cm)	Lm (cm)	Prob. L < Lm (cm)	Prob. of breeding	L mean $\pm$ S. E (cm)	Max. Length (cm)	Z
<i>H. ciliaris</i>	147	20.88	24.3	p = 0.24	p = 0.76	26.58 $\pm$ 0.2672	45 (Allen, 1998)	3.19
<i>H. tricolor</i>	46	11.32	19	p = 0.71	p = 0.29	17.61 $\pm$ 0.3646	35 (Allen, 1998)	2.68
<i>P. paru</i>	200	13.44	25	p = 0.30	p = 0.70	28.30 $\pm$ 0.4510	41 (Cervigón, 1993)	0.96
<i>C. striatus</i>	37	9.09	13.2	p = 0.92	p = 0.07	11.26 $\pm$ 0.2257	16 (Lieske and Myers, 1994)	2.17

Caption: n – number of individuals, L' – length at first capture, Lm – length at maturity, Prob – probability, Max. Length – maximum length, Z – mortality index.

The correspondence analysis for resilience revealed that *H. tricolor* and *C. striatus* are linked to the removal index. *H. ciliaris* and *P. paru* are associated through maturity criteria. *Chaetodon striatus* and *P. paru* are grouped with maximum size and mortality index criteria, respectively. Negative relationships were determined for *H. ciliaris* and the removal rate and mortality index; *H. tricolor* and *C. striatus* with maturity criteria; and *P. paru* with maximum size. The percentage of the cumulative variation for the two axes was 99.53% (Fig. 3).

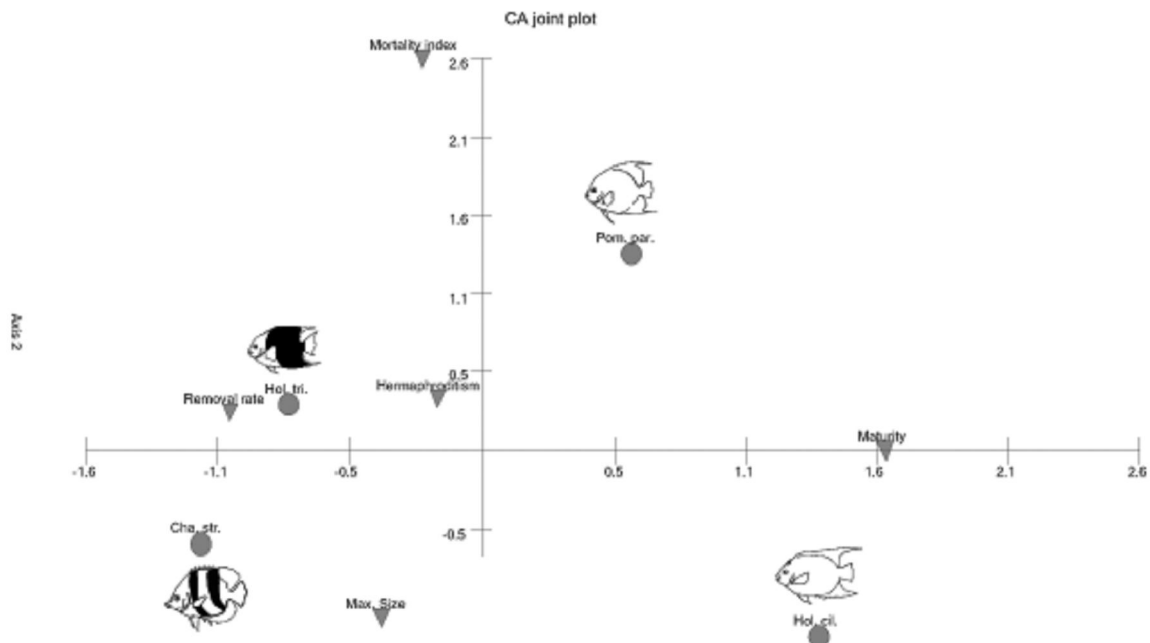


Figure 3: Correspondence analysis showing the relationship between species and criteria used to evaluate the recovery capacity.

Caption: *Hol. cil.* – *Holacanthus ciliaris*, *Hol. tri.* – *Holacanthus tricolor*, *Cha. str.* – *Chaetodon striatus*, *Pom. par.* – *Pomacanthus paru*, Max. Size – Maximum size.

## Discussion

The present study demonstrates that trap fishery is not selective. The catching of ornamental fish occurs frequently and likely causes considerable impact. Ornamental species are already considered seriously endangered due to fishery efforts (Cheung *et al.* 2007). Individuals caught are not landed and are usually returned to the sea (many still alive, according to the fishermen). Despite this, the survival of these individuals is



doubtful, as the traps are hauled quickly to surface and problems related to decompression have often been reported (Burns and Restrepo 2002).

The method applied in the present study was developed with the purpose of classifying ornamental fish species according to possible risk of extinction due to being captured as bycatch. This analysis is influenced by the length range and proportion of young and immature individuals sampled, as criteria that are directly related to specimen size are employed, such as length at first maturity and mortality index. Different mesh sizes are used in this fishery activity, which may have contributed toward the broad length range among the individuals sampled. Due to the small sample size for *H. tricolor* and *C. striatus*, the length range of the specimens caught may not be representative. Thus, for these species, the analysis must be considered only for demonstration (exploratory) purposes, even though there was a trend toward normality in the length distribution data. The present study is the first to test a method with such goals directed at trap fisheries and using criteria based on life history.

Specific biological information is optimised when well-defined criteria are applied, including characteristics that affect species sensitivity or risk of extinction (Dye *et al.* 1994; Roberts and Hawkins 1999; Stobutzki *et al.* 2001). Stobutzki *et al.* (2001) apply the value 1 in the matrix when there is no specific information on certain criterion. In the present study, all biological information was species-specific. Although the analysis presented is subjective, this may be the only one available to evaluate and monitor multi-specific fisheries (Dulvy *et al.* 2003), especially newly developed fisheries.

#### *Susceptibility to capture by traps*

Species caught by a given equipment share similar characteristics, such as habitat, diet and period of activity. Thus, reef fish are considered highly susceptible to trap fisheries. Pomacanthidae are widely distributed in depths ranging from 1 to 100 m (Allen 1998; Michael 2004; Froese and Pauly 2007), whereas Chaetodontidae occur between 3 to 55 m (Allen 1998; Robins and Ray 1986; Froese and Pauly 2007). The former feed on sponges, zoanthids, tunicates, hydroids, bryozoans and algae (Randall 1967; Allen 1998; Michael 2004). Chaetodontidae eat worms, coral polyps and mollusc eggs (Allen 1998). The diet and period of activity of these fish favour their capture, as they only feed during the day (Starck and Davis 1966) and the organisms they eat are

sessile, which are generally encrusted on reefs. Dependence on a particularly susceptible habitat (e.g. coral reefs), either alone or combined with overfishing, leads to considerable captures of bycatch species and makes these species all the more vulnerable (Reynolds and Jennings 2000).

#### *Resilience*

Species with late sexual maturity are highly vulnerable to the risk of extinction and consequently have a poor recovery capability (Dye *et al.* 1994; Roberts and Hawkins 1999; Sadovy 2005). The correspondence analysis reveals the clustering of *H. ciliaris* and *P. paru* with the maturity criterion, due to the fact that more than a half of the sampled individuals had bred at least once. The opposite was found for *H. tricolor* and *C. striatus*, for which respectively 70% and 92% of individuals sampled were immature. Myers and Worm (2005) state that the risk of extinction is very high if recruits are being captured and low when fish are caught after first sexual maturity. Thus, the maturity criterion had a positive influence on the classification of *H. ciliaris* and *P. paru* in relation to resilience.

Large body size is often correlated with other vulnerability factors, such as late maturity, and substantially increases the possibility of being caught in traps (Reynolds and Jennings 2000; Reynolds *et al.* 2002, Cheung *et al.* 2007). Body size is a useful measure in evaluating species resilience due to its relationship with the intrinsic rate of increase of a population. Thus, it can be applied to populations and species for which minimal biological data are available (Denney *et al.* 2002). In the Pomacanthidae family, the genera *Holacanthus* and *Pomacanthus* have the largest lengths (Bellwood 2004). Among the analysed fish, *P. paru* was the second largest, but exhibited the greatest resilience. This can be explained by the fact that most individuals caught were fit for reproduction and the species had the lowest mortality rate.

The demand for marine ornamental fish increased in the 1980s and 1990s (Chan and Sadovy, 1998). Concerns have been raised regarding its negative effect on reef fish and coral habitats, as nearly all ornamental marine fish are taken from the wild (Andrew 1990; Wood 2001a). The potential for over-exploitation from the aquarium trade is high due to the high selectivity of this activity and the large number of fish collected (Wood 2001a). It is estimated that most commercialised ornamental fish are juveniles and less than 10 cm in standard length (Chan and Sadovy, 1998). Pomacanthidae and

335 Chaetodontidae are often the most important families for export in terms of number and value (Wood 2001a). In the Brazilian aquarium trade, these two families are among the most representative in species richness (Gasparini *et al.* 2005). In the state of Ceará (one of the main centres of the Brazilian ornamental fish trade), *H. ciliaris*, *P. paru*, *Hippocampus erectus*, *Pomacanthus arcuatus* and *Holacanthus tricolor* made up 50%  
340 of the total trade between 1995 and 2000 (Monteiro Neto *et al.* 2003). Among the species evaluated, *H. ciliaris* is the most harvested by the Brazilian aquarium trade (IBAMA - Federal Act 56/04). The correspondence analysis corroborates this statement, revealing a negative relationship between *H. ciliaris* and removal rate.

Regarding reproductive strategy, all species evaluated are broadcast spawners  
345 (Thresher 1980; 1984). Therefore, they are less vulnerable to extinction due to their widespread distribution and the high re-colonisation potential of depleted populations (Malakoff 1997; McKinney 1997). However, larval behaviour, spawning events and local oceanography processes can influence larval dispersion by increasing local retention (Cowen *et al.* 2006; Rocha *et al.* 2007). Hawkins *et al.* (2000) state that,  
350 despite reef fish being broadcast spawners, their geographic distribution is small; 24% are restricted to less than 800,000 km<sup>2</sup> and 9% are restricted to less than 50,000 km<sup>2</sup>. Based on these statements, it is possible to infer that being a broadcast spawner does not ensure low vulnerability to extinction.

