



UNIVERSIDADE FEDERAL DE PERNAMBUCO
CENTRO DE BIOCIENTÍCIAS
PROGRAMA DE PÓS-GRADUAÇÃO EM CIÊNCIAS BIOLÓGICAS

CÍCERO RAMON BEZERRA DOS SANTOS

AVALIAÇÃO FITOQUÍMICA E ANTIPARASITÁRIA DO ÓLEO ESSENCIAL DE
***Eugenia stipitata* McVaugh**

Recife

2018

CÍCERO RAMON BEZERRA DOS SANTOS

**AVALIAÇÃO FITOQUÍMICA E ANTIPARASITÁRIA DO ÓLEO ESSENCIAL DE
Eugenia stipitata McVaugh**

Dissertação apresentada ao Programa de Pós-Graduação em Ciências Biológicas da Universidade Federal de Pernambuco, como parte dos requisitos para obtenção do grau de mestre.

Orientadora: Profª. Drª. Márcia Vanusa da Silva – UFPE

Recife

2018

Dados Internacionais de Catalogação na Publicação (CIP) de acordo com ISBD

Santos, Cícero Ramon Bezerra dos

Avaliação fitoquímica e antiparasitária do óleo essencial de *Eugenia stipitata* Mc Vaugh/ Cícero Ramon Bezerra dos Santos- 2018.

83 folhas: il., fig., tab.

Orientadora: Márcia Vanusa da Silva

Dissertação (mestrado) – Universidade Federal de Pernambuco. Centro de Biociências. Programa de Pós-Graduação em Ciências Biológicas.

Recife, 2018.

Inclui referências e anexo

1. Plantas medicinais 2. Essencias e óleos essenciais- uso terapêutico 3. Caatinga I. Silva, Márcia Vanusa da (orient.) II. Título

581.634

CDD (22.ed.)

UFPE/CB-2018-333

CÍCERO RAMON BEZERRA DOS SANTOS

AVALIAÇÃO FITOQUÍMICA E ANTIPARASITÁRIA DO ÓLEO ESSENCIAL DE
Eugenia stipitata McVaugh

Dissertação apresentada ao Programa de
Pós-Graduação em Ciências Biológicas da
Universidade Federal de Pernambuco,
como parte dos requisitos para obtenção do
grau de mestre.

Aprovada em: 30/ 07 /2018.

BANCA EXAMINADORA:

Prof^a. Dr^a. Márcia Vanusa da Silva - UFPE
(Orientadora)

Prof^o. Dr^o. Alexandre Gomes da Silva - UFPE
(1º Examinador)

Prof^o. Dr^o. Thiago Henrique Napoleão - UFPE
(2º Examinador)

Primeiramente a Deus, pela força contínua durante minha caminhada. Dedico também à minha amada tia Mônica Gonçalves Bezerra (in memoriam), que deixa em meu coração uma saudade eterna.

AGRADECIMENTOS

A Deus, por ter me dado forças.

Aos meus pais, pela educação que a mim dedicaram. Exemplos de sabedoria, garra e honestidade. Obrigado por todo amor e dedicação de todos esses anos.

À minha orientadora, Professora Doutora Márcia Vanusa, por ter aceito a orientação e confiado no meu trabalho.

Ao meu co-orientador, Professor Doutor Irwin Rose de Alencar, obrigado pela confiança depositada em mim.

A Douglas Wylliam por ser tão importante na minha vida. Graças ao seu companheirismo, apoio, amizade, paciência, compreensão, alegria e amor, esse trabalho também pôde ser concretizado.

Às minhas amigas e colegas de profissão, Mariana Gomes e Lilian Vandesmet, pela amizade, por sempre me indicarem o caminho e pelo apoio constante.

Ao meu amigo Doutor Eduardo Brandão, pela amizade que me devotas, por ter me incentivado nesse projeto e por ter me acolhido em Recife.

À Universidade Federal do Pernambuco, em especial ao Centro de Biociências, pela oportunidade.

Ao Centro Universitário Católica de Quixadá, pela compreensão e pelas oportunidades.

À banca examinadora, por ter aceito o convite.

A todos que contribuíram para a realização desse sonho, meu muito obrigado.

*“O tempo é algo que não volta atrás. Por isso
plante seu jardim e decore sua alma, ao invés
de esperar que alguém lhe traga flores ...”*
(William Shakespeare).

RESUMO

Os óleos essenciais (EOs) de plantas utilizadas na medicina tradicional são conhecidos como uma rica fonte de compostos quimicamente diferentes com diversas atividades biológicas. Neste estudo, analisamos a composição química e investigamos o efeito tripanocida, leishmanicida e a citotoxicidade *in vitro* do óleo essencial obtido a partir das folhas de *Eugenia stipitata* McVaugh. O OE foi extraído pelo método de destilação por arraste à vapor e sua composição química analisada por GC/MS e testado em sete concentrações (15.75, 31.25, 62.5, 125, 250, 500 e 1000 µg/mL) contra as formas epimastigotas de *Trypanosoma cruzi* e as formas promastigotas de *Leishmania braziliensis* e de *Leishmania infantum*, bem como contra fibroblastos. De modo geral, 48 componentes foram identificados na análise química do óleo e os constituintes mais abundantes foram: β-Eudesmol (15,28%), γ-Eudesmol (10,85%), Elemol (10,21%), Caryophyllene oxide (6,5%), Clovène (6,18%) e Epatulenol (6,14%). Na avaliação das atividades antiparasitárias, o OE de *Eugenia stipitata* McVaugh em concentrações de 125 µg/mL e 62.5 µg/mL inibiu, respectivamente, 80.69% e 71.91% da forma promastigota de *L. braziliensis*. Inibição semelhante foi observada para as mesmas concentrações do óleo contra *L. infantum*, que inibiu 76.51% da forma promastigona de *L. infantum* na concentração de 125 µg/mL e 73.51% na concentração de 62.5 µg/mL. Contudo, na concentração de 125 µg/mL foi observada uma menor inibição parasitária do óleo essencial de *Eugenia stipitata* McVaugh frente às formas epimastigotas de *T. cruzi*, com 22.85%. O óleo essencial de *Eugenia stipitata* McVaugh apresentou uma baixa citotoxicidade na concentração de 62.5 µg/mL e nenhuma citotoxicidade nas concentrações de 31.25 µg/mL e 15.75 µg/mL. Desse modo, o OE estudado apresentou-se como promissora fonte antiparasitária contra as formas de vida da *L. braziliensis* e *L. infantum*.

Palavras chaves: Óleo essencial. Caatinga. Antiprotozoário. *Eugenia stipitata* McVaugh.

ABSTRACT

The essential oils (EOs) of plants used in traditional medicine are known as a rich source of chemically different compounds with different biological activities. In this study, we analyzed the chemical composition and investigated the trypanocidal, leishmanicide, effect and in vitro cytotoxicity of essential oil obtained from the leaves of *Eugenia stipitata* McVaugh. EO was extracted by the distillation method by steam stripping and its chemical composition analyzed by GC/MS and tested in seven concentrations (15.75, 31.25, 62.5, 125, 250, 500 e 1000 µg/mL) against the forms of *Trypanosoma cruzi* epimastigotes and promastigotes forms of *Leishmania braziliensis* and *Leishmania infantum*, as well as against fibroblasts. Generally, 48 components were identified in the oil chemical analysis and the most abundant constituents were: β -eudesmol (15.28%), γ -eudesmol (10.85%), Elemol (10.21%), caryophyllene oxide (6.5%), Clovane (6.18%) and Espatulenol (6.14%). In the evaluation of the parasitic activities, EO of *Eugenia stipitata* McVaugh at concentrations of 125 µg/mL and 62.5 µg/mL inhibited, respectively, 80.69% and 71.91% of the promastigote form of *L. braziliensis*. Similar inhibition was observed for the same concentrations of the oil against *L. infantum*, which inhibited 76.51% of promastigote form of *L. infantum* in concentration of 125 µg/mL and 51.1% in the concentration of 62.5 µg/mL. However, in the concentration of 125 µg/mL a smaller parasitic inhibition was observed of essential oil of *Eugenia stipitata* McVaugh before the epimastigotes forms of *T. cruzi*, with 22.85%. The essential oil of *Eugenia stipitata* McVaugh presented a low cytotoxicity in the concentration of 62.5 µg/mL and no cytotoxicity in concentrations of 31.5 µg/mL and 15.75 µg/mL. Therefore, the studied EO presented itself as a promising antiparasitic source against the ways of life of *L. braziliensis* and *L. infantum*.

Keywords: Essential Oil. Caatinga. Antiprotozoal. *Eugenia stipitata* McVaugh.

LISTA DE FIGURAS

Figura 1 - Localização das Caatingas na América do Sul.	18
Figura 2 - Arbusto de <i>Eugenia stipitata</i> , conhecida popularmente como araçá-boi.	21
Figura 3 – Ciclo Biológico da <i>Leishmania</i> .	23
Figura 4 - A) Forma Promastigota de <i>Leishmania sp</i> ; B) Forma Amastigota de <i>Leishmania sp</i> .	23
Figura 5 – Ciclo Biológico do <i>Trypanosoma cruzi</i> .	25
Figura 6 - A: Forma amastigota do <i>T. cruzi</i> .; B: Forma tripomastigota do <i>T. cruzi</i> ; C: Forma epimastigota do <i>T. cruzi</i> .	25

LISTA DE TABELAS

Table 1 - Chemical composition of the essential oil from <i>Eugenia stipitata</i> McVaugh	43
Table 2 - Anti-Leishmani, trypanocidal activity and cytotoxicity of the essential oil of <i>Eugenia stipitata</i> .	45
Table 3 - Positive control of benzonidazole against <i>Trypanosoma cruzi</i> ($\mu\text{g/mL}$)	46
Table 4 - Positive control of miltefosine against <i>Leishmania braziliensis</i> and <i>Leishmania infantum</i> (μM).	46

LISTA DE ABREVIAÇÕES E SIGLAS

cm	Centímetros
CD	Chagas disease
EOS	Essential oils
FBS	Fetal Bovine Serum
GC/MS	Gas chromatography-mass spectrometry
LTA	Leishmaniose Tegumentar Americana
LV	Leishmaniose visceral
µg/mL	Micrograma por mililitro
µM	Micrômetro
mm	Milímetros
nm	Nanômetros
EOESL	Oil from the leaves of <i>E. stipitata</i>
OE	Óleo essencial
OMS	Organização Mundial de Saúde
km ²	Quilômetros quadrados
SDTF	Seasonally dry tropical forests
UFPE	Universidade Federal de Pernambuco
URCA	Universidade Regional do Cariri

SUMÁRIO

1 INTRODUÇÃO	13
2 OBJETIVOS	16
2.1 OBJETIVO GERAL	16
2.2 OBJETIVOS ESPECÍFICOS	16
3 REVISÃO BIBLIOGRÁFICA	17
3.1 O BIOMA CAATINGA	17
3.2 CONSIDERAÇÕES GERAIS SOBRE MYRTACEAE	18
3.3 GÊNERO <i>EUGENIA</i>	19
3.4 CONSIDERAÇÕES GERAIS SOBRE A ESPÉCIE <i>EUGENIA STIPITATA</i>	21
3.5 <i>LEISHMANIA</i>	22
3.6 <i>TRYPANOSOMA CRUZI</i>	24
3.7 ÓLEOS ESSENCIAIS	26
3.8 ÓLEOS ESSENCIAIS COM ATIVIDADE ANTIPARASITÁRIA	27
4 ARTIGO 1 - CHEMICAL COMPOSITION AND EVALUATION OF ANTIPARASITIC ACTIVITY OF THE ESSENTIAL OIL OF <i>EUGENIA STIPITATA</i> MCVAUGH (MYRTACEAE)	29
5 CONCLUSÃO GERAL	57
REFERÊNCIAS	58
ANEXO A – Normas da Revista	68

1 INTRODUÇÃO

A utilização das plantas como medicamento é uma das mais antigas armas empregadas para o tratamento das enfermidades humanas e muito já se conhece a respeito de seu uso por parte da sabedoria popular. A Organização Mundial de Saúde comenta que, na busca incessante do homem por bem-estar e qualidade de vida, a fitoterapia tornou-se uma alternativa devido à credibilidade terapêutica e ao baixo custo, o que favorece sua utilização por uma grande parcela da população mundial (LUCHESSI et al., 2005).

Em regiões que abrangem a Caatinga, espécies vegetais são amplamente utilizadas pelas comunidades no tratamento de suas enfermidades. Estas comunidades possuem uma vasta farmacopeia natural, grande parte proveniente dos recursos vegetais encontrados nos ambientes naturais habitados por estas populações, ou cultivados em ambientes transformados pelo próprio homem (GOMES et al., 2008). De acordo com Matos (2002), 90% da população economicamente carente do Nordeste utiliza a medicina tradicional, em busca da cura de seus problemas de saúde.

A busca de plantas medicinais e seus derivados como agentes terapêuticos naturais vem sendo priorizada, a fim de amenizar as enfermidades sofridas pela população. A utilização destes fitoterápicos é incentivada por apresentarem um custo mais acessível à população e aos serviços públicos de saúde, em comparação aos fármacos obtidos por síntese química (TOLEDO et al., 2003; SILVA, 2006).