Hermaphroditic species are likely to be very sensitive to over-fishing, as  
355 individuals need to attain a certain length for the gender change to occur (Francis 1992). If fishing affects the growth of the fish such that sexual inversion does not occur, this could result in a non-reproductive population and even cause the species to disappear in intensely exploited areas (Hawkins and Roberts 2003). Among the species analysed in the present study, those from the *Holacanthus* genus are protogynous hermaphrodites  
360 (Thresher 1980; 1984; Allen 1998; Moyer *et al.* 2003). These species are heavily exploited by the aquarium trade, in which most individuals collected are small in size (Chan and Sadovy 1998) and probably belong to the same sex (i.e., female). *H. tricolor* and *H. ciliaris* revealed the lowest recovery capacity. However, according to the correspondence analysis, the hermaphroditism criterion had no influence over their  
365 classification regarding resilience. This may be explained by the absence of protandric

species in the analysis, as such species are more sensitive than protogynous species (Sadovy and Shapiro 1987; Hawkins and Roberts 2003).

Throughout the life cycle, individuals are exposed to different types of environmental pressure. The natural mortality rate decreases as the fish grows and becomes less vulnerable to predators. The mortality coefficient used in the present study considered data on maximum and mean catch length (Beverton and Holt 1956). *H. ciliaris* had the highest mortality index among the species analysed due to the mean catch size, which was below that found in the literature (Allen 1998). This coefficient – along with the criterion related to the removal rate – had a negative influence on the resilience of this species.

Resilience ranks were different for the four species analysed, even for those belonging to the same family. Stobutzki *et al.* (2001) state that the catch of a species is not sustainable if it has both high resilience and susceptibility. The results obtained for *C. striatus* and *P. paru* are in agree with this statement, as these species revealed considerable resilience, but were highly susceptible according to the ecological information (such as diet, preferred habitat and depth range). This suggests that catching these species is not a sustainable practice.

The method employed in the present study (susceptibility and resilience criteria) is efficient for evaluating the impact of recently-formed fisheries with few available data that occur in areas with high species richness (Dulvy *et al.* 2003, 2004), such as reef environments. However, in order to incorporate the results of this study to fishery management measures, the vulnerability analysis (VA) can be utilised as a component of a risk analysis when only qualitative data is available and researchers lack an understanding of cause/effect relationships (Walker *et al.* 2002; McFadden 2007).

Most fishing equipment is not selective and many fish are caught as bycatch even when there is only one target species. These species are highly affected by fishery efforts, as bycatch will continue to be fished as long as it remains viable to catch the target species. Trap fisheries targeting reef fish have recently developed in Brazil and are on the rise in the north-eastern region. Regulatory agencies often take a long time to react and severe declines in non-target species go unperceived. Thus, little is known regarding the impact of this activity and such lack of information hinders regulatory efforts. The method presented here is an appropriate, practical tool for fishery

management and the assessment of catch sustainability in multi-species fisheries on which there is limited available data. It can also be employed to determine which species are at a possible risk of extinction due to bycatch. Furthermore, it is important to consider that to reduce the extinction threat posed on those species, a socio-economic vulnerability analysis is required in order to develop the appropriate institutional framework/outreach program to the fishers necessary to address this ecological challenge.

#### Acknowledgements

We would like to thank Sérgio Mattos, Simone Teixeira and Paulo Travassos for helpful comments on the first draft of the paper. We also thank the anonymous reviewers for constructive suggestions on the manuscript. This study was sponsored by the Brazilian Scientific Council - Conselho Nacional de Pesquisa e Desenvolvimento (CNPq).

#### References

- Allen, G. R. (1998). Introducing angelfish. In 'A Guide to Angelfish and Butterflyfish', pp. 8 – 12. (Mergus Publishers:Melle, Germany).
- Andrew, C. (1990). The ornamental fish trade and conservation. *Journal of Fish Biology* **37**, 53 – 59.
- Araújo, M. E. and Albuquerque Filho, A. C. (2005). Biologia das principais espécies de peixes ornamentais marinhos do Brasil: uma revisão bibliográfica e documental. *Boletim Técnico Científico do CEPENE* **13**, 109 – 154.
- Bellwood, D. R., van Herwerden, L. and Konow, N. (2004). Evolution and biogeography of marine angelfish (Pisces: Pomacanthidae). *Molecular Phylogenetics and Evolution* **33**, 140 – 155.
- Beverton, R. J. H. and Holt, S. J. (1956). A review of methods for estimating mortality rates in exploited fish populations, with special reference to sources of bias in catch sampling. *Rapp. P. -v. Réun. CIEM* **140**, 67 – 83.
- Burns, K. M. and Restrepo, V. (2002). Survival of reef fish after rapid depressurization: field and laboratory studies. *American Fisheries Society Symposium* **30**, 148 – 151.
- Carvalho-Filho, A. (1999). As espécies e suas famílias. In: *Peixes: Costa Brasileira*, pp. 40 – 283. (Editora Melro: São Paulo, SP).

- 430 Casey, J. M. and Myers, R. A. (1998). Near extinction of a large, widely distributed fish. *Science* **281**, 690 -691.
- Cervigón, F. (1993). Pomacanthidae. In 'Los Peces Marinos de Venezuela', pp. 339 – 346. (Fundación Científica Los Roques : Caracas, Venezuela).
- Chan, T. T. C. and Sadovy, Y. (1998). Profile of the marine aquarium fish trade in Hong Kong. *Aquarium Sciences and Conservation* **2**, 197 – 213.
- 435 Cheung, W. W. L., Watson, R., Morato, T, Pitcher, T. J. and Pauly, D. (2007). Intrinsic vulnerability in the global fish catch. *Marine Ecology Progress Series* **333**, 1 – 12.
- Cowen, R. K., Paris, C. B. and Srinivasan, A. (2006). Scaling of connectivity in marine populations. *Science* **311**, 522–527
- 440 Dalzell, P., Adams, T. J. H. and Polunin, N. V. C. (1996) Coastal fisheries in the Pacific Islands. *Oceanography and Marine Biology: an annual review* **34**, 395-531.
- Denney, N. H., Jennings, S. and Reynolds, J. D. (2002). Life-history correlates of maximum population growth rates in marine fish. *Proceedings of Royal Society London* **269**, 2229 – 2237.
- 445 Dulvy, N. K., Ellis, J. R., Goodwin, N. B., Grant, A., Reynolds, J. D. and Jennings, S. (2004). Methods of assessing extinction risk in marine fish. *Fish and Fisheries* **5**, 255 – 276.
- Dulvy, N. K., Sadovy, Y. and Reynolds, J. D. (2003). Extinction vulnerability in marine populations. *Fish and Fisheries* **4**, 25 – 64.
- 450 Dye, A. H., Schleyer, M. H., Lambert, G. and Lasiak, T. A. (1994). Biological option for the management of the exploitation of intertidal and subtidal resources. In 'Rocky Shores: Exploitation in Chile and South Africa'. (Ed.W.R. Siegfried) pp. 131 – 154. (Springer-Verlag).
- Eayrs, S. (2007). Guía para Reducir la Captura de Fauna Incidental (Bycatch) en las Pesquerías por Arrastre de Camarón Tropical. FAO, Rome.
- 455 Francis, R. C. (1992). Sexual lability in teleosts: developmental factors. *Quarterly Review of Biology* **67**, 1-18.
- Froese, R. and Pauly, D. (2007). FishBase. Available at [http:// www.fishbase.org](http://www.fishbase.org). [Accessed 29 October 2007]

- 460 Garrison, V. H., Rogers, C. S. and Beets, J. (1998). Of reef fish, overfishing and *in situ* observations of fish traps in St. John, U. S. Virgin Islands. *Revista de Biologia Tropical* **46**, 41 – 59.
- Gasparini, J. L., Floeter, S. R., Ferreira, C. E. L. and Sazima, I. (2005). Marine ornamental trade in Brazil. *Biodiversity and Conservation* **14**, 2883 – 2899.
- 465 Hawkins, J. P. and Roberts, C. M. (2003). Effects of fishing on sex-changing Caribbean parrotfish. *Biological Conservation* **115**, 213-226.
- Hawkins, J. P., Roberts, C. M. and Clark, V. (2000). The threatened status of restricted-range coral reef fish species. *Animal Conservation* **3**, 81-88.
- Hourigan, T. F., Stanton, F. G., Motta, P. J., Kelley, C. D. and Carlson, B. (1989). The  
470 feeding ecology of three species of Caribbean angelfish (family Pomacanthidae). *Environmental Biology of Fish* **24**, 105 – 116.
- Hutchings, J. A. (2001). Conservation biology of marine fish: perceptions and caveats regarding assignment of extinction risk. *Canadian Journal of Aquatic Sciences* **58**, 108 – 121.
- 475 Kelleher, K. (2005). Discards in the world's marine fisheries. An update. *FAO Fisheries Technical Paper*. N° **470**. FAO: Rome. 131p.
- Lieske, E. and Myers, R. (1994). Spadefish and Butterflyfish, In 'Collins Pocket Guide. Coral reef fish. Indo-Pacific and Caribbean including the Red Sea'. pp. 161 – 162 (Princeton University Press: Princeton, NJ).
- 480 Mahon, R. and Hunte, W. (2001). Trap mesh selectivity and the management of reef fish. *Fish and Fisheries* **2001**, 356 – 375.
- Malakoff, D. (1997). Extinction on the high seas. *Science* **277**, 486 – 488.
- McFadden, L. (2007). Vulnerability analysis in environmental management: widening and deepening its approach. *Environmental Conservation* **34**, 195 – 204.
- 485 McKinney, M. L. (1997). Extinction vulnerability and selectivity: combining ecological and paleontological views. *Annual Review of Ecology and Systematics* **28**, 495 – 516.
- Michael, S. W. (2004). Family Pomacanthidae/Angelfish. In 'Angelfish and Butterflyfish' pp. 210 – 321 (T. F. H Publications: Neptune City).