Dentre os agentes terapêuticos vegetais destacam-se os óleos essenciais (OEs), produzidos nas folhas em estruturas chamadas tricomas glandulares, provenientes do metabolismo secundário das plantas que atuam nos mecanismos de defesa contra as condições adversas (MICHELIN, 2008). A obtenção de uma droga vegetal de qualidade tem correlação direta com o conhecimento prévio das classes de componentes químicos encontrados nesses vegetais. Quando confirmada a presença de determinados grupos químicos, o estudo fitoquímico e biológico é direcionado através de ensaios que avaliam substâncias oriundas de plantas, em extratos ou substâncias isoladas. Apesar do interesse na utilização desses produtos vegetais, ainda são poucas as pesquisas que relatam as propriedades biológicas de espécies em vegetações predominantemente brasileiras como a Caatinga (LÔBO et al., 2010).

A família Myrtaceae, no Brasil, é representada por 23 gêneros e cerca de 1000 espécies. Estudos apontam que as plantas pertencentes a essa família são fontes de óleos essenciais responsáveis por importantes benefícios associados à saúde (D'ANGELIS; NEGRELLE, 2014). O gênero *Eugenia* encontra-se representado por 400 espécies nos diversos domínios

fitogeográficos do Brasil, e muitas destas espécies destacam-se pela produção de óleos essenciais que apresentam diversas propriedades farmacológicas (QUEIROZ et al., 2015).

Especificamente, as espécies *E. brasiliensis* e *E. uniflora* já tiveram seus potenciais antiparasitários investigados anteriormente frente ao *T. cruzi* e o OE das flores de *E. klotzschiana* apresentou satisfatória atividade tripanocida frente às formas tripomastigota desse parasita (AZEVEDO et al., 2014; CARNEIRO et al., 2017; DONATO; MORRETES, 2007). Além disso, atividade leishmanicida de produtos naturais, com atividade citotóxica contra o parasita, obtidos de plantas, tem sido considerado promissor no desenvolvimento de fármacos (RODRIGUES et al., 2015).

Eugenia stipitata McVaugh é uma frutífera, conhecida popularmente como araçá-boi (MCVAUGH, 1956). Na medicina popular, é utilizado no tratamento de desordens intestinais e urinárias, além de também aliviar os sintomas do resfriado, o que indica que os compostos bioativos presentes na planta podem ter potenciais benefícios para a saúde humana (FERNÁNDEZ-TRUJILLO et al., 2011). Entre as propriedades biológicas investigadas, podem-se destacar a atividade antioxidante na eliminação de radicais livres, potencial antimicrobiano do OE produzido a partir de suas folhas e atividades antimutagênica e antigenotóxica de extratos do fruto, sugerindo que estes podem funcionar como agentes preventivos contra o câncer (DUKE, 2017; MEDEIROS et al., 2003; NERI-NUMA et al., 2013).

As infecções por protozoários são consideradas um problema mundial de saúde pública, especialmente em países subdesenvolvidos, onde aproximadamente 14% da população são considerados como risco de infecção (KONDRASHIN et al., 2011; WALDRON et al., 2011). Neste contexto, destacam-se: a doença causada pelo protozoário *Trypanosoma cruzi*, denominada doença de Chagas, cujo tratamento quimioterápico apresenta vários efeitos colaterais, como: anorexia, náuseas, alterações gastrointestinais, dermatopatia alérgica, polineurite, depressão da medula óssea, neuropatia periférica entre outros; e a leishmaniose, causada pelo protozoário *Leishmania*, considerada pela Organização Mundial de Saúde (OMS) como uma das seis principais doenças infecciosas com alta incidência e capacidade de produzir deformidades, cujos tratamentos convencionais são limitados, inseguros e têm uma série de efeitos colaterais, além de contribuírem para a resistência do parasita ao tratamento (ALIZADEH et al., 2008; FERREIRA, 2012; MITROPOULOS et al., 2010; OLIVEIRA et al., 2008).

Devido a estas complicações causadas pelos fármacos disponíveis, torna-se cada vez mais relevante a busca por novos agentes quimioterápicos, com menos efeitos colaterais quando

comparado à terapia alopática e mais acessível à população. Dessa forma, o isolamento, a caracterização e a aplicação biológica de compostos naturais, sobretudo de plantas nativas da Caatinga devido à sua grande biodiversidade, possuem um elevado grau de significância na produção de novas drogas com perfil farmacológico.

Nesse contexto, o presente estudo objetiva avaliar as atividade citotóxica, leishmanicida e tripanocida do óleo essencial de *Eugenia stipitata* McVaugh, além de quantificar os componentes químicos do mesmo utilizando cromatografia gasosa acoplada à espectrometria de massa.

2 OBJETIVOS

2.1 OBJETIVO GERAL:

Avaliar a composição química e os efeitos tripanocida, leishmanicida e a citotoxicidade *in vitro* do óleo essencial obtido a partir das folhas de *Eugenia stipitata* McVaugh.

2.2 OBJETIVOS ESPECÍFICOS:

- Obter o óleo essencial das folhas de *Eugenia stipitata* McVaugh.
- Quantificar os constituintes químicos do óleo essencial de *Eugenia stipitata* McVaugh através da cromatografia gasosa acoplada à espectrômetro de massas.
- Avaliar a citotoxicidade e as atividades tripanocida, contra a forma epimastigota de *Trypanosoma cruzi* e leishmanicida, contra as formas promastigotas de *Leishmania braziliensis* e *Leishmania infantum* do óleo essencial.

3 REVISÃO BIBLIOGRÁFICA

3.1 O BIOMA CAATINGA

A caatinga é um ecossistema de florestas tropicais secas localizadas na região semiárida do nordeste do Brasil. O termo caatinga (“floresta branca”) se refere à um mosaico de fisionomias vegetais de florestas secas, xerófitas, lenhosas, espinhosas e decíduas, com uma vegetação arbustiva sazonal. Essas variações são atribuídas principalmente em decorrência das alterações climáticas, padrões orográficos, modificações em pequena escala na topografia e nos solos (ANDRADE-LIMA, 1981; SAMPAIO, 1995).

Abrangendo a maioria dos estados do Nordeste como: Piauí, Ceará, Rio Grande do Norte, Paraíba, Pernambuco, Alagoas, Sergipe, Bahia e no nordeste de Minas Gerais, no Vale de Jequitinhonha, ocupando uma área que se estende por cerca de 850.000 km² e está cercada por regiões Amazônicas e da Mata Atlântica a leste e oeste, e de savanas do Cerrado ao sul (Figura 1) (BASSO et al., 2005). Isso corresponde a 10% do território brasileiro e 60% da região nordeste. Sua precipitação média anual varia de 240 a 1500 mm, estando restrita a três meses consecutivos, com temperatura média anual de 27,5 °C (ANDRADE-LIMA, 1981; LEAL et al., 2005; ALVES et al., 2011).

Anteriormente, a Caatinga era considerada um bioma pobre em espécies vegetais e, portanto, indigno de medidas de conservação. Entretanto, é de conhecimento geral que existem 510 gêneros e 5344 espécies de plantas vasculares na Caatinga, dentre as quais 18 gêneros e 318 espécies são endêmicos (GIULIETTI et al., 2002). Dessa forma, o bioma é dominado por um dos poucos tipos de vegetação cuja distribuição é totalmente restrita ao Brasil (SILVA; ALBUQUERQUE, 2005).

Espécies de plantas medicinais pertencentes à Caatinga, como *Myracrodruon urundeuva* Allemão, *Amburana cearensis* (Arr. Cam.) A.C. Smith., e *Anadenanthera colubrina* (Vell.) Brenan var. *cebil* (Griseb) Altschul, são amplamente conhecidas e utilizadas na medicina popular, e para confecção de produtos fitoterápicos (ALMEIDA; ALBUQUERQUE, 2002; ALBUQUERQUE et al., 2007). Apesar da sua diversidade de recursos, a vegetação da Caatinga faz parte de um bioma altamente ameaçado e ainda pouco estudado do ponto de vista etnobotânico, fitossociológico e farmacológico, dada a riqueza cultural e biológica que lá existe (VANDESMET, 2015).

Figura 1: Localização das Caatingas na América do Sul



Fonte: LEAL; SILVA; TABARELLI, 2005.

3.2 CONSIDERAÇÕES GERAIS SOBRE MYRTACEAE

Myrtaceae, uma das maiores famílias de Angiospermas, engloba cerca de 129 gêneros e aproximadamente 4.620 espécies de árvores e arbustos, que por suas características e atributos são incluídas em duas subfamílias: Psiloxiloideae, com duas tribos e Myrtoideae, com quinze tribos (WILSON et al., 2001, 2005). A família apresenta ampla distribuição geográfica, com particular ocorrência na Austrália, Índia, América tropical e várias outras regiões de clima temperado e semiárido (DI STASI; HIRUMA-LIMA, 2002; OLIVEIRA et al., 2005).

Apresenta-se distribuída em todos os domínios fitogeográficos brasileiros, ocorrendo quase 1000 espécies subordinadas a 23 gêneros. As mirtáceas estão entre as 10 famílias com maior riqueza de espécies na flora do país, sendo a Floresta Atlântica um de seus centros de diversidade, onde é a sexta maior família em níveis de diversidade. A mesma engloba os gêneros *Myrtus*, *Psidium*, *Pimenta*, *Eugenia*, *Syzygium*, *Eucalyptus*, *Leptospermum* e *Malaleuca* (FORZZA et al., 2010; FERNANDES, 2011).

As mirtáceas desempenham um grande papel na economia, uma vez que várias espécies são cultivadas, seja por seus frutos comestíveis, com propósito ornamental, pela extração de essências ou madeiras com alto valor comercial. Seus representantes também possuem várias propriedades medicinais, grande parte destas atribuídas aos óleos essenciais das estruturas secretoras de seus órgãos vegetativos e reprodutivos (BARROSO et al., 1984; MELO, 2009).

Do ponto de vista químico, várias substâncias com atividade farmacológica foram isoladas em estudos a partir de espécies dessa família, tais como o cariofileno, biciclogermacreno, espatulenol, carotenoides, ácido ascórbico e compostos fenólicos (MORENO et al., 2014; MIRANDA et al., 2017; PEREIRA et al., 2017).

O uso medicinal das muitas espécies desta família justifica-se pelas diversas atividades comprovadas, como antimicrobiana, antiviral, hipoglicemiante, antioxidante e anticancerígena. Entre os exemplos mais frequentes, podem-se citar a goiabeira, araçá, jabuticaba, ponhema, sabará, pitanga, uvária, cabeludinha, quimixama, guabiroba e o cambuci (SILVA; CASALI, 2000; APEL et al., 2006; SERAFIM, 2006). Ainda revisando a literatura, encontra-se algumas pesquisas que revelam inúmeras plantas pertencentes a família Myrtaceae com atividade antiparasitária como, por exemplo, o estudo realizado por Silva (2007) que revelou potente atividade tripanocida e leishmanicida da planta *Myrcia hiemalis* (Myrtaceae), além da pesquisa de Correia et al (2016) que mostrou satisfatória atividade Leishmanicida do extrato de *Myrciaria dubia* (Myrtaceae).

3.3 GÊNERO *EUGENIA*

O gênero *Eugenia* encontra-se bem representado nos diversos domínios fitogeográficos do Brasil, destacando-se pelo vasto potencial medicinal, econômico, alimentício, bem como para exploração comercial de madeiras e óleos essenciais, além de seu uso como plantas ornamentais (ROMAGNOLO; SOUZA, 2006; ALVES; TRESMONDI; LONGUI, 2008; LAGO et al., 2011).

Possuem representantes na forma de arbustos, subarbustos e árvores, nas quais o caule pode atingir de três a doze metros de altura. As flores exibem-se em racemos, dicásios ou isoladas, com antopódio presente e profilos livres, persistentes ou caducos. Os botões florais abertos apresentam quatro sépalas que frequentemente são desiguais, e as pétalas tetrâmeras semelhantes. Os estames são numerosos; ovário bilocular podendo apresentar de quatro a vinte óvulos por lóculo. Os frutos apresentam coloração heterogênea podendo ser amarelos, alaranjados, vermelhos, vináceos e até pretos quando maduros, sendo as bagas globosas a

elipsóides, com cálice persistente. O número de sementes pode variar de uma a três, o embrião é do tipo eugenióide. Entre os cotilédones existe uma linha de separação e o eixo hipocótilo-radícula apresenta-se pouco desenvolvido (QUEIROZ et al., 2015).

As espécies do gênero *Eugenia* são alvos constantes de pesquisas, uma vez que são detentoras de óleos essenciais e diversos metabólitos secundários que apresentam importantes ações biológicas (VICTORIA et al., 2012). Em estudos realizados com a espécie *Eugenia uniflora* L. importantes propriedades terapêuticas foram descritas, tais como: antiparasitária, antibacteriana, antioxidante, antidepressiva, antinociceptiva e hipotérmicas. Já no viés fitoquímico, os principais marcadores descritos são: ácido gálico, ácido elágico e miricitrina, porém, podem ser encontrados nesta planta outros metabólitos, tais como as antacioninas e sesquiterpenos (OGUNWANDE et al., 2005; AMORIM et al., 2009; BEZERRA et al., 2017; OLIVEIRA et al., 2017; SOUZA et al., 2017).