- 490 Milton, D. A. (2001) Assessing the susceptibility to fishing of populations of rare trawl  
bycatch: sea snakes caught by Australia's Northern Prawn Fishery. *Biological  
Conservation* **101**, 281 – 290.
- Monteiro Neto, C., Cunha, F. E. A., Nottingham, M. C, Araújo, M. E, Rosa, I. L. and  
Barros, G. M. L. (2003). Analysis of the marine ornamental fish trade at Ceará  
495 State, northeast Brazil. *Biodiversity and Conservation* **12**, 1287 – 1295.
- Moyer, J. T., Thresher, R. E. and Colin, P. L. (1983). Courtship, spawning and inferred  
social organization of American angelfish (genera *Pomacanthus*, *Holacanthus* and  
*Centropyge*: Pomacanthidae). *Environmental Biology of Fish* **9**, 25-39.
- Myers, R. A. and Worm, B. (2005). Extinction, survival or recovery of large predatory  
500 fish. *Philosophical Transactions of the Royal Society* **360**, 13 – 20.
- Nottingham, M. C., Cunha, F. E. and Monteiro Neto, C. (2000). Captura de peixes  
ornamentais marinhos no Ceará. *Arquivos Ciências do Mar* **33**, 113 – 118.
- Pope, J. G., MacDonald, D. S., Dann, N., Reynolds, J. D. and Jennings, S. (2000).  
Gauging the impact of fishing mortality on nontarget species. *ICES Journal of*  
505 *Marine Science* **57**, 689 – 696.
- Reynolds, J. D. and Jennings, S. (2000). The role of animal behavior in marine  
conservation. In 'Behavior and Conservation' (Eds. L. M. Gosling and W. J.  
Shuterland.) pp. 147 – 168. (Cambridge University Press: Cambridge, UK)
- Reynolds, J. D., Jennings, S. and Dulvy, N. K. (2001). Life histories of fish and  
510 population responses to exploitation exploitation. In 'Conservation of Exploited  
Species' (Eds. J. D. Reynolds, G. M. Mace, K. H. Redford and J. G. Robinson.) pp.  
147 – 168. (Cambridge University Press: Cambridge, UK.).
- Reynolds, J. D., Dulvy, N. K. and Roberts, C. M. (2002). Exploitation and other threats  
to fish conservation. In 'Fish and Fisheries Handbook' (Eds. P. J. B. Hart and J. D  
515 Reynolds.) pp. 319 – 341. (Blackwell Science: Oxford).
- Ribeiro, F. P. (2004). Composição da biocenose e abundância relativa de peixes  
capturados com covos nos Estados do Rio Grande do Norte e Pernambuco. *Boletim  
Técnico Científico do CEPENE* **12**, 113 – 128.
- Rocha, L. A., Craig, M. T. and Bowen B. W. (2007). Phylogeography and the  
520 conservation of coral reef fish. *Coral reefs* Online First.



- Roberts, C. M. and Hawkins, J. P. (1999). Extinction risk in the sea. *Trends in Ecology and Evolution* **14**, 241-246.
- Robins, C. R. and Ray, C. G. (1986). Butterflyfish: Family Chaetodonidae. In 'A field Guide to Atlantic Coast Fish of North America' pp. 192 – 194. (Houghton Mifflin Company: New York).
- 525 Sadovy, Y. and Shapiro, D. Y. (1987). Criteria for the diagnosis of hermaphroditism in fishes. *Copeia* **1**, 136 -156.
- Sadovy, Y. (2005). Trouble on the reef: the imperative for managing vulnerable and valuable fisheries. *Fish and Fisheries* **6**, 167–185.
- 530 Sparre, P. and Venema, S.C. (1992) Introduction to tropical fish stock assessment, part 1. *FAO Fisheries Technical Paper. N° 306*. FAO: Rome. 376 p.
- Starck, W. A. I. and Davis, W. P. (1966). Night habits of fish of Alligator Reef, Florida. *Ichthyologica* **38**, 313 – 356.
- Stobutzki, I., Blaber, S., Brewer, D., Fry, G., Heales, D., *et al.* (2000). Ecological sustainability of bycatch and biodiversity in prawn trawl fisheries. Fisheries Research and Development Corporation, Final Report Project 96/257, Australia.
- 535 Swingle, W. E., Dammann, A. E. and Yntema, A. N. (1970). Survey of the commercial fishery of the Virgin Islands of the United States. *Proceedings of the Gulf and Caribbean Fisheries Institute* **20**, 110 -121.
- 540 Stobutzki, I., Miller, M. and Brewer, D. (2001). Sustainability of fishery bycatch: a process for assessing highly diverse and numerous bycatch. *Environmental Conservation* **28**, 167-181.
- Thresher, R. E. (1980). Angelfish. In 'Reef fish: Behavior and Ecology on the Reef and in the Aquarium' pp 37 – 46 (The Palmetto Publishing Company: St Petersburg, Fl)
- 545 Thresher, R. E. (1984). Angelfish (Pomacanthidae). In 'Reproduction in Reef Fish' pp. 244 – 261. (T. F. H. Publications: Neptune City).
- Walker, B., Carpenter, S., Anderies, J., Abel, N., Cumming, G., Janssen, M., Lebel, L., Norberg, J., Peterson, G. D. and Pritchard, R. (2002). Resilience management in social-ecological systems: a working hypothesis for a participatory approach.
- 550 *Conservation Ecology* **6**:14.

Wolff, N., Grober-Dunsmore, R., Rogers, C. S., and Beets, J. (1999). Management implications of fish trap effectiveness in adjacent coral reef and gorgonian habitats. *Environmental Biology of Fish* **55**, 81-90.

555 Wood, E. M. (2001a). Collection of coral reef fish for aquaria: global trade, conservation issues and management strategies. Marine Conservation Society, Ross-on-Wye, UK.

Wood, E. M. (2001b). Global Advances in Conservation and Management of Marine Ornamental Resources. *Aquarium Science and Conservation* **3**, 65–77.

### **Considerações finais**

O conhecimento do ciclo de vida de espécies como *Pomacanthus paru*, que são extraídas diretamente dos recifes para o aquarismo, é fundamental para a sua preservação. Desde o ano 2000, esta espécie vem sendo classificada como a segunda mais exportada pelo Brasil, principalmente pelo Estado do Ceará. Nas últimas reuniões técnicas sobre a exploração de peixes ornamentais marinhos, promovidas pelo IBAMA, foi enfatizada a carência de pesquisas sobre a biologia e a dinâmica populacional das principais espécies exportadas (e.g. *Holacanthus ciliaris* e *Pomacanthus paru*).

As considerações aqui apresentadas sobre a diversidade genética dos peixes anjo representam um primeiro passo para se compreender as estruturas populacionais de espécies irmãs e simpátricas, como *P. paru* e *P. arcuatus*, embora com histórias evolutivas diferentes. Estudos complementares sobre assentamento larval, características fisiológicas e ações antrópicas dirigidas a essas populações são necessários para inferir sobre essa dinâmica.

As pesquisas relacionadas à biologia reprodutiva indicaram que *P. paru* é gonocorística e os dados de reprodução, idade e crescimento forneceram informações importantes sobre sua biologia populacional. Algumas hipóteses, como a preferência de habitat entre os sexos, uma maior susceptibilidade de captura das fêmeas por meio das armadilhas e a migração das fêmeas no período reprodutivo para a área de pesca, podem explicar a prevalência de fêmeas na estrutura populacional de *Pomacanthus paru*. Entretanto, mais pesquisas serão necessárias para responder esta questão.

Os resultados mostraram também que é possível obter estimativas de crescimento para espécies tropicais baseadas em métodos convencionais para determinação de idade. Os dados obtidos nos trabalhos de biologia reprodutiva e idade e crescimento deverão ser utilizados no manejo de *P. paru*. Algumas medidas de manejo, relacionadas ao aquarismo, já são aplicadas em *P. paru*. Nos Estados Unidos, o ordenamento da pesca desta espécie para fins ornamentais é realizado por meio da regulamentação de tamanho. No Brasil, o ordenamento é

feito com cotas de exportação específicas. Estas regulamentações são fundamentais e ajudam na conservação dos estoques e na prevenção contra a sobreexploração. Entretanto, essas medidas foram tomadas sem nenhum conhecimento dos parâmetros populacionais da espécie, como crescimento, longevidade, idade e tamanho de primeira maturação sexual. Esses parâmetros são requisitos mínimos para um manejo pesqueiro efetivo de uma determinada espécie. Diante do exposto, o manejo de *P. paru* deve incluir medidas baseadas nos parâmetros populacionais aqui estabelecidos, antes que mais restrições sejam impostas. Entretanto, esses parâmetros aqui estabelecidos não se adequam ao manejo das espécies utilizadas no aquarismo, pois a maioria dos indivíduos comercializados é juvenil e inferior a 10 cm de comprimento padrão. Desta forma, sugere-se que as medidas conservacionistas estejam relacionadas a dados quantitativos (captura) por área e criação de áreas de proteção ambiental.

Somada à pressão pesqueira sofrida por esta espécie através do aquarismo, *P. paru* é constantemente capturada como *bycatch* em armadilhas de pesca no nordeste do Brasil. Nesta pescaria, mais de 70% dos indivíduos capturados estavam aptos para a reprodução. É possível que essa espécie sofra pressão pesqueira em todas as faixas de comprimento, uma vez que indivíduos menores que 10 cm são comumente utilizados no aquarismo e aqueles com comprimento entre 15 e 40 cm são frequentemente pescados em armadilhas. A constante captura de indivíduos de maior porte representa um possível risco ao recrutamento para o estoque reprodutivo.

No caso da pesca com armadilhas, não há nenhum tipo de regulamentação para esta atividade. O manejo desta atividade é difícil, por ser considerada multi-específica e com impacto desconhecido. A maioria das artes de pesca não é seletiva e muitos peixes são capturados como *bycatch*, mesmo quando há apenas uma espécie alvo. Esses peixes são altamente afetados, pois enquanto for viável capturar a espécie alvo, o *bycatch* continuará a ser pescado.

A metodologia estabelecida nesta tese é uma ferramenta prática e apropriada para o manejo pesqueiro e avaliação da sustentabilidade da pesca de espécies que são capturados em pescarias multi-específicas e com poucos dados disponíveis sobre o impacto da atividade. Este método também pode ser empregado para determinar quais as espécies estão em um possível risco de extinção devido ao *bycatch*. Além disso, faz-se necessária uma análise de vulnerabilidade sócio-econômica a fim de desenvolver um programa institucional apropriado para os pescadores, no intuito de resolver essa problemática ecológica.

## APÊNDICE

# A rapid new method for assessing sustainability of ornamental fish by-catch from coral reefs

Caroline Vieira Feitosa<sup>A,B</sup>, Beatrice Padovani Ferreira<sup>A</sup>  
and Maria Elisabeth de Araújo<sup>A</sup>

<sup>A</sup>Departamento de Oceanografia, Centro de Tecnologia e Geociências, Universidade Federal de Pernambuco, 50670-901, Recife, Pernambuco, Brazil.

<sup>B</sup>Corresponding author. Email: carol\_feitosa@hotmail.com

**Abstract.** The ornamental fish trade has considerable potential for damaging reef environments. Nearly 120 species are currently harvested in the Brazilian trade and little attention has been paid to the effects of the accidental catch of these species. Records of ornamental fish by-catch and the susceptibility and resilience of species from the major Brazilian families in the aquarium trade were examined to test a method and determine the sustainability of these catches. Each species was assessed based on two groups of criteria: (1) relative vulnerability to capture; and (2) population resilience to mortality due to fishing efforts. Each species was also ranked (1–3) for criteria regarding ecology, biology, catchability, numbers of individuals exported by aquarium companies and mortality index. These criteria were then weighted and each species rank in the two groups of criteria was plotted. Nineteen species from 10 families were recorded. The species least likely to have sustainable catch rates included *Holacanthus tricolor*, *H. ciliaris*, *Chaetodon striatus* and *Pomacanthus paru*. This approach is a useful alternative for management efforts and the assessment of catch sustainability in newly developed multi-species fisheries with limited available data, which occur in areas of high species richness such as reefs.