Em consonância, as atividades biológicas de *Eugenia jambolana* L. incluem: antibacteriana, antidiarreica, antiparasitária, antidiabética, antinociceptiva e anti-inflamatória (MUKHERJEE et al., 1988; SAHA et al., 2013; LI et al., 2017; PEREIRA et al., 2017; SOUZA et al., 2017). A avaliação dos constituintes presentes nessa espécie aponta para a presença de compostos fenólicos nos extratos, como taninos, alcaloides, esteroides, flavonoides, terpenóides, ácidos graxos, fenóis, minerais, carboidratos e vitaminas (MARGARET; SHAILAJA; RAO, 2015; VEBER et al., 2015). Além disso, um rastreio químico e quimiopreventivo realizado com extratos e frações de folhas e frutos de *E. jambolana* levaram à purificação de flavonoides (tricetina-4'-O- α -l-rhamnopiranósido e miricitrina) e antacioninas (malvidina-3-O-gentibiosídeo), sugerindo que esta espécie pode agir como um agente preventivo contra danos ao DNA (DAMETTO et al., 2017).

Outras espécies pertencentes a este gênero que vêm sendo estudadas são *Eugenia punicifolia* (Kunth) DC. e *Eugenia dysenterica* DC. apresentando, respectivamente, tais atividades biológicas: neuroprotetora, gastroprotetora, antioxidante, antidiarreica, antileucêmica e antifúngica; e anti-inflamatórias, antinociceptiva, gastroprotetora, anti-diabética e antioxidante. Análises químicas dessas espécies sugerem a presença de terpenos, flavonoides, saponinas e taninos como responsáveis por suas propriedades farmacológicas (LEITE et al., 2014; GALENO et al., 2014; SALES et al., 2014; BASTING et al., 2014; COSTA et al., 2000; PRADO et al., 2014; GALHEIGO et al., 2015; PEIXOTO, 2015; VITEK et al., 2017).

É cabível citar outros estudos, como o de Carneiro et al (2017), cujos resultados demonstraram satisfatória atividade tripanocida do óleo essencial de *Eugenia klotzschiana* e

Santos et al (2013) em que os resultados indicam que *Eugenia uniflora* apresenta atividade leishmanicida.

3.4 CONSIDERAÇÕES GERAIS SOBRE A ESPÉCIE *EUGENIA STIPITATA*

O araçazeiro-boi (*Eugenia stipitata* McVaugh) é uma frutífera pertencente à família *Myrtaceae*. É originária da Amazônia Ocidental, comumente cultivada em pequena escala no Peru, Bolívia, Equador, Colômbia e Brasil (MCVAUGH, 1956).

O arbusto do araçazeiro-boi alcança de três a cinco metros de altura (Figura 2), com abundante ramificação e folhagem, se adaptando muito bem ao clima tropical úmido. Suas folhas são elípticas, verde-escuras e as pequenas inflorescências possuem de três a dez flores hermafroditas, com pétalas brancas e 75 a 100 estames (CHÁVES-FLORES; CLEMENT, 1984; EMBRAPA, 1996).

Figura 2: Arbusto de *Eugenia stipitata*, conhecida popularmente como araçá-boi.



Fonte: http://www.fruitipedia.com/araca_boi.htm

Seu fruto é uma baga globosa, de formato redondo ou achatao, com casca delgada e coloração amarelo-canário quando madura. Sua polpa é suculenta, pouco fibrosa, e bastante ácida, de coloração amarelo-clara, possuindo de 4 a 10 sementes oblongas de 0,5 a 1,0 cm de comprimento (SACRAMENTO; BARRETTO; FARIA, 2008).

Devido sua intensa acidez, a polpa do araçá-boi não é adequada para o consumo *in natura*. Porém, esta possui aroma e sabor agradáveis, podendo ser consumida em forma de

sucos, sorvetes, geleias, néctar, licores e outros (EMBRAPA, 1996; ANDRADE et al., 1997; SOARES, 2009).

Entre as propriedades biológicas investigadas, podem-se destacar a atividade antioxidante na eliminação de radicais livres (DUKE, 2017), potencial antimicrobiano do óleo essencial produzido a partir de suas folhas (MEDEIROS et al., 2003) e atividades antimutagênica e antigenotóxica de extratos do fruto, sugerindo que estes podem funcionar como agentes preventivos contra o câncer (NERI-NUMA et al., 2013).

Na medicina popular local, o araçá-boi é utilizado no tratamento de desordens intestinais e urinárias, além de também aliviar os sintomas do resfriado, o que indica que os compostos bioativos presentes na planta podem ter potenciais benefícios para a saúde humana (FERNÁNDEZ-TRUJILLO et al., 2011).

3.5 LEISHMANIA

As leishmanioses são antropozoonoses, causadas por protozoários do gênero *Leishmania*, consideradas um grande problema de saúde pública, representando um conjunto de doenças com grande importância clínica e diversidade epidemiológica (BRASIL, 2007).

A Leishmaniose Tegumentar Americana (LTA) é uma enfermidade causada por protozoários do gênero *Leishmania*, transmitida ao homem pela picada de mosquitos flebotomíneos. Existem atualmente 6 espécies de *Leishmania* responsáveis pela doença humana no Brasil, e mais de 200 espécies de flebotomíneos envolvidos em sua transmissão (BASANO, 2004).

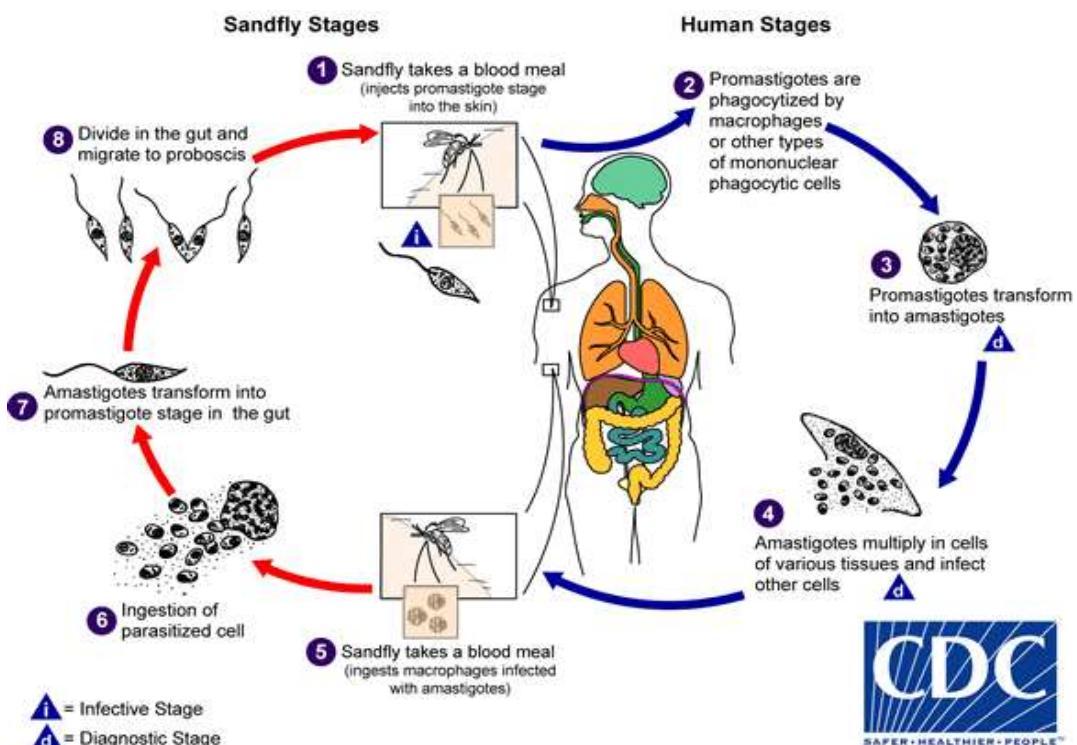
A enfermidade apresenta-se na forma cutânea ou mucosa (PALMEIRO et al., 2007), sendo que a forma mucosa é, na maioria das vezes, secundária às lesões cutâneas, surgindo, geralmente, meses ou anos depois das lesões de pele (BRASIL, 2007). Quando acometem a mucosa bucal, a doença se torna destrutiva ou ulcerovegetativa e granulomatosa, acompanhada pela presença de granulações grosseiras e sulcos profundos. Geralmente, a dificuldade de deglutição e a dor são os principais sintomas relatados, além de sialorréia, odor fétido e o sangramento (GONTIJO; CARVALHO, 2003).

O protozoário *Leishmania infantum chagasi*, no Brasil, é o agente causador da leishmaniose visceral (LV), enfermidade que atinge cerca de 65 países, com incidência estimada de 500 mil novos casos e 59 mil óbitos anuais, e é transmitida por flebotomíneos do gênero *Lutzomyia*, sendo o cão considerado a principal fonte de infecção no meio urbano. Considerada uma doença grave com poucas opções terapêuticas, mesmo quando

adequadamente tratada, a mesma tem letalidade de cerca de 5% (WERNECK, 2010).

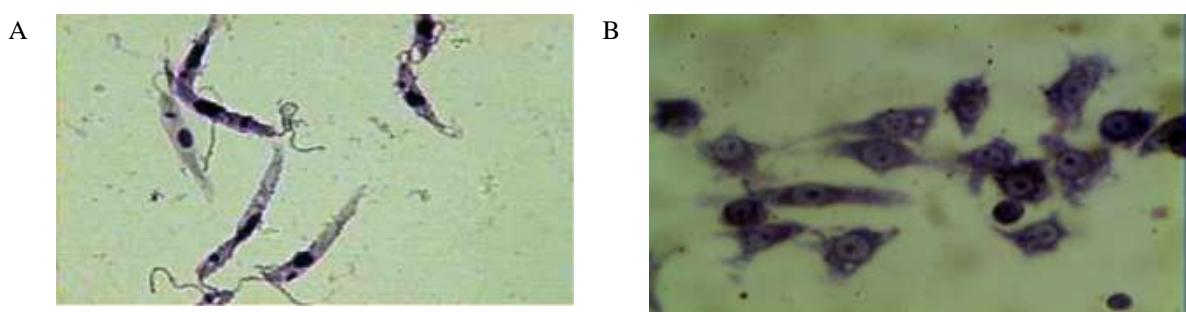
As formas promastigotas metacíclicas de *Leishmania* são responsáveis por causar infecção e são transmitidas aos hospedeiros vertebrados pelos vetores (Figura 3). Além da forma promastigotas metacíclicas, o parasito também apresenta as formas paramastigotas e promastigota não metacíclica, que se multiplicam no trato digestivo do vetor, e as formas amastigotas, que são parasitas obrigatórios das células do sistema fagocítico mononuclear (Figura 4) (ALBERNAZ, 2010).

Figura 3: Ciclo Biológico da *Leishmania*



Fonte: CDC, 2014

Figura 4: A) Forma Promastigota de *Leishmania sp*; B) Forma Amastigota de *Leishmania sp*.



Fonte: VANDESMET, 2014.

No tratamento, além da necessidade do desenvolvimento de melhores fármacos para o tratamento da Leishmaniose, ainda não há vacinas contra o parasita *Leishmania* e, também, não

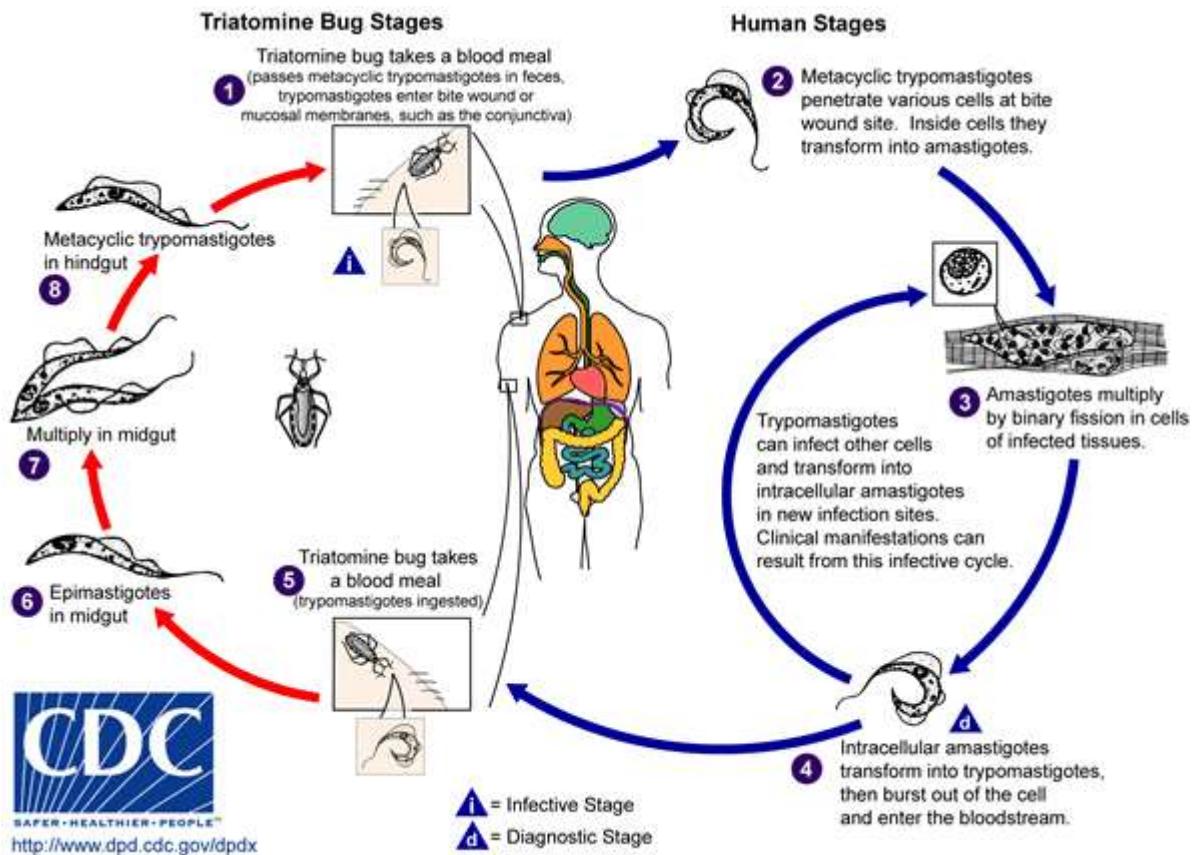
há ações eficazes no controle do vetor. Desta forma, o tratamento utilizado atualmente é baseado na quimioterapia com fármacos caros, tóxicos e ineficientes que não atendem a variedade de agentes etiológicos da doença (REBOLLO; OLIVERO-VERBEL; REYES, 2013).

3.6 *TRYPANOSOMA CRUZI*

Trypanosoma cruzi é o protozoário causador da doença de Chagas, uma infecção sistêmica crônica que atinge cerca de 18 milhões de pessoas na América Latina e 30% dos infectados apresentam a forma sintomática da enfermidade. Os problemas cardíacos são os principais responsáveis pelo óbito dos pacientes chagásicos, por arritmias ventriculares ou disfunção ventricular grave (RASSI JÚNIOR et al., 2006). A infecção por *T. cruzi* pode ser transmitida por um vetor invertebrado, os triatomíneos, conhecidos no Brasil como barbeiros, a principal espécie é o *Triatoma infestans* (DE ARAÚJO-JORGE; DE CASTRO, 2000).

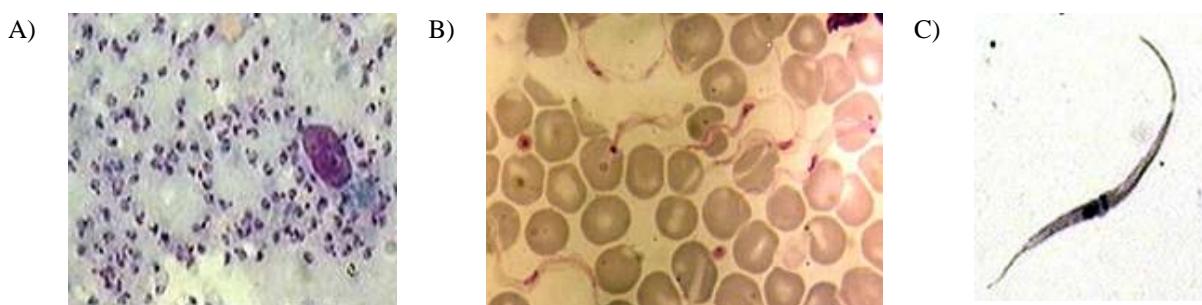
O *Trypanosoma cruzi* possui ciclo de vida complexo que requer passagem obrigatória por um hospedeiro invertebrado, o inseto vetor triatomíneo, e um hospedeiro vertebrado (Figura 5) (BONNEY; ENGMAN, 2010). Durante seu ciclo de vida, o parasito apresenta basicamente três formas evolutivas: 1- epimastigota, forma replicativa não infectante, presente no intestino superior do inseto; 2- tripomastigota, forma não replicativa infectante, presente na porção posterior do intestino do inseto; 3- amastigota, forma replicativa intracelular (Figura 6). As formas são classificadas e diferenciadas a partir da posição do cinetoplasto em relação ao núcleo, flagelo e morfologia (FERRAZ et al., 2007; ABE et al., 2002; ROMERO; MORILLA, 2010).

Figura 5: Ciclo Biológico do *Trypanosoma cruzi*



Fonte: CDC

Figura 6: A: Forma amastigota do *T. cruzi*; B: Forma tripomastigota do *T. cruzi*; C: Forma epimastigota do *T. cruzi*.



Fonte: VANDESMET, 2014

O ciclo no inseto tem início quando durante o repasto sanguíneo no vertebrado infectado, o vetor ingere os tripomastigotas sanguíneos que, no intestino do vetor, serão transformados na forma epimastigota (exclusiva do hospedeiro invertebrado) onde se prolifera e, novamente, transforma-se na forma tripomastigota metacíclica que será eliminada com as fezes durante o repasto sanguíneo, podendo penetrar no organismo do hospedeiro vertebrado por meio da picada ou mucosas (DUTRA et al., 2006).

As formas tripomastigotas metacíclicas são capazes de sobreviver e reproduzir-se em uma variedade de células nucleadas (SILVA et al., 2007), onde se diferenciam em formas amastigotas; ocorre proliferação intracelular por sucessivas divisões binárias e diferenciação em formas tripomastigotas que rompem a célula e caem na circulação sanguínea (LIMA et al., 2010). Após a ruptura celular, as formas tripomastigotas liberadas na corrente sanguínea podem infectar células vizinhas, se espalhar para outros órgãos ou serem ingeridas por outro inseto vetor reiniciando o ciclo (MUÑOZ-SARAIVA et al., 2012).

Após a invasão, a doença de Chagas pode ser classificada em fases aguda e crônica, de acordo com o tempo da infecção e características do soro do vertebrado infectado. Por cerca de 30 a 90 dias, observa-se a fase aguda, sendo uma característica marcante desta fase o grande número de parasitas presentes na circulação, enquanto a fase crônica é caracterizada pela parasitemia baixa e pelo elevado número de imunoglobulinas circulantes. As manifestações observadas nesta fase são decorrentes do efeito intracelular do parasitismo (FERRAZ et al., 2007).

A infecção aguda tende a ser uma doença febril autolimitante leve. Aproximadamente 30% dos indivíduos infectados desenvolvem doença de Chagas crônica, que na maioria dos casos afeta o coração (causando cardiomiopatia e disritmias) ou o intestino (causando mega-esôfago ou mega-côlon). A supressão imune de qualquer causa pode resultar na reativação da infecção latente, causando graves problemas cardíacos e sequelas neurológicas (TEIXEIRA et al., 2006).

Nifurtimox e benznidazol são os medicamentos disponíveis para o tratamento de pacientes infectados com *T. cruzi*. Entretanto, devido à sua conhecida toxicidade e efeito limitado para diferentes isolados de parasitas e fases da doença, a produção de novos fármacos é urgentemente necessária (DUSCHAK; COUTO, 2007; MCKERROW et al., 2009; URBINA, 2009; SOEIRO et al., 2009).

3.7 ÓLEOS ESSENCIAIS

Os óleos essenciais (OE) são misturas complexas de substâncias voláteis e lipofílicas usadas desde tempos antigos por suas propriedades farmacêuticas e como aromatizantes de cosméticos e perfumes. São conhecidos cerca de cerca de 3000 OE, dos quais 300 são comercialmente importantes por suas aplicações agronômicas, alimentícias, farmacêuticas, sanitárias, cosméticas e na perfumaria (BAKKALI et al., 2008; FIGUEIREDO et al., 2008).

Essas substâncias podem ser obtidas de diversos órgãos vegetais, tais como flores,

caules, folhas, galhos, frutas, sementes, raízes, madeira ou casca, e são armazenados em células secretórias, canais, cavidades, células epidérmicas ou tricomas glandulares (NIETO, 2017).

Os OE são produtos do metabolismo secundário vegetal e desempenham importantes papéis protetores, atuando como agentes alelopáticos, antivirais, antifúngicos, antibacterianos, inseticidas, sendo capazes de reduzir o apetite de animais herbívoros. Além disso, possuem função crucial para o processo de polinização, atraindo insetos para favorecer a dispersão de pólens e sementes, e repelindo aqueles indesejáveis. As mais bem estabelecidas atividades de óleos essenciais são antimicrobiana, sedativa, anti-inflamatória, bactericida, antiviral, fungicida e como conservante de alimentos (BAKKALI et al., 2008; NIETO, 2017).

Do ponto de vista químico, os OE são constituídos de misturas de 20-60 compostos orgânicos nas mais diversas concentrações, que podem pertencer a classe dos álcoois, éteres ou óxidos, aldeídos, cetonas, ésteres, aminas, amidas, fenóis, heterociclos e principalmente os terpenos. Além destes, também podem conter compostos não-terpênicos como os derivados de fenilpropanóides (DHIFI et al., 2016).

O fato destes óleos serem naturais e biodegradáveis, aliado a baixa toxicidade em mamíferos, podendo atuar sobre várias moléculas-alvo simultaneamente, torna essas substâncias peças-chave na pesquisa de novos medicamentos. Assim, os óleos podem ser explorados como alternativa ou complemento aos compostos sintéticos utilizados pela indústria farmacêutica, diminuindo ou sem induzir os efeitos secundários indesejáveis (FIGUEIREDO et al., 2008; DHIFI et al., 2016).

3.8 ÓLEOS ESSENCIAIS COM ATIVIDADE ANTIPARASITÁRIA

A ação antiparasitária de plantas medicinais, atualmente, pode ser considerada, ao revisar a literatura, como um dos campos de mais interesse aos pesquisadores, principalmente acerca de OE. Dito isto, se faz necessário citar alguns estudos que comprovam a atividade antiparasitária de OE, como Estevam e colaboradores (2018) que fez análise do OE dos frutos verdes de *Protium ovatum* (*Burseraceae*) que apresentou forte atividade tripanocida contra as formas tripomastigota do *Trypanosoma cruzi* ($IC_{50} = 1,2 \mu\text{g/mL}$), os principais compostos encontrados no OE dos frutos verdes de *P. ovatum* foram: β -mirceno (62,0 %), α -pineno (11,3 %) e limoneno. Carneiro e colaboradores (2017) onde o OE das flores de *Eugenia klotzschiana* (*Myrtaceae*) apresentou promissora atividade tripanocida contra formas tripomastigotas de *Trypanosoma cruzi* ($20,2 \mu\text{g/mL}$) e os principais compostos encontrados no OE das flores foram sesquiterpenoides: β -cariofileno (21,1%), biciclogermacreno (10,2%) e espatulenol (20,9%).

Em especial, o potencial Leishmanicida de OE tem sido descrito na literatura. Por exemplo, Rosa e colaboradores (2003) que relata que o óleo das folhas de *Croton cajucara*, apresentou IC₅₀ de 0,008 µg/mL frente a promastigotas da espécie *L. amazonenses* e Dutra e colaboradores (2009) que estudou o OE e frações (6,25 – 100 µg/mL) obtidas das sementes de *P. emarginatus* que apresentou nas frações hexânica (IC₅₀= 50,06 µg/mL) e butanólica (IC₅₀= 46,65 µg/mL) atividade leishmanicida frente às formas promastigotas de *L. amazonensis*.

**4 ARTIGO 1 - CHEMICAL COMPOSITION AND EVALUATION OF
ANTIPARASITIC ACTIVITY OF THE ESSENTIAL OIL OF *EUGENIA
STIPITATA* MCVAUGH (MYRTACEAE)**

Chemical composition and evaluation of antiparasitic activity of the essential oil of *Eugenia stipitata* Mcvaugh (myrtaceae)

CÍCERO RAMON BEZERRA DOS SANTOS^a, MÁRCIA VANUSA DA SILVA^a, IRWIN ROSE DE ALENCAR MENEZES^c, ALEXANDRE GOMES DA SILVA^b, MARIANA GOMES VIDAL SAMPAIO^a, LILIAN CORTEZ SOMBRA VANDESMET^a, SAULO ALMEIDA DE MENEZES^d, BENEDITO YAGO MACHADO PORTELA^d, EDUARDO BRANDÃO^f, DOUGLAS WILLYAM RODRIGUES GOMES^d, FERNANDO GOMES FIGUEREDO^e, LEANDRO BEZERRA DE LIMA^g, MARIA CELESTE VEJA GOMEZ^h.

^a Biochemistry Departament, Biocience Center, Federal University of Pernambuco – UFPE, Recife, Pernambuco, Brazil.