**Additional keywords:** aquarium, Chaetodontidae, fishery, Pomacanthidae, resilience, susceptibility, trap.

## Introduction

Species classified as by-catch (Eayrs 2007) are caught in fisheries that target other groups and are then either used or discarded. It is estimated that 7.3 million tonnes of fish were discarded globally in commercial fisheries between 1995 and 2005 (Kelleher 2005). This is equivalent to 8% of worldwide marine fish production (Eayrs 2007). The global effect of fisheries on by-catch species is poorly known, although some studies have documented ecological changes at the species level (Casey and Myers 1998; Pope *et al.* 2000). Even less attention has been paid to accidental catches affecting reef fish, including ornamental species.

Beauty and perfect patterns on ornamental reef fish are requirements imposed by the international aquarium trade (Araújo and Albuquerque Filho 2005), which is one of the world's largest industries. The import value worldwide of marine fish and invertebrates is estimated to be between US\$24 and 40 million annually (Wood 2001a). Nearly 120 species are currently harvested in the Brazilian aquarium trade (Monteiro Neto *et al.* 2003). Most belong to the family Pomacanthidae (Araújo and Albuquerque Filho 2005) and species of the Chaetodontidae are among the most exported (Monteiro Neto *et al.* 2003; Nottingham *et al.* 2000). Although other fisheries that use traps have an effect on ornamental marine fish stocks (through by-catch), these stocks have only been managed through a Federal Act (56/04) established in 2004 by the Brazilian environmental protection agency (Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis – IBAMA), which monitors

the ornamental fish trade through export quotas. The purpose of this Federal Act is to administer this activity through the establishment of export quotas.

Traps are the predominant gear employed to capture reef fish in the Caribbean (Swingle *et al.* 1970; Garrison *et al.* 1998) as well as other places in the world (e.g. Arabian Gulf, Seychelles, Papua New Guinea) (Dalzell *et al.* 1996) and are responsible for the decline of many reef species which are fishery targets (Wolff *et al.* 1999). Among the organisms caught by these devices, the most frequently caught ornamental fish belong to the following genera: *Acanthurus*, *Chaetodon*, *Bodianus*, *Holacanthus* and *Pomacanthus* (Garrison *et al.* 1998; Wolff *et al.* 1999). In Brazil, trap fisheries targeting reef fish have been on the rise in the last 10 years (Ribeiro 2004). The target species of these fisheries belong to the genera *Acanthurus*, *Cephalopholis*, *Lutjanus*, *Pseudupeneus* and *Sparisoma* (Ribeiro 2004). Because by-catch species, including ornamental species, are returned to the sea and not recorded, there are no data or regulations available about the actual catch of ornamental species.

Owing to the increasing pressure to manage the impact of fisheries on non-target species and the uncertainty of how this is to be done, Stobutzki *et al.* (2001) developed a broad method using qualitative measures of the biological and ecological characteristics of species to rank vulnerability to a known threat. Stobutzki *et al.* (2001) examined the impact of trawling on the catch sustainability of teleost by-catch species, identifying the species with the least likelihood of catch sustainability in by-catch. They argued that such species should then be the focus

of research and management. Stobutzki *et al.* (2001) recorded at least 411 species from 99 families as by-catch, identifying 15 species with the least likelihood of catch sustainability as well as the 13 with the most likelihood of catch sustainability. This approach was designed to overcome problems such as the high diversity of by-catch in tropical fisheries and the lack of information on individual species. The method is versatile and can also be applied to other fisheries, although the criteria employed may vary.

The present study describes a rapid, innovative method, based on the method developed by Stobutzki *et al.* (2001), for evaluating the impact of trap fisheries on ornamental species using biological data and the portion of the population caught. The intention is to answer the following two questions. (1) Is the method appropriate for ornamental reef fish? (2) Is the capture of ornamental reef fish as by-catch sustainable? The present study records the occurrence of ornamental fish caught as by-catch and assesses the susceptibility and resilience of fish from the Pomacanthidae and Chaetodontidae families.

## Materials and methods

### Sampling

The fish were acquired from the fishing colony of Itamaracá island located on Pilar Beach (7°45'17.80"S, 34°49'26.46"W) (Itamaracá, Pernambuco, Brazil – Fig. 1) and were caught as by-catch while trap fishing for the spotted goatfish, *Pseudupeneus maculatus* and parrotfish, *Sparisoma* spp. This kind of fishing occurs throughout the year and is carried out on alternating days. The boats leave Pilar Beach at midnight and arrive at Tambaba Beach, 46 km away (7°22'2.93"S, 34°47'48.51"W) (Paraíba, Brazil) in the early morning. Four boats are employed in this fishery. Each has a fishing effort of 15 to 20 traps and trap mesh varying between 1 and 7 cm. The traps are deployed unbaited next to reefs and hauled 6 h later. Occasionally, depending on factors such as wind and currents, some traps are left for longer periods and only hauled after 48 h. The fish are stored for ~7 h in 100-L containers. For the present study, weekly samplings were performed from April to June 2006. The fishermen were asked to bring back all specimens with ornamental characteristics, which were then bought. In the case of *Holacanthus ciliaris* and *Pomacanthus paru*, this procedure was carried out over a longer period (March 2006 to February 2007), as these two species were the focus of a concurrent study.

The sampled fish were placed in polystyrene chests with ice and taken to the Nekton and Aquaculture Laboratory of the Oceanography Department of the Universidade Federal de Pernambuco, where they were identified, measured and photographed.

### Analysis of catch sustainability of ornamental reef fish by-catch

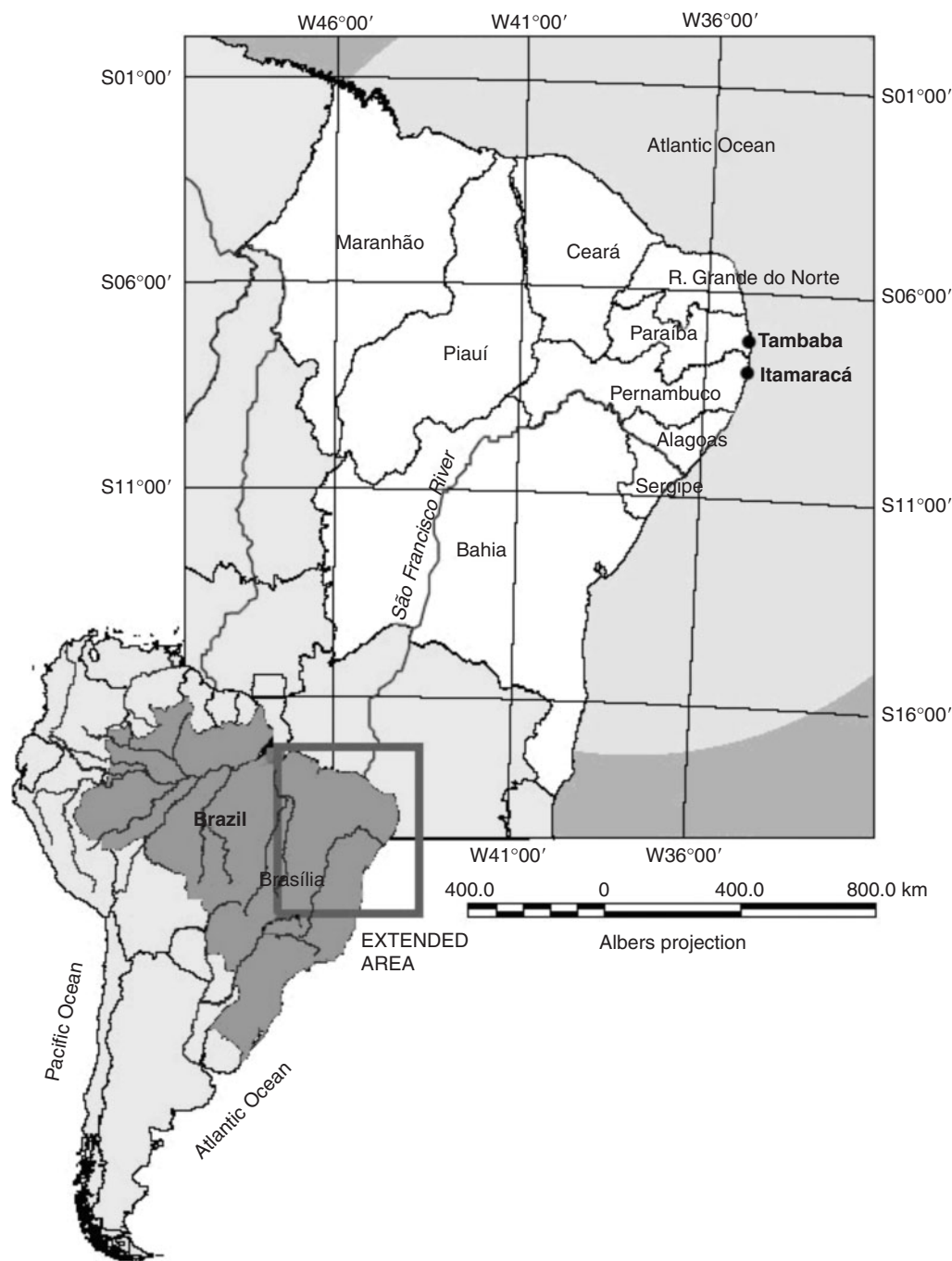
Only the most abundant species were analysed, i.e. species from the Pomacanthidae and Chaetodontidae. Rare species for Brazil, such as *Chaetodon ocellatus* and *Pomacanthus arcuatus* (Thresher 1980; Carvalho Filho 1999) were not surveyed owing to the small sample size. Information on the biological characteristics of the selected species was taken from published literature including Robins and Ray (1986), Thresher (1980,

1984), Allen (1998), Hourigan *et al.* (1989) and Froese and Pauly (2007).

The method proposed to assess the sustainability of selected by-catch species was adapted from Stobutzki *et al.* (2001) and applied based on two scored sets of criteria: (1) relative vulnerability to capture by traps and mortality due to trap fishery; and (2) the recovery capacity of a species after the population is depleted. The criteria were originally developed during a study on the sustainability of all North Australian Prawn Fishery (NPF) prawn-trawl vertebrate by-catch (Stobutzki *et al.* 2000). These criteria were modified and assigned relative importance weights by the NPF Assessment Group (Milton 2001). Values ranging from 1 to 3 were attributed to each sustainability indicator, for which 1 denotes the highest value for the susceptibility criteria and the lowest for resilience. Depending on the criterion, these values were based on either categorical or continuous data. When no information was available to assign the divisions between values, the range of the data was divided into thirds to create categories (Table 1). The rank matrix constructed from these values for each set of criteria are plotted as axes (resilience versus susceptibility) on a graph indicating the species with the greatest catch sustainability. This graph is recommended only if the different species have different values on each axis. According to Stobutzki *et al.* (2001), when species-specific information is unavailable, a species is given the same value as other species within the same family. With criteria for which family members are not similar or for which no family information is available, the rank of 1 is given as a precautionary measure.

The criteria adopted to analyse species susceptibility (axis 1) to capture and possible mortality due to fishery efforts were adapted from Stobutzki *et al.* (2001) and are summarised as follows. (1.1) Water column position – as traps are deployed on the substrate (Mahon and Hunte 2001; Garrison *et al.* 2004), benthic and demersal species are more susceptible to capture than pelagic species. (1.2) Preferred habitat – species are ranked according to their preference for rocky habitats. This criterion addresses the likelihood that a species habitat overlaps the area in which a trap fishery occurs. Coral reef habitats have been historically targeted by trap fisheries (Wolff *et al.* 1999). Thus, species that live near shore and depend on a specific habitat, such as coral reefs, are highly vulnerable to a possible risk of extinction (Dye *et al.* 1994; Roberts and Hawkins 1999). (1.3) Day/night catchability – because trap fishery is a daytime operation, species with diurnal habits are more susceptible to capture by traps. (1.4) Diet – whether eating habits attract species to trap fishery areas. Thus, species that generally feed on benthic or demersal organisms found on reefs are more vulnerable to capture. Invertebrate feeders (mobile and sessile) are the dominant trophic guild in traps deployed in coral reef habitats (Garrison *et al.* 2004). (1.5) Depth range – the trap fishery that we studied occurs in depths between 30 and 40 m. According to Wolff *et al.* (1999), trap fisheries in adjacent coral reef and gorgonian habitats range from 3 to 38 m. Overlap between the depth range of the trap fishery area and that of the preferred habitat will affect the vulnerability of a species to capture more than that of others that live in either deeper or shallower areas. Fish that have a narrow vertical depth range are highly vulnerable to extinction (Dye *et al.* 1994; Roberts and Hawkins 1999) (Table 1).





**Fig. 1.** Location of sampled area.

Six criteria adapted from Stobutzki *et al.* (2001) were employed to analyse species resilience. (2.1) Maturity – if most individuals caught are larger than their size at first maturity, the species has a greater probability of recovering from a declining population. (2.2) Maximum size – species that attain large sizes tend to live longer and therefore the resilience of the population is lower than species with smaller body sizes. A slow life history (e.g. late maturity, long reproductive span and large body size) is related to low maximum population growth

rates and, consequently, such species are more vulnerable and have less capacity to recover from exploitation (Hutchings 2001; Reynolds *et al.* 2001, 2002). (2.3) Removal rate – species-based quotas established by IBAMA to ensure that harvests are maintained at a sustainable level. The ornamental species fishery is highly selective and certain species, such as chaetodontids and pomacanthids, are preferred targets. Among those, the species that are rare may be vulnerable to over-exploitation (Wood 2001b). (2.4) Reproductive strategy – species capable

**Table 1. The criteria used to assess the susceptibility of by-catch species to capture and mortality due to traps and their recovery**  
For each criterion the weighting score and definition of the three ranks is given

Criteria	Weighting score	1	2	3
Susceptibility				
Water column position	3	Demersal or benthic	Benthopelagic	Pelagic
Preferred habitat	3	Rocky habitats with gravel and sand	Habitats with soft or muddy bottoms	No fishing habitats
Day/night catchability	2	Higher catch rate at day	No difference between day and night	Higher catch rate at night
Diet	3	Feed on demersal or bent. organisms	Feed on benthopelagic organisms	Feed on pelagic organisms
Depth range	1	Less than 40 m	–	Deeper than 40 m
Resilience				
Maturity	3	Mature individuals <50%		Mature individuals >50%
Maximum size	3	Max. size $\geq 50$ cm	16 cm < max. size < 50 cm	Max. size $\leq 16$ cm
Removal rate	3	Catch $\geq 3500$ inds/comp/yr	1000 < catch < 3500 inds/comp/yr	Catch $\leq 1000$ inds/comp/yr
Reproductive strategy	2	Bear live young or brood young	Guard eggs and/or young	Broadcast spawners
Hermaphroditism	1	Protandric hermaphrodites	Protogynous hermaphrodites	Dioecious
Mortality index	1	Mortality index $\geq 3.19$	0.96 < mortality index < 3.19	Mortality index $\leq 0.96$

of dispersal over long distances are less vulnerable to extinction (Dye *et al.* 1994; Roberts and Hawkins 1999). Thus, the resilience of R-strategist species is generally higher than that of K-strategist species. (2.5) Hermaphroditism – hermaphroditic species, especially protandric species, may be at great risk of extinction because fisheries select individuals of certain sizes and hermaphroditic fish generally only change sex when they reach a certain size, so the fishing effort is, in effect, directed at only one sex – which can have a negative effect on reproduction success (Dye *et al.* 1994; Roberts and Hawkins 1999; Hawkins and Roberts 2004). (2.6) Mortality index – this index was proposed by Sparre and Venema (1992) and Stobutzki *et al.* (2001). This index can be derived from the length–frequency of a species and von Bertalanffy growth parameters. Such parameters are not available for most by-catch species, but the index can be calculated as follows:

$$\text{Mortality index (Z): } (L_{\max} - L_{\text{mean}})/(L_{\text{mean}} - L_{\min})$$

in which  $L_{\max}$  is the maximum length recorded in the literature,  $L_{\text{mean}}$  is the mean length of total number of individuals caught and  $L_{\min}$  is the minimum length of the individuals in the sample. In the present study, the mortality index range was calculated (0.13–0.74) and divided into thirds for the division between ranks (Table 1).

#### Analysis of criteria

The overall ranks of susceptibility or resilience were calculated according to the equation below:

$$S_i = \sum_{j=1}^n R_i / \sum_{j=1}^n w_j$$

in which  $S_i$  is the total susceptibility or resilience rank for species  $i$ ,  $w_j$  is the weight for criterion  $j$ ;  $R_i$  is the rank of species  $i$  for criterion  $j$  and  $n$  is the number of criteria for each axis.

#### Statistical analysis

Data normality was evaluated using the Kolmogorov–Smirnov test (BioEstat 5.0 software – Instituto de Desenvolvimento

Sustentável Mamirauá – Amazonas). The parametric Z-test (BioEstat 5.0 software) was used to determine the probability of a captured individual being smaller than size at first maturity and the probability of a captured individual having bred at least once. Correspondence analysis (Multivariate Statistical Package – MVSP 3.1 software – Kovach Computing Services – Anglesey, Wales) was used to determine and describe the correspondence between species and criteria on the same graph. This multivariate analysis was used only for criteria related to resilience because values regarding susceptibility did not vary among species. Since the variance of the reproductive strategy criterion was equal to zero, this criterion was not applied in the present analysis.

#### Results

Among the fish caught, 19 were recorded as by-catch species and belonged to 14 genera and 10 families (Table 2). Of these, 100% were from reef environments and are used in the aquarium trade and only one species (*Equetus lanceolatus*) was not authorised for export. In total, 430 individuals were sampled: 147 *Holacanthus ciliaris*, 46 *Holacanthus tricolor*, 200 *Pomacanthus paru* and 37 *Chaetodon striatus*. Length–frequency distribution by species (Fig. 2) revealed a broad length range among the individuals sampled.

All susceptibility criteria are species-specific and the four species evaluated in the sustainability analysis obtained the same value (1) for this criterion. Rank resilience ranged from 1.92 to 2.54; *H. tricolor* and *P. paru* obtained the smallest and the largest values respectively (Table 3). Based on these results, it can be affirmed that *H. tricolor* has the least catch sustainability, followed by *H. ciliaris* and *C. striatus*. *P. paru* is the most capable of recovering from a population decline.

Mean and smallest lengths for *H. tricolor* and *C. striatus* individuals were under size-at-first-maturity. There was a 71% probability of *H. tricolor* individuals and a 92% probability of *C. striatus* being caught at sizes smaller than first maturity. Contrarily, the probability of captured individuals being larger than maturity size was 76% for *H. ciliaris* and 70% for *P. paru*. *H. ciliaris* had the highest mortality index ( $Z = 3.19$ ) and *P. paru* had the lowest ( $Z = 0.96$ ) (Table 4).

**Table 2.** Species registered as by-catch, with the respective common names

Family	Species	Common name
Syngnathidae	<i>Hippocampus reidi</i> Ginsburg, 1933	Longsnout seahorse
Synodontidae	<i>Synodus foetens</i> (Linnaeus, 1766)	Inshore lizardfish
Aulostomidae	<i>Aulostomus maculatus</i> Valenciennes, 1841	Trumpetfish
Sciaenidae	<i>Equetus lanceolatus</i> (Linnaeus, 1758)	Jack-knife fish
Sciaenidae	<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)	Highhat
Chaetodontidae	<i>Chaetodon ocellatus</i> Bloch, 1787	Spotfin butterflyfish
Chaetodontidae	<i>Chaetodon striatus</i> Linnaeus, 1758	Banded butterflyfish
Pomacanthidae	<i>Holacanthus ciliaris</i> (Linnaeus, 1758)	Queen angelfish
Pomacanthidae	<i>Holacanthus tricolor</i> (Bloch, 1795)	Rock beauty
Pomacanthidae	<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	Gray angelfish
Pomacanthidae	<i>Pomacanthus paru</i> (Bloch, 1787)	French angelfish
Labridae	<i>Bodianus rufus</i> (Linnaeus, 1758)	Spanish hogfish
Labridae	<i>Bodianus pulchellus</i> (Poey, 1860)	Spotfin hogfish
Labridae	<i>Halichoeres dimidiatus</i> (Agassiz, 1831)	Brazilian yellowcheek wrasse
Labridae	<i>Halichoeres brasiliensis</i> (Bloch, 1791)	Brazilian wrasse
Monacanthidae	<i>Cantherhines macrocerus</i> (Hollard, 1855)	Whitespotted filefish
Monacanthidae	<i>Stephanolepis hispidus</i> (Linnaeus, 1766)	Planehead filefish
Ostraciidae	<i>Lactophrys trigonus</i> (Linnaeus, 1758)	Buffalo trunkfish
Tetraodontidae	<i>Canthigaster figueiredoi</i> Moura & Castro, 2002	Southern Atlantic sharpnose-puffer

The correspondence analysis for resilience revealed that *H. tricolor* and *C. striatus* are linked to the removal index. *H. ciliaris* and *P. paru* are associated through maturity criteria. *C. striatus* and *P. paru* are grouped with maximum size and mortality index criteria respectively. Negative relationships were determined for: *H. ciliaris* and the removal rate and mortality index; *H. tricolor* and *C. striatus* with maturity criteria; and *P. paru* with maximum size. The percentage of the cumulative variation for the two axes was 99.53% (Fig. 3).