^b Departament Antibiotics, Biocience Center, Federal University of Pernambuco - (UFPE) – Recife, Pernambuco, Brazil.

^c Pharmacology and Medicinal Chemistry, Regional University of Cariri, Crato, Ceara, Brazil.

^d Universitary Center Católica de Quixadá, Quixadá, Ceará, Brazil.

^e Laboratories of Microbiology and Molecular Biology, Regional University of Cariri, Crato, Ceara, Brazil.

^f Research Center Aggeu Magalhães, Fundação Oswaldo Cruz, Recife, PE, Brasil

^g Laboratory of Biotechnology and Ogenic Synthesis, Universidade Federal do Ceará, Fortaleza, Ceara, Brazil.

^h Centre for the Development of Scientific Investigation - CEDIC, Fundación Moisés Bertoni /Laboratorios Diaz Gill, Asuncion, Paraguay.

*Corresponding Author: Cícero Ramon Bezerra dos Santos. Biochemistry Departament, Biocience Center. Federal University of Pernambuco – UFPE, Recife (PE), Brazil. Address: Av. Prof. Moraes Rego, 1235, Cidade Universitária, CEP: 50.670-901. Phone: +55 (81) 2126.8540

E-mail: ramonsantosbezerra@hotmail.com

ABSTRACT

The essential oils (EOs) of plants used in traditional medicine are known as a rich source of chemically different compounds with different biological activities. In this study, we analyzed the chemical composition and investigated the trypanocidal, leishmanicide, effect and in vitro cytotoxicity of essential oil obtained from the leaves of *Eugenia stipitata* McVaugh. EO was extracted by the distillation method by steam stripping and its chemical composition analyzed by GC/MS and tested in seven concentrations (15.75, 31.25, 62.5, 125, 250, 500 e 1000 µg/mL) against the forms of *Trypanosoma cruzi* epimastigotes and promastigotes forms of *Leishmania braziliensis* and *Leishmania infantum*, as well as against fibroblasts. Generally, 48 components were identified in the oil chemical analysis and the most abundant constituents were: β -eudesmol (15.28%), γ -eudesmol (10.85%), Elemol (10.21%), caryophyllene oxide (6.5%), Clovane (6.18%) and Epatulenol (6.14%). In the evaluation of the parasitic activities, EO of *Eugenia stipitata* McVaugh at concentrations of 125 µg/mL and 62.5 µg/mL inhibited, respectively, 80.69% and 71.91% of the promastigote form of *L. braziliensis*. Similar inhibition was observed for the same concentrations of the oil against *L. infantum*, which inhibited 76.51% of promastigote form of *L. infantum* in concentration of 125 µg/mL and 51.1% in the concentration of 62.5 µg/mL. However, in the concentration of 125 µg/mL a smaller parasitic inhibition was observed of essential oil of *Eugenia stipitata* McVaugh before the epimastigotes forms of *T. cruzi*, with 22.85%. The essential oil of *Eugenia stipitata* McVaugh presented a low cytotoxicity in the concentration of 62.5 µg/mL and no cytotoxicity in concentrations of 31.5 µg/mL and 15.75 µg/mL. Therefore, the studied EO presented itself as a promising antiparasitic source against the ways of life of *L. braziliensis* and *L. infantum*.

Keywords: Essential Oil. Caatinga. Antiprotozoal. *Eugenia stipitata* McVaugh.

1. INTRODUCTION

Essential oils (EOs) are a complex mixture of compounds arising from plants secondary metabolism, which serve as a protective mechanism, acting as antimicrobial , antiviral and antifungal agents (Bakalli et al., 2008; Sharifi-Rad et al., 2017). Since the middle age, EOS has been widely used as a bactericide, virucides, fungicides and pesticides. Furthermore, their use has been shown to be safe, with few side effects and low toxicity to mammalian cells (Bakkali et al., 2008; Dubey et al., 2010; Santoro et al., 2007).

American trypanosomiasis, also known as Chagas disease (CD), is a neglected tropical disease which is Brazil in Latin America. CD was first described in 1909 and is caused by the parasite *Trypanosoma cruzi* (Moncayo; Silveira, 2017). According to the World Health Organization more than 65 million people live at risk of infection and of 6 to 7 million are infected (WHO, 2018a). The transmission occurs by contact with hematophagous triatomine insects of the genus *Triatoma*, *Panstrongylus* and *Rhodnius* infected by the protozoan parasite, by blood transfusion, via the placenta, by eating contaminated food with the flagellate parasite or, less frequently, ingestion of contaminated raw meat, organ transplants from infected donors and sexual contact (Coura, 2015).

There is no vaccine for CD and the treatment for this disease is based on two chemotherapeutic agents: benzonizadole and nifurtimox. These drugs have a low effectiveness during the chronic phase of the disease and high toxicity (Bermudez et al., 2016). Moreover, the absence of a liquid pharmaceutical form leads to a low adherence of the patient, especially for elderly people and children (Bernardes; Zani; Carvalho, 2013). Therefore, it is necessary to seek more effective and less toxic treatment to this disease (De Paula et al., 2015; Pinheiro et al., 2017). In an attempt to develop new drugs, many compounds of natural and synthetic sources, have been tested against *Trypanosoma cruzi* (Cerecetto; González, 2010; González;

Cerecetto, 2011).

The leishmaniasis is caused by parasites of the genus *Leishmania*, also being considered neglected tropical diseases (Arango, Descoteaux, 2014). Leishmaniasis is classified in cutaneous, subcutaneous and visceral forms, affecting 12 million people in 98 countries (Hamzavi et al., 2018; WHO, 2018b). The treatment occurs through the administration of antimonial multipurpose drugs, liposomal amphotericin B and pentamidine, paromomycin, miltefosine, among others; all with high cost and a lot of side effects. Therefore, drugs originating from natural products have been studied (Carneiro et al., 2015; Dias; Dassoy, 2013).

Seasonally dry tropical forests (SDTF) present the circum-Amazonian distribution in South America and have been the main focus of many recent biogeographic and conservation studies (DryFlor et al., 2016). These forests are usually associated with fertile soils and climates marked by highly seasonal rainfall with the severe dry season from three to six months, when most of the vegetation is leafless (Mcleod et al., 2006; DryFlor et al., 2016). SDTFs are patchily distributed in South America in several "nuclei" of which the Caatinga biogeographical province (Morrone, 2014) from Northeastern Brazil is the largest one (Prado; Gibbs, 1993).

Chapada do Araripe, Ceará, Pernambuco and Piauí state, Brazil, harbors a humid forest enclave within the dry forest mosaic of the Caatinga dryland domain. The humid forest grows in an area with relatively abundant water resources provided by many streams and springs, in spite of the long, regional dry season (DNPM, 1996) constituting a type of oasis in the midst of a semiarid region that supports a wide floral and faunal diversity (FLONA, 2004; Silva et al., 2011). The humid forest acts as a refuge for many specific species to that region, including endemic taxa such *Antilophia bokermanni* (the Araripe Soleil Bleu, "soldadinho do Araripe") which is critically threatened with extinction (Auler et al., 2004; Linhares; Silva, 2015), and other taxa with disjunct distributions between the Atlantic and Amazon forests (e.g., spermatophytes, ferns and angiosperms) (Loiola et al., 2015; Collinet et al., 2015) -

illustrating the biological importance of that area and the necessity of its protection (MMA, 2000).

The genus *Eugenia* is well represented in the various phytogeographic domains in Brazil and are constant targets of research, once they are in possession of essential oils and various secondary metabolites which have important biological actions (Romagnolo; Souza, 2006; Victoria et al., 2012). *Eugenia stipitata* McVaugh, popularly known as araçá-boi, is used in folk medicine in the treatment of intestinal and urinary disorders, in addition to relieving the symptoms of cold, which indicates that the bioactive compounds present in the plant may have potential benefits for human health (Fernández-Trujillo et al., 2011).

This study aimed to investigate the trypanocidal, the leishmanicide effect and *in vitro* cytotoxicity, as well as to identify the chemical components present in the essential oil obtained from the leaves of *E. stipitata*.

2 MATERIALS AND METHODS

2.1 Collection Area

The leaves of the species *E. stipitata* were collected in the area of native vegetation, located in Serra dos Paus - Chapada do Araripe, municipality of Exu, Pernambuco ($7^{\circ}21'S$, $39^{\circ}53'W$ and altitude 884m), in the month of May in the year 2017.

2.2 Botanical Material

The plant material was identified by PhD. Maria Arlene Pessoa da Silva of the Laboratory of Microbiology and Molecular Biology, Regional University of Cariri - URCA.

Specimens of the species were produced and deposited at the Herbarium Caririense Dárdano de Andrade Lima of the University aforementioned, under the number 13.054.

2.3 Extraction and chemical composition of the essential oil

To obtain the oil from the leaves of *E. stipitata* (EOESL), the plant material was ground into small pieces and subjected to the distillation method by stream stripping (4500g, 3h). The obtained EO was filtered, weighed and the oil yield was calculated in % (w/w). EO was then stored in a dark bottle and chilled (at + 5 °C) until use.

EO was analyzed through the Model of Gas chromatography-mass spectrometry (GC-MS) Shimadzu GCMS-QP2010 using capillary column Rtx®-5MS (30 m x 0.25 mm x 0.25 µm). The MS operation conditions were optimized in the following way: 70 eV, stream stripping gas (He) 13.6 mL·min⁻¹ and pressure of 53.5 KPa. The following temperature program was used: 100 °C (3 min) to 310 °C (3.5 °C/min) (Karmeling; Verluis; schauer, 1975).

2.4 Cell lines used

For the in-vitro tests with *T. cruzi*, CL-B5 clone was used (Buckner et al., 1996). The parasites stably transected with the gene for β-galactosidase of *Escherichia coli* (lacZ) were provided by PhD. F. Buckner through Commemorative Gorgas Institute (Panama). Epimastigotes forms were cultured at 28 °C in Liver Infusion Tryptose Broth (Difco, Detroit, MI) supplemented with 10% Fetal Bovine Serum (FBS) (Gibco, Carlsbad, CA), penicillin (Ern, SA, Barcelona, Spain) and streptomycin (Reig Jofr SA, Barcelona, Spain) as described by Le Senne et al. (2002). The cells were collected for testing in the exponential growth phase. Cultures of *Leishmania* spp. were obtained from the Health Science Research Institute,

Asunción, Paraguay - IICS and identified by isoenzyme analysis. The lines maintenance, forms of cultivation and isolation of promastigotes forms of *Leishmania* spp. followed procedures described by Roldos et al. (2008). The inhibitory action of these promastigotes forms were performed using the *L. braziliensis* (MHOM / CO / 88 / UA301) and *L. infantum* lines (MHOM/ES/92/BCN83), cultured at 22 °C in Schneider's Drosophila supplemented with 20% FBS. In the cytotoxicity assays, the NCTC-929 murine fibroblast line, obtained from the National Collection of Type Cultures and cultivated in Minimal Essential Medium, was used. The culture medium was supplemented with heat inactivated FBS (10%), penicillin G (100 U/mL) and streptomycin (100 mg/mL). Cultures were maintained at 37 °C in a humidified atmosphere with 5% CO². The lines viability was evaluated through the use of the resazurin colorimetric method (Rolón et al., 2006).

2.5 Reagents

Resazurin sodium was obtained from Sigma-Aldrich (St Louis, MO) and stored at 4 °C away from light. The Resazurin solution was prepared with 1% phosphate buffer, pH 7 and sterilized by filtration before use. Chlorophenol red-β-D galactopyranoside (CPRG; Roche, Indianapolis, IN) was dissolved in a 0.9% Triton X-100 (pH 7.4) solution. Penicillin G (Ern, S.A., Barcelona, Spain), streptomycin (Reig Jofré S.A., Barcelona, Spain) and dimethyl sulfoxide (DMSO) were also used.

2.6 Trypanocidal activity

For the trypanocidal assays, 96-well microdilution plates (Sarstedt, Sarstedt, Inc.) were used with cultures that did not reach the stationary phase, as described by Vega et al. (2005).

The *T. cruzi* epimastigote forms were seeded at 1×10^5 per millilitre in 200 μ L and the plates were incubated with EOESL at the 15.75, 31.25, 62.5, 125, 250, 500 and 1000 μ g/ mL concentrations at 28°C for 72h, with 50 μ L of the CPRG solution to give the final concentration of 200 μ M. The plates were incubated at 37°C for an additional 6h period and then read at 595nm in a spectrophotometer.

Benznidazole was the reference drug tested in triplicates at 1, 10, 50 and 100 μ g/mL concentrations. Each experiment was performed twice separately. The antiepimastigote percentage (%AE) was calculated following the formula: %AE= [(AE - AEB)/(AC - ACB)] x 100, where AE=absorbance of the experimental group; AEB=blank compound; AC=absorbance of the control group; ACB=culture medium blank. The natural product solution to be analyzed was diluted in dimethyl sulfoxide, with the final concentration DMSO never exceeding 0.2% of the final solvent.