## Discussion

The present study demonstrates that the trap fishery is not selective. The catching of ornamental fish occurs frequently and likely causes considerable impact. Ornamental species are already considered seriously endangered owing to fishery efforts (Cheung *et al.* 2007). Individuals caught are not landed and are usually returned to the sea (many still alive, according to the fishermen). Despite this, the survival of these individuals is doubtful, as the traps are hauled quickly to surface and problems related to decompression have often been reported (Burns and Restrepo 2002).

The method applied in the present study was developed with the purpose of classifying ornamental fish species according to possible risk of extinction owing to being captured as by-catch. This analysis is influenced by the length range and proportion of young and immature individuals sampled because criteria that are directly related to specimen size are employed, such as length at first maturity and mortality index. Different mesh sizes are used in this fishery, which may have contributed to the broad length range among the individuals sampled. Owing to the small sample size for *H. tricolor* and *C. striatus*, the length range of the specimens caught may not be representative. Thus, for these species, the analysis must be considered only for demonstration (exploratory) purposes, even though there was a trend towards normality in the length distribution data. The present study is the

first to test a method with such goals directed at trap fisheries and using criteria based on life history.

Specific biological information is optimised when well defined criteria are applied, including characteristics that affect species sensitivity or risk of extinction (Dye *et al.* 1994; Roberts and Hawkins 1999; Stobutzki *et al.* 2001). Stobutzki *et al.* (2001) applied the value 1 in the matrix when there was no specific information for certain criteria. In the present study, all biological information was species-specific. Although the analysis presented is subjective, this may be the only one available to evaluate and monitor multi-specific fisheries (Dulvy *et al.* 2003), especially newly developed fisheries.

## Susceptibility to capture by traps

Species caught by a given equipment share similar characteristics, such as habitat, diet and period of activity. Thus, reef fish are considered highly susceptible to trap fisheries. Species of Pomacanthidae are widely distributed in depths ranging from 1 to 100 m (Allen 1998; Michael 2004; Froese and Pauly 2007), whereas species of Chaetodontidae occur between 3 and 55 m (Robins and Ray 1986; Allen 1998; Froese and Pauly 2007). The former feed on sponges, zoanthids, tunicates, hydroids, bryozoans and algae (Randall 1967; Allen 1998; Michael 2004). Chaetodontids eat worms, coral polyps and mollusc eggs (Allen 1998). The diet and period of activity of these fish favour their capture because they only feed once during the day (Starck and Davis 1966) and the organisms they eat are sessile, which are generally encrusted on reefs. Dependence on a particularly susceptible habitat (e.g. coral reefs), either alone or combined with overfishing, leads to considerable captures of by-catch species and makes these species all the more vulnerable (Reynolds and Jennings 2000).

## Resilience

Species with late sexual maturity are highly vulnerable to the risk of extinction and consequently have a poor recovery capability

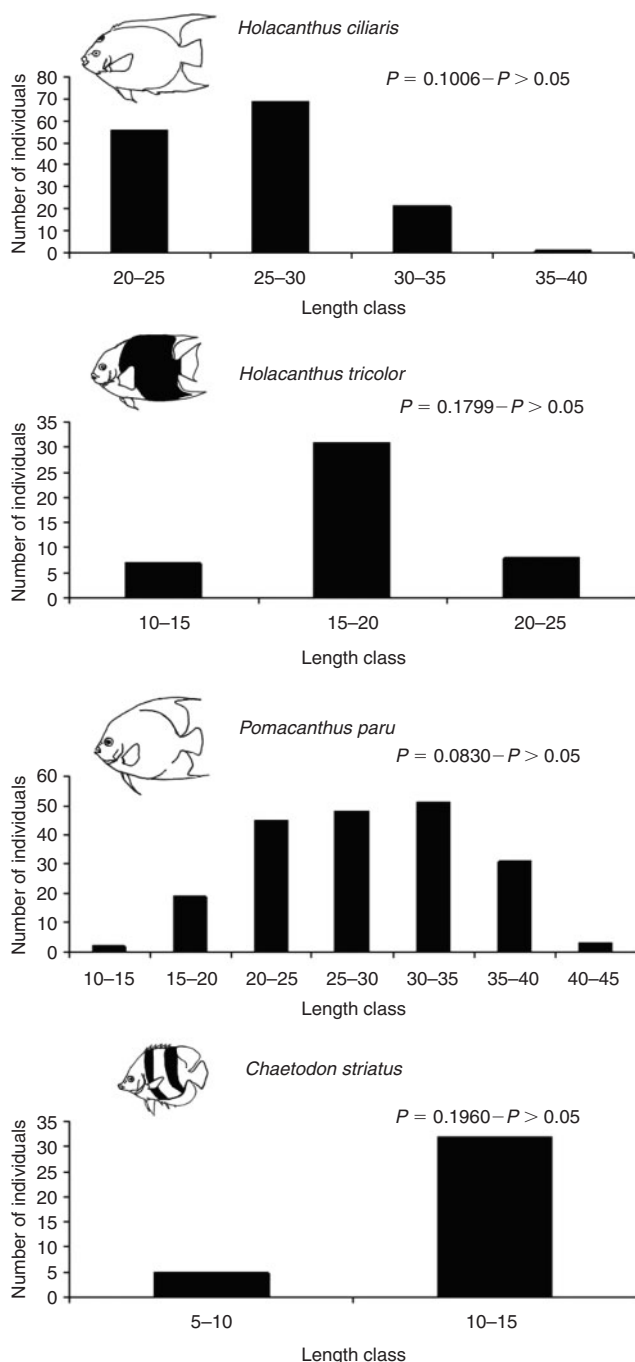


Fig. 2. Length distributions for the four analysed species.

(Dye *et al.* 1994; Roberts and Hawkins 1999; Sadovy 2005). The correspondence analysis reveals the clustering of *H. ciliaris* and *P. paru* with the maturity criterion owing to the fact that more than a half of the sampled individuals had bred at least once. The opposite was found for *H. tricolor* and *C. striatus*, for which, respectively, 70% and 92% of individuals sampled were immature. Myers and Worm (2005) stated that the risk of extinction is very high if recruits are being captured and low when fish are caught after first sexual maturity. Thus, the maturity criterion

had a positive influence on the classification of *H. ciliaris* and *P. paru* in relation to resilience.

Large body size is often correlated with other vulnerability factors, such as late maturity, and substantially increases the possibility of being caught in traps (Reynolds and Jennings 2000; Reynolds *et al.* 2002; Cheung *et al.* 2007). Body size is a useful measure in evaluating species resilience owing to its relationship with the intrinsic rate of increase of a population. Thus, it can be applied to populations and species for which minimal biological data are available (Denney *et al.* 2002). In the Pomacanthidae, the genera *Holacanthus* and *Pomacanthus* are the longest (Bellwood *et al.* 2004). Among the analysed fish, *P. paru* was the second largest, but exhibited the greatest resilience. This can be explained by the fact that most individuals caught were fit for reproduction and the species had the lowest mortality rate.

The demand for marine ornamental fish increased in the 1980s and 1990s (Chan and Sadovy 1998). Concerns have been raised regarding its negative effect on reef fish and coral habitats because nearly all ornamental marine fish are taken from the wild (Andrews 1990; Wood 2001a). The potential for over-exploitation from the aquarium trade is high owing to the high selectivity of this activity and the large number of fish collected (Wood 2001a). It is estimated that most commercialised ornamental fish are juveniles and less than 10 cm in standard length (Chan and Sadovy 1998). Species of Pomacanthidae and Chaetodontidae are often the most important families for export in terms of number and value (Wood 2001a). In the Brazilian aquarium trade, these two families are among the most representative in species richness (Gasparini *et al.* 2005). In the state of Ceará (one of the main centres of the Brazilian ornamental fish trade), *H. ciliaris*, *P. paru*, *Hippocampus erectus*, *Pomacanthus arcuatus* and *Holacanthus tricolor* made up 50% of the total trade between 1995 and 2000 (Monteiro Neto *et al.* 2003). Among the species evaluated, *H. ciliaris* is the most harvested by the Brazilian aquarium trade (IBAMA – Federal Act 56/04). The correspondence analysis corroborates this statement, revealing a negative relationship between *H. ciliaris* and removal rate.

Regarding reproductive strategy, all species evaluated are broadcast spawners (Thresher 1980, 1984). Therefore, they are less vulnerable to extinction owing to their widespread distribution and the high recolonisation potential of depleted populations (Malakoff 1997; McKinney 1997). However, larval behaviour, spawning events and local oceanography processes can influence larval dispersal by increasing local retention (Cowen *et al.* 2006; Rocha *et al.* 2007). Hawkins *et al.* (2000) stated that, despite reef fish being broadcast spawners, their geographic distribution is small: 24% are restricted to less than 800 000 km<sup>2</sup> and 9% are restricted to less than 50 000 km<sup>2</sup>. Based on these statements, it is possible to infer that being a broadcast spawner does not ensure low vulnerability to extinction.

Hermaphroditic species are likely to be very sensitive to over-fishing because individuals need to attain a certain length for the sex change to occur (Francis 1992). If fishing affects the growth of the fish such that sexual inversion does not occur, this could result in a non-reproductive population and even cause the species to disappear in intensely exploited areas (Hawkins and Roberts 2004). Among the species analysed in the present study, those from the *Holacanthus* genus are protogynous hermaphrodites (Thresher 1980, 1984; Moyer *et al.* 1983;

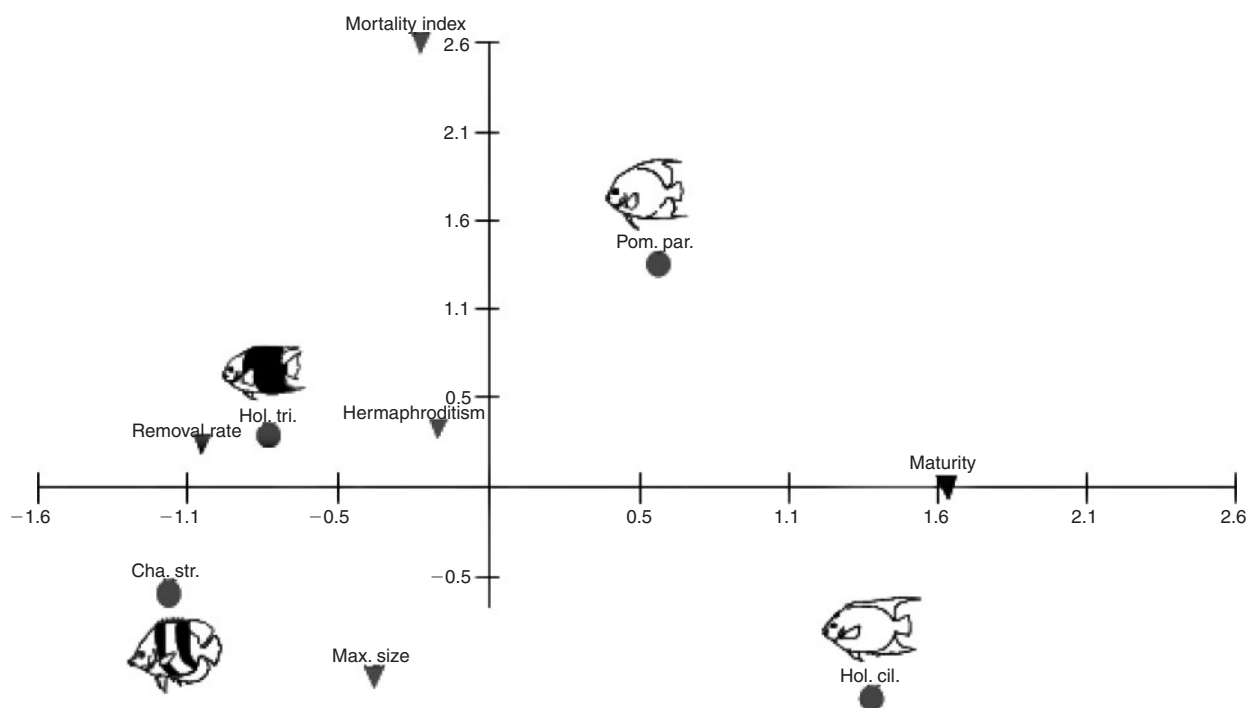
**Table 3.** The ranking of the species on the criteria on the resilience axis

Species/weight	Resilience						Rank
	3 Maturity	3 Maximum size	3 Removal rate	2 Reproductive strategy	1 Hermaphroditism	1 Mortality index	
<i>Holacanthus ciliaris</i>	3	2	1	3	2	1	2.08
<i>Holacanthus tricolor</i>	1	2	2	3	2	2	1.92
<i>Pomacanthus paru</i>	3	2	2	3	3	3	2.54
<i>Chaetodon striatus</i>	1	3	3	3	3	2	2.46

**Table 4.** The length at first capture, length at maturity, probability that individuals caught are below the length at maturity, probability of breeding, mean and maximum length (max. length) of the species and the mortality index (*Z*) of four species of fish

Species	<i>n</i>	<i>L'</i> (cm)	<i>L<sub>m</sub></i> (cm)	Prob. <i>L' &lt; L<sub>m</sub></i> (cm)	Prob. of breeding	<i>L<sub>mean</sub> ± s.e.</i> (cm)	Max. length (cm)	<i>Z</i>
<i>Holacanthus ciliaris</i>	147	20.88	24.3	<i>P</i> = 0.24	<i>P</i> = 0.76	26.58 ± 0.2672	45 (Allen 1998)	3.19
<i>Holacanthus tricolor</i>	46	11.32	19	<i>P</i> = 0.71	<i>P</i> = 0.29	17.61 ± 0.3646	35 (Allen 1998)	2.68
<i>Pomacanthus paru</i>	200	13.44	25	<i>P</i> = 0.30	<i>P</i> = 0.70	28.30 ± 0.4510	41 (Cervigón 1993)	0.96
<i>Chaetodon striatus</i>	37	9.09	13.2	<i>P</i> = 0.92	<i>P</i> = 0.07	11.26 ± 0.2257	16 (Lieske and Myers 1994)	2.17

*n* = number of individuals, *L'* = length at first capture, *L<sub>m</sub>* = length at maturity, Prob. = probability.

**Fig. 3.** Correspondence analysis showing the relationship between species and criteria used to evaluate the recovery capacity. Hol. cil. = *Holacanthus ciliaris*, Hol. tri. = *Holacanthus tricolor*, Cha. str. = *Chaetodon striatus*, Pom. par. = *Pomacanthus paru*, Max. size = maximum size.

Allen 1998). These species are heavily exploited by the aquarium trade, in which most individuals collected are small in size (Chan and Sadovy 1998) and probably belong to the same sex (i.e. female). *H. tricolor* and *H. ciliaris* revealed the lowest recovery capacity. However, according to the correspondence

analysis, the hermaphroditism criterion had no influence over their classification regarding resilience. This may be explained by the absence of protandric species in the analysis because such species are more sensitive than protogynous species (Sadovy and Shapiro 1987; Hawkins and Roberts 2004).

Throughout the life cycle, individuals are exposed to different types of environmental pressure. The natural mortality rate decreases as the fish grows and becomes less vulnerable to predators. The mortality coefficient used in the present study considered data on maximum and mean catch length (Beverton and Holt 1956). *H. ciliaris* had the highest mortality index among the species analysed owing to the mean catch size, which was below that found in the literature (Allen 1998). This coefficient – along with the criterion related to the removal rate – had a negative influence on the resilience of this species.

Resilience ranks were different for the four species analysed, even for those belonging to the same family. Stobutzki *et al.* (2001) stated that the catch of a species is not sustainable if it has both high resilience and susceptibility. The results obtained for *C. striatus* and *P. paru* are in agreement with this statement because these species revealed considerable resilience, but were highly susceptible according to the ecological information (such as diet, preferred habitat and depth range).

The method employed in the present study (susceptibility and resilience criteria) is efficient for evaluating the impact of newly formed fisheries with few available data that occur in areas with high species richness (Dulvy *et al.* 2003, 2004), such as reef environments. However, in order to incorporate the results of this study in fishery management measures, vulnerability analysis (VA) can be utilised as a component of a risk analysis when only qualitative data is available and researchers lack an understanding of cause/effect relationships (Walker *et al.* 2002; McFadden 2007).

Most fishing equipment is not selective and many fish are caught as by-catch, even when there is only one target species. These species are highly affected by fishery efforts because by-catch will continue to be fished as long as it remains viable to catch the target species. Trap fisheries targeting reef fish have recently developed in Brazil and are on the rise in the north-eastern region. Regulatory agencies often take a long time to react and severe declines in non-target species go unperceived. Thus, little is known regarding the impact of this activity and such lack of information hinders regulatory efforts. The method presented here is an appropriate, practical tool for fishery management and the assessment of catch sustainability in multi-species fisheries on which there is limited available data. It can also be employed to determine which species are at a possible risk of extinction owing to being by-catch. Furthermore, it is important to consider that to reduce the extinction threat posed on those species, a socioeconomic VA is required in order to develop the appropriate institutional framework/outreach program to the fishers necessary to address this ecological challenge.

## Acknowledgements

We would like to thank Sérgio Mattos, Simone Teixeira and Paulo Travassos for helpful comments on the first draft of the paper. We also thank the anonymous reviewers for constructive suggestions on the manuscript. This study was sponsored by the Brazilian Scientific Council – Conselho Nacional de Pesquisa e Desenvolvimento (CNPq).

## References

Allen, G. R. (1998). Introducing angelfish. In 'A Guide to Angelfish and Butterflyfish'. pp. 8–12. (Mergus Publishers: Melle.)

- Andrews, C. (1990). The ornamental fish trade and conservation. *Journal of Fish Biology* **37**, 53–59. doi:10.1111/J.1095-8649.1990.TB05020.X
- Araújo, M. E., and Albuquerque Filho, A. C. (2005). Biologia das principais espécies de peixes ornamentais marinhos do Brasil: uma revisão bibliográfica e documental. *Boletim Técnico Científico do CEPENE* **13**, 109–154.
- Bellwood, D. R., van Herwerden, L., and Konow, N. (2004). Evolution and biogeography of marine angelfish (Pisces: Pomacanthidae). *Molecular Phylogenetics and Evolution* **33**, 140–155. doi:10.1016/J.YMPEV.2004.04.015
- Beverton, R. J. H., and Holt, S. J. (1956). A review of methods for estimating mortality rates in exploited fish populations, with special reference to sources of bias in catch sampling. *Rapports et procès-verbaux des Reunions Conseil permanent International pour l'Exploration de la mer* **140**, 67–83.
- Burns, K. M., and Restrepo, V. (2002). Survival of reef fish after rapid depersurization: field and laboratory studies. *American Fisheries Society Symposium* **30**, 148–151.
- Carvalho-Filho, A. (1999). As espécies e suas famílias. In 'Peixes: Costa Brasileira'. pp. 40–283. (Editora Melro: São Paulo.)
- Casey, J. M., and Myers, R. A. (1998). Near extinction of a large, widely distributed fish. *Science* **281**, 690–691. doi:10.1126/SCIENCE.281.5377.690
- Cervigón, F. (1993). Pomacanthidae. In 'Los Peces Marinos de Venezuela'. pp. 339–346. (Fundación Científica Los Roques: Caracas.)
- Chan, T. T. C., and Sadovy, Y. (1998). Profile of the marine aquarium fish trade in Hong Kong. *Aquarium Sciences and Conservation* **2**, 197–213. doi:10.1023/A:1009644730784
- Cheung, W. W. L., Watson, R., Morato, T., Pitcher, T. J., and Pauly, D. (2007). Intrinsic vulnerability in the global fish catch. *Marine Ecology Progress Series* **333**, 1–12. doi:10.3354/MEPS333001
- Cowen, R. K., Paris, C. B., and Srinivasan, A. (2006). Scaling of connectivity in marine populations. *Science* **311**, 522–527. doi:10.1126/SCIENCE.1122039
- Dalzell, P., Adams, T. J. H., and Polunin, N. V. C. (1996). Coastal fisheries in the Pacific Islands. *Oceanography and Marine Biology: An Annual Review* **34**, 395–531.
- Denney, N. H., Jennings, S., and Reynolds, J. D. (2002). Life-history correlates of maximum population growth rates in marine fish. *Proceedings of the Royal Society of London* **269**, 2229–2237. doi:10.1098/RSPB.2002.2138
- Dulvy, N. K., Sadovy, Y., and Reynolds, J. D. (2003). Extinction vulnerability in marine populations. *Fish and Fisheries* **4**, 25–64. doi:10.1046/J.1467-2979.2003.00105.X
- Dulvy, N. K., Ellis, J. R., Goodwin, N. B., Grant, A., Reynolds, J. D., and Jennings, S. (2004). Methods of assessing extinction risk in marine fish. *Fish and Fisheries* **5**, 255–276. doi:10.1111/J.1467-2679.2004.00158.X
- Dye, A. H., Schleyer, M. H., Lambert, G., and Lasiak, T. A. (1994). Biological options for the management of the exploitation of intertidal and subtidal resources. In 'Rocky Shores: Exploitation in Chile and South Africa'. (Ed. W. R. Siegfried.) pp. 131–154. (Springer-Verlag: New York).
- Eayrs, S. (2007). Guía para Reducir la Captura de Fauna Incidental (Bycatch) en las Pesquerías por Arrastre de Camarón Tropical. FAO, Rome. Available at <ftp://ftp.fao.org/FI/DOCUMENT/rebyc/a1008s.pdf> [Accessed 9 December 2008].
- Francis, R. C. (1992). Sexual lability in teleosts: developmental factors. *The Quarterly Review of Biology* **67**, 1–18. doi:10.1086/417445
- Froese, R., and Pauly, D. (2007). FishBase. Available at <http://www.fishbase.org> [Accessed 29 October 2007].
- Garrison, V. H., Rogers, C. S., and Beets, J. (1998). Of reef fish, overfishing and *in situ* observations of fish traps in St. John, U. S. Virgin Islands. *Revista de Biología Tropical* **46**, 41–59.
- Garrison, V. H., Rogers, C. S., Beets, J., and Friedlander, A. M. (2004). The habitats exploited and the species trapped in a Caribbean