2.7 Leishmanicidal activity

The method used in this assay was developed by Mikus; Steverding (2000) and modified, where the promastigote cultures (2.5×10^5 parasites/well) were grown in microdilution plates. The EOESL was dissolved in DMSO at 15.75, 31.25, 62.5, 125, 250, 500, 1000 μ g/mL concentrations. After an incubation period of 48h at 26°C, 20 μ l of resazurin solution was added and the oxidation was quantified in a spectrophotometer with 570–595nm wavelengths. The concentrations were tested in triplicates along with miltefosine which was used as a positive control. The antipromastigote percent (%AP) was calculated following the formula %AP = $((A_{570} X 117,216 - A_{595} X 80,586) / (A_{570} X 117,216 - A_{595} X 80,586))$ of test compounds / $((A_{570} X 117,216 - A_{595} X 80,586) / (A_{570} X 117,216 - A_{595} X 80,586))$ of untreated posite growth control))X100. Thus testing the substances efficacy.

2.8. Cytotoxicity test

The clones were seeded (3×10^4) in 96-well flat-bottom microdilution plates with 100 μ L culture medium. RPMI 1640 per well. Cells were grown overnight at 37°C and 5% CO₂ atmosphere. The medium was then replaced with the isolated tested substance and added at different concentrations in 200 μ L of the medium for 24h. Growth control wells were also included. After the incubation, 20 μ L of Resazurin solution was added to each well. The plates were incubated again for 3h; The Resazurin reduction was determined by wavelength absorbance measurement at 570 and 595 nm in a microplate reader.

Each concentration of EOESL (15.75, 31.25, 62.5, 125, 250, 500, 1000 μ g/mL) was tested three times. The percent cytotoxicity (%C) of the natural products was determined as follows: $\%C = [(A_{570} \times 117,216 - A_{595} \times 80,586) \text{ test samples} / (A_{570} \times 117,216 - A_{595} \times 80,586) \text{ control}] \times 100$, where A₅₇₀ and A₅₉₅ represented the values 80,586 and 117,216 of optical media density at 570 and 595nm, respectively, recorded for wells with cells containing different natural product doses or the value recorded for wells with cells and without natural product(respectively positive growth controls)(Rolón et al., 2006). Then, the Selectivity Index (SI) was calculated as the 50 inhibitory concentration rate (IC₅₀) of the cells / 50 inhibitory concentration (IC₅₀) of parasites. The value resulting from this calculation reflects how many times the used EO is more selective to parasites than the host cell.

2.9 Statistical analysis

Statistical analysis was performed using ANOVA. The data were analyzed by GraphPad Prism 5.0 program (GraphPad Software, San Diego, CA, USA).

3 RESULTS

The fresh leaves of *E. stipitata* provided oil yield of 0.13%. Table 1 presents the chemical composition of the essential oil (EO) analyzed by GC/MS. In the EO of *Eugenia stipitata* leaves 46 compounds were identified, corresponding to 100% of the total oil, which is composed by 45.83% of sesquiterpene hydrocarbons, 28.26% of oxygenated sesquiterpenes, 15.22% of oxygenated monoterpenes, 4.35% of monoterpenes hydrocarbons and 4.35% other elements. The most abundant constituents of the analyzed EO were β -eudesmol (15.28%), γ -eudesmol (10.85%), Elemol (10.21%), caryophyllene oxide (6.5%), Clovane (6.18%) and Espatulenol (6.14%).

Table 1. Chemical composition of the essential oil from *Eugenia stipitata* McVaugh

Peak	RT (min)	Compound	Area (%)
1	4.440	β -Pinene	0.41
2	5.140	o-Cymene	0.12
3	5.315	Eucalyptol	2.61
4	6.451	Linalool	0.24
5	7.465	Pinocarveol	0.49
6	8.274	Terpinen-4-ol	1.51
7	8.558	α -Terpineol	2.62
8	8.717	Myrtenol	0.40
9	8.770	Myrtenal	0.26
10	11.147	(E)-Pinocarvyl acetate	0.24
11	12.076	δ -Elemene	0.53

12	12.960	Ylangene	0.19
13	13.077	α -Cubebene	0.95
14	13.330	β -Bourbonene	1.99
15	13.431	β -Elemene	1.09
16	14.028	α -Chamigrene	0.49
17	14.204	Caryophyllene	4.38
18	14.280	β -Cedrene	4.33
19	14.460	β -Cubebene	4.19
20	14.547	α -Amorphene	0.86
21	14.674	Aromadendrene	0.45
22	14.779	γ -Cadinene	4.03
23	15.026	1,4,7,-Cycloundecatriene, 1,5,9,9-tetramethyl-, <i>Z,Z,Z</i> -	0.76
24	15.211	Espatulenol	6.14
25	15.818	β -Selinene	0.51
26	15.870	Anthracene, 1,2,3,4,5,6,7,8-octahydro-2-methyl, (R)-	0.24
27	16.012	Ledene	0.79
28	16.074	α -Murolene	0.92
29	16.189	β -Bisabolene	0.70
30	16.629	Cadina-1,3,5-triene	2.08
31	16.974	α -Cadinene	0.52
32	17.126	α -Calacorene	0.31
33	10.21	Elemol	10.21
34	17.392	Alloaromadendrene oxide-(1)	1.06

35	17.761	Viridiflorol	1.56
36	18.152	Caryophyllene oxide	6.50
37	18.589	Epiglobulol	0.81
38	18.812	Cubenol	0.48
39	18.969	γ -Eudesmol	10.85
40	19.118	8-Cedren-13-ol	1.20
41	19.389	tau-Cadinol	2.53
42	19.688	β -Eudesmol	15.28
43	19.730	Clovene	6.18
44	20.063	β -Caryophyllene oxide	0.15
45	20.421	Xenitorin A	0.29
1s,4R,7R,11R-1,3,4,7-			
46	25,458	Tetramethyltricyclo[5.3.1.0(4,11)]undec-2-en-8-one	0.18
Total			100.00

RT: retention time (minutes); Area (%): percentage of compound on the sample

In the evaluation of the antiparasitic activities, OEESL at concentrations of 125 μ g/mL and 62.5 μ g/mL inhibited, respectively, 80.69% and 71.91% of the promastigote form of *L. braziliensis*. Similar inhibition was observed for the same concentrations of the oil against *L. infantum*, which inhibited 76.51% of promastigote form of *L. infantum* in concentration of 125 μ g/mL and 51.1% in the concentration of 62.5 μ g/mL. However, in the concentration of 125 μ g/mL a smaller parasitic inhibition of EOESL was observed for the epimastigotes forms of *T. cruzi*, with 22.85%. EOESL presented a low cytotoxicity in the concentration of 62.5 μ g/mL and no cytotoxicity in concentrations of 31.5 μ g/mL and 15.75 μ g/mL. At the dose of 125 μ g/mL toxicity of 25.73% was observed, staying on the margin the maximum tolerated

cytotoxicity to mammalian cells (25%). The cytotoxicity against mammalian cells was compared to those of parasite by determining the SI: *L. braziliensis*, SI= 13.59; *L. infantum*, SI= 9.35; *T. cruzi*, SI= 0.50.

It is noteworthy that in relation to the parasitic activities that presented results less than or equal to 20% of inhibition, the results were not considered because they did not have important biological activities.

Table 2: Anti-Leishmani, trypanocidal activity and cytotoxicity of the essential oil of *Eugenia stipitata*.

Natural Product	Conc. ($\mu\text{g/mL}$)	% AP <i>L.braziliensis</i>	% AP <i>L.infantum</i>	% AE <i>T. cruzi</i>	%C
LC50 ($\mu\text{g/mL}$)		37.65	54.71	1019	511.5
	1000	91.04 \pm 0.64	84.80 \pm 0.45	41.46 \pm 0.28	*
	500	88.39 \pm 0.39	84.69 \pm 0.01	30.85 \pm 0.53	*
	250	87.61 \pm 0.51	83.82 \pm 0.42	27.97 \pm 0.81	*
EOESL	125	80.69 \pm 0.75	76.51 \pm 0.32	22.85 \pm 0.70	25.73 \pm 2.69
	62.5	71.91 \pm 0.14	73.91 \pm 0.15	-	5.09 \pm 0.34
	31.25	41.40 \pm 0.42	27.71 \pm 8.15	-	0.00 \pm 1.12
	15.75	22.17 \pm 0.99	0.00 \pm 0.14	-	0.00 \pm 0.95

EOESL – essential oil from *Eugenia stipitata* McVaugh; SI – selectivity index; Conc. - Concentration; $\mu\text{g/mL}$ - micrograms per milliliters; % AP - Percent Antipromastigote; % AE- Percent Antiepimastigote; % C - Percent of Cytotoxicity in Fibroblasts; * - No determined and \pm - Standard Deviation.

Table 3 presents the results for the positive control of benznidazole. At a concentration of 100 µg/mL, the drug inhibited 96.59% of forms of *T. cruzi* epimastigotes and caused the death of 1.43% of the fibroblasts.

Table 3: Positive control of benzonidazole against *Trypanosoma cruzi* (µg/mL)

Drug	Conc. (µg/mL)	% AE (<i>T. cruzi</i>)
Benzomidazole	100	96.59±2.13
	50	81.84±2.74
	10	50.61±5.83
	1	10.53±5.87

Conc. - Concentration; µg/mL - micrograms per milliliters; % AE - Percent Antiepimastigote and ± - Standard Deviation.

Table 4 presents the results for the positive control of Miltefosine, which behaved differently before the two species of *Leishmania*. For *L. infantum*, the concentration of 128 µg/mL had the largest percentage inhibition of parasites, however, caused the death of 2.42% of the fibroblasts. In relation to *L. brasiliensis*, the concentration of 64 µg/mL of miltefosine had the highest rate of inhibition, 83.69% of the promastigotes forms without registration of fibroblasts deaths.

Table 4: Positive control of miltefosine against *Leishmania brasiliensis* and *Leishmania infantum* (µM).

Drug	Conc. (µg/mL)	% AP <i>L.braziliensis</i>	% AP <i>L.infantum</i>

Miltefosine	256	97.07±0.41	80.45±0.27
	128	98.75±0.28	81.64±0.19
	64	98.65±0.21	83.69±0.32
	36	95.43±0.79	83.22±0.22
	16	50.49±0.70	73.69±0.03
	8	1.57±0.80	46.45±0.14

µM- Micromols; % AP - Percent Antipromastigote; ± - Standard Deviation; Conc. - Concentration; µg/mL- Micrograms per microliter.

4 DISCUSSION

The analysis of chemical composition performed by GC/MS, of essential oil of *E. stipitata* leaves , collected in São Miguel Island, in Portugal, identified that the main components of the oil were monoterpenes and sesquiterpenes, as α -pinene (14.1%), β -caryophyllene (22.7%), caryophyllene oxide (15.4%) (Medeiros et al., 2003). In another study, the analysis of the essential oil from *E. stipitata* leaves of the city of Macas, in Ecuador, revealed the presence of 30 compounds, of which γ -Murolene, E-Caryophyllene and δ -Cadinene were the majoritarian (Ronquillo; Galarza, 2016). The mentioned EOs chemical analysis showed distinct chemical elements or the same element in different concentrations of those observed in the present study, as shown in Table 1. According to Douglas et al. (2004), variations in the EO chemical composition of the same species from distinct regions can be attributed to differences in the climatic and geographical parameters, such as temperature, altitude, wind direction, rainfall, soil type, etc.

The chemical analysis of volatile compounds from the fruit of *E. stipitata* collected in the city of Manaus, was characterized by the presence of complex sesquiterpenes and the main

majoritarian elements were germacrene D (37.65%), β -pinene (12.2%) and α -pinene (10.4%) (Franco; Shibamoto, 2000). According to Garzón et al. (2012) this fruit has important antioxidant activity. This analysis showed different chemical composition of the oil composition of the leaves of this study.

The EO analysis of *Eugenia hiemalis* Cambess leaves collected in Blumenau, Santa Catarina, in four different seasons of the year, revealed that the elements espatulenol, δ -cadinene, Bicyclogermacrene and β -caryophyllene were among the four main compounds present in at least three of the tations analyzed (Zatelli et al., 2016). Similar results were observed in the EO of *Eugenia brejoensis* Mazine leaves collected in Catimbau, Pernambuco, whose majoritarian constituents were δ -cadinene, β -caryophyllene and bicyclogermacrene (Silva, 2015). Souza et al. (2017) verified the presence of δ -cadinene, trans-caryophyllene and α -Muurolol as majority elements of EO of leaves of the species *Eugenia brejoensis*.

The composition of the oil of the *Eugenia natalitia* leaves, collected in the Southern Africa, was dominated by oxygenated sesquiterpenes (54.4%), consisting mainly of selina-1,3,7 (11)-trien-8-one (27.5%) and oxidoselina-1,3,7 (11)-trien-8-one (21.9%). The main constituents of a class of compounds of hydrocarbons monoterpenes (33.0%) were α -phellandrene (13.0%) and sabinene (9.7%) (Lawal et al., 2016). While the EO of *Eugenia platysema* leaves had in diterpnes the main class of identified compounds (66.05%), followed by the sesquiterpenes (32.95%). No monoterpenes were found in EO, different from other species of the genus Eugenia (Tenfen et al., 2016).