- island trap fishery. *Environmental Biology of Fishes* **71**, 247–260. doi:10.1007/S10641-004-0301-X
- Gasparini, J. L., Floeter, S. R., Ferreira, C. E. L., and Sazima, I. (2005). Marine ornamental trade in Brazil. *Biodiversity and Conservation* **14**, 2883–2899. doi:10.1007/S10531-004-0222-1
- Hawkins, J. P., and Roberts, C. M. (2004). Effects of fishing on sex-changing Caribbean parrotfish. *Biological Conservation* **115**, 213–226. doi:10.1016/S0006-3207(03)00119-8
- Hawkins, J. P., Roberts, C. M., and Clark, V. (2000). The threatened status of restricted-range coral reef fish species. *Animal Conservation* **3**, 81–88. doi:10.1111/J.1469-1795.2000.TB00089.X
- Hourigan, T. F., Stanton, F. G., Motta, P. J., Kelley, C. D., and Carlson, B. (1989). The feeding ecology of three species of Caribbean angelfish (family Pomacanthidae). *Environmental Biology of Fishes* **24**, 105–116. doi:10.1007/BF00001281
- Hutchings, J. A. (2001). Conservation biology of marine fish: perceptions and caveats regarding assignment of extinction risk. *Canadian Journal of Aquatic Sciences* **58**, 108–121. doi:10.1139/CJFAS-58-1-108
- Kelleher, K. (2005). Discards in the world's marine fisheries. An update. FAO Fisheries Technical Paper No 470. FAO, Rome. Available at <http://www.fao.org/docrep/008/y5936e/y5936e00.htm> [Accessed 9 December 2008].
- Lieske, E., and Myers, R. (1994). Spadefish and butterflyfish. In 'Collins Pocket Guide. Coral Reef Fish. Indo-Pacific and Caribbean Including the Red Sea'. pp. 161–162. (Princeton University Press: Princeton, NJ.)
- Mahon, R., and Hunte, W. (2001). Trap mesh selectivity and the management of reef fish. *Fish and Fisheries* **2**, 356–375. doi:10.1046/J.1467-2960.2001.00054.X
- Malakoff, D. (1997). Extinction on the high seas. *Science* **277**, 486–488. doi:10.1126/SCIENCE.277.5325.486
- McFadden, L. (2007). Vulnerability analysis in environmental management: widening and deepening its approach. *Environmental Conservation* **34**, 195–204. doi:10.1017/S0376892907004195
- McKinney, M. L. (1997). Extinction vulnerability and selectivity: combining ecological and paleontological views. *Annual Review of Ecology and Systematics* **28**, 495–516. doi:10.1146/ANNUREV.ECOLSYS.28.1.495
- Michael, S. W. (2004). Family Pomacanthidae/angelfish. In 'Angelfish and Butterflyfish'. pp. 210–321. (T. F. H. Publications: Neptune City, NJ.)
- Milton, D. A. (2001). Assessing the susceptibility to fishing of populations of rare trawl bycatch: sea snakes caught by Australia's Northern Prawn Fishery. *Biological Conservation* **101**, 281–290. doi:10.1016/S0006-3207(00)00232-9
- Monteiro Neto, C., Cunha, F. E. A., Nottingham, M. C., Araújo, M. E., Rosa, I. L., and Barros, G. M. L. (2003). Analysis of the marine ornamental fish trade at Ceará State, northeast Brazil. *Biodiversity and Conservation* **12**, 1287–1295. doi:10.1023/A:1023096023733
- Moyer, J. T., Thresher, R. E., and Colin, P. L. (1983). Courtship, spawning and inferred social organization of American angelfish (genera *Pomacanthus*, *Holocanthus* and *Centropyge*: Pomacanthidae). *Environmental Biology of Fishes* **9**, 25–39. doi:10.1007/BF00001056
- Myers, R. A., and Worm, B. (2005). Extinction, survival or recovery of large predatory fish. *Philosophical Transactions of the Royal Society* **360**, 13–20. doi:10.1098/RSTB.2004.1573
- Nottingham, M. C., Cunha, F. E., and Monteiro Neto, C. (2000). Captura de peixes ornamentais marinhos no Ceará. *Arquivos Ciências do Mar* **33**, 113–118.
- Pope, J. G., MacDonald, D. S., Dann, N., Reynolds, J. D., and Jennings, S. (2000). Gauging the impact of fishing mortality on nontarget species. *ICES Journal of Marine Science* **57**, 689–696. doi:10.1006/JMSC.2000.0729
- Randall, J. E. (1967). Food habits of reef fishes of the West Indies. *Studies on Tropical oceanography* **5**, 665–847.
- Reynolds, J. D., and Jennings, S. (2000). The role of animal behavior in marine conservation. In 'Behavior and Conservation'. (Eds L. M. Gosling and W. J. Shuterland.) pp. 147–168. (Cambridge University Press: Cambridge, UK.)
- Reynolds, J. D., Jennings, S., and Dulvy, N. K. (2001). Life histories of fish and population responses to exploitation. In 'Conservation of Exploited Species'. (Eds J. D. Reynolds, G. M. Mace, K. H. Redford and J. G. Robinson.) pp. 147–168. (Cambridge University Press: Cambridge, UK.)
- Reynolds, J. D., Dulvy, N. K., and Roberts, C. M. (2002). Exploitation and other threats to fish conservation. In 'Fish and Fisheries Handbook'. (Eds P. J. B. Hart and J. D. Reynolds.) pp. 319–341. (Blackwell Science: Oxford.)
- Ribeiro, F. P. (2004). Composição da biocenose e abundância relativa de peixes capturados com covos nos Estados do Rio Grande do Norte e Pernambuco. *Boletim Técnico Científico do CEPENE* **12**, 113–128.
- Roberts, C. M., and Hawkins, J. P. (1999). Extinction risk in the sea. *Trends in Ecology & Evolution* **14**, 241–246. doi:10.1016/S0169-5347(98)01584-5
- Robins, C. R., and Ray, C. G. (1986). Butterflyfish: family Chaetodonidae. In 'A Field Guide to Atlantic Coast Fish of North America'. pp. 192–194. (Houghton Mifflin Company: New York.)
- Rocha, L. A., Craig, M. T., and Bowen, B. W. (2007). Phylogeography and the conservation of coral reef fish. *Coral Reefs* **26**, 501–512.
- Sadovy, Y. (2005). Trouble on the reef: the imperative for managing vulnerable and valuable fisheries. *Fish and Fisheries* **6**, 167–185. doi:10.1111/J.1467-2979.2005.00186.X
- Sadovy, Y., and Shapiro, D. Y. (1987). Criteria for the diagnosis of hermaphroditism in fishes. *Copeia* **1987**, 136–156. doi:10.2307/1446046
- Sparre, P., and Venema, S. C. (1992). Introduction to tropical fish stock assessment, part 1. FAO Fisheries Technical Paper No 306. FAO: Rome. Available at <ftp://ftp.fao.org/docrep/fao/w5449e/w5449e00.pdf> [Accessed 9 December 2008].
- Starck, W. A. I., and Davis, W. P. (1966). Night habits of fish of Alligator Reef, Florida. *Ichthyologica* **38**, 313–356.
- Stobutzki, I., Blaber, S., Brewer, D., Fry, G., Heales, D., et al. (2000). Ecological sustainability of bycatch and biodiversity in prawn trawl fisheries. Final Report Project 96/257. Fisheries Research and Development Corporation, Canberra.
- Stobutzki, I., Miller, M., and Brewer, D. (2001). Sustainability of fishery bycatch: a process for assessing highly diverse and numerous bycatch. *Environmental Conservation* **28**, 167–181. doi:10.1017/S0376892901000170
- Swingle, W. E., Dammann, A. E., and Yntema, A. N. (1970). Survey of the commercial fishery of the Virgin Islands of the United States. *Proceedings of the Gulf and Caribbean Fisheries Institute* **20**, 110–121.
- Thresher, R. E. (1980). Angelfish. In 'Reef fish: Behavior and Ecology on the Reef and in the Aquarium'. pp. 37–46. (The Palmetto Publishing Company: St Petersburg, FL.)
- Thresher, R. E. (1984). Angelfish (Pomacanthidae). In 'Reproduction in Reef Fish'. pp. 244–261. (T. F. H. Publications: Neptune City, NJ.)
- Walker, B., Carpenter, S., Anderies, J., Abel, N., Cumming, G., Janssen, M., Lebel, L., Norberg, J., Peterson, G. D., and Pritchard, R. (2002). Resilience management in social-ecological systems: a working hypothesis for a participatory approach. *Conservation Ecology* **6**, 14.
- Wolff, N., Grober-Dunsmore, R., Rogers, C. S., and Beets, J. (1999). Management implications of fish trap effectiveness in adjacent coral reef and gorgonian habitats. *Environmental Biology of Fishes* **55**, 81–90. doi:10.1023/A:1007430407540
- Wood, E. M. (2001a). 'Collection of Coral Reef Fish for Aquaria: Global Trade, Conservation Issues and Management Strategies.' (Marine Conservation Society: Ross-on-Wye, UK.)
- Wood, E. M. (2001b). Global advances in conservation and management of marine ornamental resources. *Aquarium Science and Conservation* **3**, 65–77. doi:10.1023/A:1011391700880

Manuscript received 27 February 2008, accepted 22 September 2008