Sesquiterpenes predominate in OE composition of *Eugenia klotzschiana* leaves, collected in Goiás. The following main terpenic components were identified: α -copaeno (10.6%), β -Bisabolene (17.4%), α - (E) -bergamottin (29.9%) and germacrene D (13.3%) (Carneiro et al., 2017). The OE obtained from the leaves of *Eugenia uniflora*, collected in the vicinity of the plant leaves this research shows a high percentage of isoflurane-Germacrene

(65.80%), followed by germacra-3,7,9-trien-6-one (16.19%), β -elemenona (4.47%), γ -elemeno (3.97%), Germacrene B (2.19%) and (Z)- β -elemeno (0.39%) (Pereira et al., 2017). Sesquiterpenos also predominated in the composition of the essential oil of the leaves of *Eugenia stipitata* of this research.

There are many biological properties attributed to EO. Because of the chemical complexity, it is difficult to correlate these properties to a single substance, since that the same may be determined by the chemical synergism (Chaieb et al., 2007). The synergism of the main chemical EO constituents or the presence of other constituents which may also be active at even lower concentrations is an important factor that can lead to good performance of antiparasitic activity (Melo et al., 2011). The EO activity in this study is probably due to the presence of sesquiterpenes (77.08%), whose biological activities are already well known (Sauter et al., 2011). The β -caryophyllene (a sesquiterpenic hydrocarbon) and the nerolidol (a oxygenated sesquiterpene) are examples of terpenic substances with well characterized anti-*Leishmania* activity, possibly related to the inhibition of the biosynthesis of isoprenoid cells (Arruda et al., 2005; Santos et al., 2008). Two sesquiterpene lactones were isolated from the organic extract of *Ambrosia teunifolia* leaves, both presenting *in vitro* trypanocidal activity against epimastigote forms of *T. cruzi* and promastigotes forms of *Leishmania* sp. (Süsen et al., 2008).

Of the majoritarian components, β -eudesmol has important anticancer activity observed in different animal models (Kotawong et al., 2018) and antibacterial activity (Bankova; Trusheva, 2014); Elemol can be considered as one of the main EO antimicrobial components (Mevy et al., 2006). β -eudesmol (51.9%) and γ -eudesmol (18.9%) were the majoritarian EO elements of *Guatteria friesiana*, plant with excellent antiparasitic activity against *Plasmodium falciparum* and different forms of *T. cruzi* (Meira et al., 2016). The EO chemical composition of *Alpinia zerumbet* presents caryophyllene oxide (18.0%) and β -eudesmol (8.9%) and a good

antiparasitic activity (Mendiola et al., 2015). Caryophyllene oxide was also isolated in different species of *Lippia spp*, known by the relevant antiparasitic activity (Escobar et al., 2010).

Phytotherapy has become a promising alternative in the parasitic diseases treatment, especially when it is a question of leishmaniasis, since the existing chemotherapy is considered inadequate due to the high drugs toxicity, to the high costs and the growing resistance of pests to these drugs (González; Cerecetto, 2011; Tasdemir et al., 2006). It is likely that the multicomponent nature of EOs may reduce the occurrence of resistance to them, because multiple targets need to adapt to hinder their action (Yap et al., 2014).

This is the first report of antiparasitic activity of *Eugenia stipitata* and essential oils from the genus Eugenia are little explored regarding this activity.

The ethanolic extract of *Eugenia uniflora* L. leaves collected in Campo Grande, Mato Grosso do Sul, showed no leishmanicide activity on the promastigote forms of *L. amazonensis* (Ribeiro et al., 2014). Studies of Braga et al. (2007) with ethanolic extract of *Eugenia uniflora* L leaves, collected in Juiz de Fora, Minas Gerais, verified the inhibition on the growth of 48.33% of the promastigote forms of *L. amazonensis* and *L. chagasi* in the extract concentrations greater than 250 µg/mL for both parasites. According to Croft et al. (2006), different results may be due to the type of solvent used, moreover, it must be taken into account the genomic difference among the used strains.

The essential oil of the *Eugenia uniflora* L. leaves, collected in São Luís, in Maranhão, showed a significant reduction dependent on the concentration on the parasite viability *L. amazonensis*, with 100% of the promastigote growth inhibition at concentrations of 400, 200 and 100 g/mL ($IC_{50}= 1.75\text{g/mL}$). In the determination of cytotoxicity, the oil was 20 times more toxic for amastigotes than for murine macrophages. The hemolytic activity was 63.22% in the highest tested concentration (400g/mL); however, showed no toxicity to $50\text{g}\cdot\text{mL}^{-1}$ (Rodrigues et al., 2013). The cytotoxic activity evaluation of natural products before the

mammalian cells is an essential target in search of active compounds with biological activity (Santos et al., 2008).

It has been demonstrated that the EO lipophilic components can affect layers of polysaccharides, fatty acids and phospholipids in plasma membranes of promastigotes forms of *Leishmania spp.*, inducing cell lysis and the release of macromolecules (Di Pasqua et al., 2007). In the cytoplasm, these substances can break down specific metabolic pathways of lipids and proteins or stimulate the mitochondrial membrane depolarization, leading to cell necrosis or apoptosis (Armstrong, 2006; Tarikou et al., 2010).

Concerning Chagas disease, the current treatment is based on the use of nifurtimox and benznidazole, very toxic drugs to patients with various side effects (Marin-Neto et al., 2009; Urbina, 2002). Thus, the development of new safer and more effective therapeutic agents are necessary.

Borges et al. (2012) proposes that the EOs trypanocidal activity occurs mainly by their terpene composition, responsible for the hydrophobic character of essential oils, which allows their diffusion through the parasite cellular membrane, affecting the metabolic pathways and intracellular organelles.

The ethanolic extract of *Eugenia uniflora* L. leaves showed 80% inhibition of *T. cruzi* epimastigotes forms at a concentration of 100 µg/mL. Low cytotoxicity of this extract against J774 macrophages was observed, 8% in the concentration of 100 µg/mL, and no toxicity in the concentration of 10 µg/mL. Nifurtimox was used as positive control and, in the highest tested concentration, 10 µg/mL inhibited 89.1% of epimastigotes forms (Santos et al., 2012a).

The trypanocidal activity of ethanolic extract of *Eugenia jambolana* L. leaves, harvested in the city of Crato, Ceará, showed activity against the CL-B5 strain of *T. cruzi*, with 100% of inhibition at a concentration of 100 µg/mL. However, it showed cytotoxicity of 37% in the same concentration (Santos et al., 2012b).

Of the six species of Caatinga plants from different families analyzed by Souza et al. (2017), EO of *Eugenia brejoensis* leaves had the best dose-dependent inhibitory effect on the epimastigotes , trypomastigote and amastigote forms of *T. cruzi* .

According to IS, EO was more selective for the promastigotes forms of *L. braziliensis* (IS=13.59) and *L.infantum* (IS=9.35), and not selective for forms of *T. cruzi* (IS=0.50). The higher the selectivity index the more efficient and less toxic the EO used (Azevedo, 2013).

5. Conclusion

The study revealed that the EO from *Eugenia stipitata* leaves showed no relevant antiparasitic effect on the epimastigotes forms of *T. cruzi*. However, presented antiparasitic effect against the ways of life of *L. braziliensis* and *L. infantum*. Probably, the majoritarian compounds of the analyzed EO are responsible for the observed effect, however additional tests are required to elucidate their action mechanisms.

REFERENCES

- Auler, A.S., Wang, X., Edwards, R.L., 2004. Palaeoenvironments in semi-arid northeastern Brazil inferred from high precision mass spectrometric speleothem and travertine ages and the dynamics of South American rainforests. *Speleogenesis and Evolution of Karst Aquifers*. 2, 1-4.
- Arango, D.G., Descoteaux, A., 2014. Macrophage cytokines: involvement in immunity and infectious diseases. *Frontiers in Immunology*. 5, 491. <https://doi.org/10.3389/fimmu.2014.00491>
- Armstrong, J.S., 2006. Mitochondrial membrane permeabilization: the sine qua non for cell death. *BioEssays*. 28, 253–260. DOI: <https://doi.org/10.1002/bies.20370>
- Arruda, D.C., D'Alexandri ,F.L., Katzin, A. M., Uliana, S.R.B., 2005. Antileishmanial activity of the terpene nerolidol. *Antimicrobial Agents and Chemotherapy*. 49, 1679–1687.
- Azevedo, M.O.A., 2013. Efeito inibitório de óleos essenciais sobre *Trypanosoma cruzi*. Dissertação (Mestrado em biologia celular e molecular) – Universidade Federal do Paraná – Brasil.
- Bakkali, F., Averbeck, S., Averbeck, D., Idaomar, M., 2008. Biological effects of essential oils: a review. *Food and Chemical Toxicology*. 46, 446–475. <https://doi.org/10.1016/j.fct.2007.09.106>
- Bermudez, J., Davies, C., Simonazzi, A., Real, J. P., Palma, S., 2016. Current drug therapy and pharmaceutical challenges for Chagas disease. *Acta Tropica*. 156, 1-16. <https://doi.org/10.1016/j.actatropica.2015.12.017>
- Bernardes, L.S.C.; Zani, C.L., Carvalho, I., 2013. Trypanosomatidae diseases: from the current therapy to the efficacious role of trypanothione reductase in drug discovery. *Current Medicinal Chemistry*. 20, 2673–2696. <https://doi.org/10.2174/0929867311320210005>
- Borges, A.R., Aires, J.R., Higino, T.M., de Medeiros, Md., Citó, A.M., Lopes, J.A., de Figueiredo, R.C., 2012. Trypanocidal and cytotoxic activities of essential oils from medicinal plants of northeast of Brazil. *Experimental Parasitology*. 132, 123-128. <https://doi.org/10.1016/j.exppara.2012.06.003>
- Buckner, F.S., Verlinde, C.L., La Flamme, A.C., Van Voorhis, W.C., 1996. Efficient technique for screening drugs for activity against *Trypanosoma cruzi* using parasites expressing beta-galactosidase. *Antimicrobial Agents and Chemotherapy*. 40, 2592–2597.
- Braga, F.G., Bouzada, M.L.M., Fabri, R.L., Matos, M.O., Moreira, F.O., Scio, E., Coimbra, E.S., 2007. Antileishmanial and antifungal activity of plants used in tradicional medicine in Brazil. *Science Direct*. 111, 396-402. <https://doi.org/10.1016/j.jep.2006.12.006>
- Carneiro, J.N.P., Albuquerque, R.S., Leite, N. F., Machado, A.J.T., Brito, D.I.V., Rólon, M., Veja, C., Coutinho, H.D.M., Morais-Braga, M.F.B., 2015. Avaliação da atividade tripanocida, leishmanicida e citotóxica do geraniol e citronelal. *Caderno de Cultura e Ciência*. 13, 29–36. <https://doi.org/10.14295/cad.cult.cienc.v13i2.841>.

- Carneiro, N.S., Alves, C.C.F., Alves, J.M., Egea, M.B., Martins, C.H.G., Silva, T.S., Bretanha, L.C., Balleste, M.P., Micke, G.A., Silveira, E.V., Miranda, M.L.D., 2017. Chemical composition, antioxidant and antibacterial activities of essential oils from leaves and flowers of *Eugenia klotzschiana* Berg (Myrtaceae). Análises da academia brasileira de ciência. 89. <http://dx.doi.org/10.1590/0001-3765201720160652>
- Cerecetto, H., González, M., 2010. Synthetic medicinal chemistry in Chagas' disease: compounds at the final stage of "Hit-to-Lead" phase. *Pharmaceuticals.* 3, 810-838. DOI: 10.3390/ph3040810
- Coura, J.R., 2015. The main sceneries of Chagas disease transmission. The vectors, blood and oral transmissions - a comprehensive review. *Memorial do Instituto Oswaldo Cruz.* 110, 277–282. <https://doi.org/10.1590/0074-0276140362>.
- Chaieb, K., Hajlaoui, H., Zmantar, T., Kahla-Nakbi, A., Rouabchia, M., Mahdouani, K., Bakhrouf, A., 2007. The chemical composition and biological activity of clove essential oil, *Eugenia caryophyllata* (Syzigium aromaticum L. Myrtaceae): a short review. *Phytotherap.* 506, 501–506. <https://doi.org/10.1002/ptr.2124>.
- Croft, S.L., Sundar, S., Fairlamb, A.H., 2006. Drug resistance in leishmaniasis. *Clin. Microbiol. Rev.* 19, 111–126. <http://dx.doi.org/10.1128/CMR.19.1.111-126.2006>.
- De Paula, J.C., Desoti, V.C., Sampiron, E.G., Martins, S.C., Ueda-Nakamura, S., Biando, E. M., Silva, S.O., Oliveira, G.G., Nakamura, C.V., 2015. Trypanocidal activity of organic extracts from the brazilian and spanish marine sponges. *Revista Brasileira de Farmacognosia.* 6, 651–656. <http://dx.doi.org/10.1016/j.bjp.2015.08.011>
- Di Pasqua, R., Betts, G., Hoskins, N., Edwards, M., Ercolini, D., Mauriello, G., 2007. Membrane toxicity of antimicrobial compounds from essential oils. *Journal of Agricultural and Food Chemistry.* 55, 4863–4870. DOI: 10.1021/jf0636465
- Dias, L.C.; Dassoy, M.A., 2013. Doenças tropicais negligenciadas: uma nova era de desafios e oportunidades. *Revista Química Nova.* 36, 1552–1556, 2013. <https://doi.org/10.1590/S010040422013001000011>.
- DNPM - Departamento Nacional de Produção Mineral. 1996. Projeto Avaliação Hidrológica da Bacia Sedimentar do Araripe. Recife, DNPM.
- Douglas, M.H., Van Klink, J.W., Smallfield, B.M., Perry, N.B., Anderson, R.E., Johnstone, P., Weavers, R.T., 2004. Essential oils from New Zealand manuka: triketone and other chemotypes of *Leptospermum scoparium*. *Phytochemistry.* 65, 1255-1264. <https://doi.org/10.1016/j.phytochem.2004.03.019>
- DryFlor, Banda-R., K., Delgado-Salinas, A.G., Dexter, K.G., Linares-Palomino, R., Oliveira-Filho, A., Prado, D., Pullan, M., Quintana, C., Riina, R., Rodríguez M., G.M., Weinritt, J., Acevedo-Rodríguez, P., Adarve, J., Álvarez, E., Aranguren B., A., Arteaga, J.C., Aymard, G., Castaño, A., Ceballos-Mago, N., Cogollo, Á., Cuadros, H., Delgado, F., Devia, W., Dueñas, H., Fajardo, L., Fernández, Á., Fernández, M.Á., Franklin, J., Freid, E.H., Galetti, L.A., Gonto, R., González-M., R., Graveson, R., Helmer, E.H., Idárraga, Á., López, R., Marcano-

- Veja, H., Martínez, O.G., Maturo, H.M., McDonald, M., McLaren, K., Melo, O., Mijares, F., Mogni, V., Molina, D., Moreno, N. del P., Nassar, J.M., Neves, D.M., Oakley, L.J., Oatham, M., Olvera-Luna, A.R., Pezzini, F.F., Dominguez, O.J.R., Ríos, M.E., Rivera, O., Rodríguez, N., Rojas, A., Särkinen, T., Sánchez, R., Smith, M., Vargas, C., Villanueva, B., Pennington, R.T., 2016. Plant diversity patterns in neotropical dry forests and their conservation implications. *Science*. 353, 1383-1387. DOI:10.1126/science.aaf5080
- Dubey, N. K., Shukla, R., Kumar, A., Singh, P., Prakash, B., 2010. Prospectives of botanical pesticides in sustainable agriculture. *Current Science*. 98, 479-480.
- Escobar, P., Leal, S.M., Herrera, L.V., Martinez, J.R., Stashenko, E., 2010. Chemical composition and antiprotozoal activities of Colombian *Lippia* spp essential oils and their major components. *Mem Inst Oswaldo Cruz*. 105, 184-190. <http://dx.doi.org/10.1590/S0074-02762010000200013>
- Fernández-Trujillo, J.P., Hernández, M.S., Carrillo, M., Barrera, J., 2011. Arazá (*Eugenia stipitata* McVaugh). In: YAHIA, E. M. (Org.). Postharvest biology and technology of tropical and subtropical fruits. England: Woodhead Publishing. 1, 98-117.
- FLONA - Floresta Nacional Araripe. 2004. Plano de Manejo, Sumário executivo da Floresta Nacional Araripe. Brasília, ICMBio.
<http://www.icmbio.gov.br/portal/unidadesdeconservacao/biomas-brasileiros> . 08 April 2018.
- Franco, M.R.B., Shibamoto, T., 2000. Volatile composition of some Brazilian fruits: umbu-caja (*Spondias citherea*), camu-camu (*Myrciaria dubia*), araca-boi (*Eugenia stipitata*), and cupuaçu (*Theobroma grandiflorum*). *Journal of Agricultural and Food Chemistry*. 48, 1263-1265. <https://doi.org/10.1021/jf9900074>
- Garzón, G.A., Narvaéz-Cuenca, C-E., Kopec, R.E., Barry, A.M., Rield, K.M., Schwartz, S.J., 2012. Determination of Carotenoids, Total Phenolic Content, and Antioxidant Activity of Arazá (*Eugenia stipitata* McVaugh), an Amazonian Fruit. *J Agric Food Chem*. 60, 4709-4717. <https://doi.org/10.1021/jf205347f>
- González, M., Cerecetto, H., 2011. Novel compounds to combat trypanosomatid infections: a medicinal chemical perspective. *Expert Opinion on Therapeutic Patents*. 21, 699-715. <https://doi.org/10.1517/13543776.2011.565334>
- Hamzavi, S.S., Dashti, A.S., Kadivar, M.R., Pouladfar, G., Pourabbas, B., 2018. Successful treatment of disseminated cutaneous leishmaniasis with liposomal amphotericin B and miltefosine in an eight-year-old girl. *The Pediatric Infectious Disease Journal*. 37, 275-277. <https://doi.org/10.1097/INF.0000000000001741>
- Karmeling J.P., Verluis, C., Schauer, R., 1975. *Carbohydrate R*. 41, 7-17.
- Kotawong, K.; Chaijaorenkul, W.; Muhamad, P.; Na-Bangchang, K., 2018. Cytotoxic activities and effects of atracylodin and β -eudesmol on the cell cycle arrest and apoptosis on cholangiocarcinoma cell line. *Journal of Pharmacological Sciences*. 136, 51-56. <https://doi.org/10.1016/j.jphs.2017.09.033>
- Lawal, O. A., Ogunwande, I. A., Owolabi, M. S., Opoku, A. R., Oyedeleji, A. O., 2016. Chemical composition, antibacterial activity, and brine shrimp lethality test of essential oil

from the leaves of *Eugenia natalitia*. Chemistry of Natural Compounds. 52, 731-733.
<http://dx.doi.org/10.1590/S0102-695X2010005000010>

Le Senne, A., Muelas-Serrano, S., Fernández-Portillo, C., Escario, J. A., Gómez-Barrio, A., 2002. Biological characterization of a beta-galactosidase expressing clone of Trypanosoma cruzi CL strain. Memórias do Instituto Oswaldo Cruz. 97, 1101–1105.
<http://dx.doi.org/10.1590/S0074-02762002000800006>

Linhares, K.V., Silva, W.A.G., 2015. Soldadinho-do-Araripe, símbolo da conservação das águas e florestas úmidas do Cariri Cearense. Caderno de Cultura e Ciência. 13, 37-50.
<http://dx.doi.org/10.14295/cad.cult.cienc.v13i2.849>

Loiola, M.I.B., Araújo, F.S., Lima-Verde, L.W., 2015. Flora da Chapada do Araripe. In: Albuquerque UP, Meiado MV. (eds.) Sociobiodiversidade na Chapada do Araripe. 6, 103-148.

Marin-Neto, J.A., Rassi Júnior, A., Avezum Júnior, A., Mattos, A.C., Rassi, A., Morillo C.A., Sosa-Estani, S., Yusuf, S., 2009. The benefit trial: testing the hypothesis that trypanocidal therapy is beneficial for patients with chronic Chagas heart disease. Memórias do Instituto Oswaldo Cruz, Rio de Janeiro. 104, 319–324.
<http://dx.doi.org/10.1590/S0074-02762009000900042>

Medeiros, J.R., Medeiros, N., Medeiros, H., Davin, L.B., Lewis, N.G., 2003. Composition of the bioactive essential oils from the leaves of Eugenia stipitata McVaugh ssp. sororia from de Azores. Journal of Essential Oil Research. 15, 293-295.
<http://dx.doi.org/10.1080/10412905.2003.9712145>

Meira, C.S., Menezes, L.R.A., Santos, T.B., Macedo, T.S., Fontes, J.E.N., Costa, E.V., Pinheiro, M.L.B., Da Silva, T.B., Teixeira, G.E, Soares, M.B.P., 2016. Chemical composition and antiparasitic activity of essential oils from leaves of Guatteria friesiana and Guatteria pogonopus (Annonaceae). Journal of essential oil research. 29, 156-162.
<https://doi.org/10.1080/10412905.2016.1210041>

Melo, N.I., Magalhaes, L.G., Carvalho, C.E., Wakabayashi, K.A., Aguiar, P.G., Ramos, R.C., Mantovani, A.L., Turatti, I.C., Rodrigues, V., Groppo, M., Cunha, W.R., Veneziani, R.C., Crotti, A.E., 2011. Schistosomicidal activity of the essential oil of Ageratum conyzoides L. (Asteraceae) against adult Schistosoma mansoni worms. Molecules. 16, 762-773.
<http://doi.org/10.3390/molecules16010762>

Mendiola, J., Pino, J.A., Fernández-Calienes, A.; Mendoza, D.; Herrera, P., 2015. Chemical composition and in vitro antiplasmodial activity of essential oils of leaves and flowers of alpinia zerumbet grown in cuba. Pharmacology Online. 2, 1-5.

Mevy, J.P., Bessiere, J.M., Dherbomez, M., Millogo, J., Viano, J., 2007. Chemical composition and some biological activities of the volatile oils of a chemotype of *Lippia chevalieri* Moldenke. Food Chemistry. 101, 682-685.
<https://doi.org/10.1016/j.foodchem.2006.01.052>

Mikus, J., Steverding, D., 2000. A simple colorimetric method to screen drug cytotoxicity against Leishmania using the dye Alamar Blue. Parasitology International. 48, 265–269.

[https://doi.org/10.1016/S1383-5769\(99\)00020-3](https://doi.org/10.1016/S1383-5769(99)00020-3)

MMA - Ministério do Meio Ambiente. 2000. Avaliação e ações prioritárias para a conservação da Biodiversidade da Floresta Atlântica e Campos Sulinos. Brasília, MMA/SBF.

Moncayo, A., Silveira, A.C., 2017. Current epidemiological trends for Chagas' disease in Latin America and future challenges in epidemiology, surveillance and health policy.

Memorial do Instituto Oswaldo Cruz, Rio de Janeiro. 104, 59-88.

<http://dx.doi.org/10.1590/S0074-02762009000900005>

Morrone, J.J., 2014. Biogeographical regionalisation of the Neotropical region. Zootaxa. 3782, 1-110. <http://dx.doi.org/10.11646/zootaxa.3782.1.1>

Pennington, R.T., Lewis, G.P., Ratter, J.A., 2006. Neotropical savannas and dry forests: plant diversity, biogeography, and conservation. Oxford, Taylor & Francis. 1, 54..

<http://dx.doi.org/10.1201/9781420004496.ch1>

Pereira, N.L.F., Aquino, P.E.A, Júnior, G.J.A.S., Cristo, J.S., Vieira Filho, M.A., Moura, F.F., Ferreira, N.M.N., Silva, M.K.N., Nascimento, E.M., Correira, F.M.A., Cunha, F.A.B., Boligon, A.A., Coutinho, H.D.M., Matias, E.F.F., Guedes, M.I.F., 2017. In vitro evaluation of the antibacterial potential and modification of antibiotic activity of the *Eugenia uniflora* L. essential oil in association with led lights. Microbial Pathogenesis. 2017, 512-518.

<http://doi.org/10.1016/j.micpath.2017.07.048>

Prado, D.E., Gibbs, P.E., 1993. Patterns of species distributions in the dry seasonal forest of South America. Annals of the Missouri Botanical Garden. 80, 902-927. <http://doi.org/10.2307/2399937>

Pinheiro, E., Brum-Soares, L., Reis, R., Cubides, J. C., 2017. Chagas disease: review of needs, neglect, and obstacles to treatment access in Latin America. Revista da Sociedade Brasileira em Medicina Tropical. 50, 296–300. <http://dx.doi.org/10.1590/0037-8682-0433-2016>

Reinaldo R.C.P.S., Saraiva, A.A.F., Santiago, A.C.P., 2015. Samambaias e licófitas da Chapada do Araripe. In: Albuquerque UP, Meiado MV. (eds.) Sociobiodiversidade na Chapada do Araripe. 1, 85-102.

Ribeiro, T.G., Chávez-Fumagalli, M.A., Valadares, D.G., Franca, J.R., Lage, P.S., Duarte, M.C., Andrade, P.H.R., Martins, V.T., Costa, L.E., Arruda, A.L.A., Faraco, A. A.G., Coelho, E.A.F., Castilho, R.O., 2014. Antileishmanial activity and cytotoxicity of Brazilian plants. Experimental Parasitology. 143, 60-68. <https://doi.org/10.1016/j.exppara.2014.05.004>

Rodrigues, K.A. F., Amorim, L.V., Oliveira, J.M.G.D., Dias, C.N., Moraes, D.F.C., Andrade, E.H.D.A., Maia J.G.S., Carneiro, S.M.P., Carvalho, F.A.D.A., 2013. *Eugenia uniflora* L. essential oil as a potential anti-*Leishmania* agent: effects on *Leishmania amazonensis* and possible mechanisms of action. Evidence-Based Complementary and Alternative Medicine. 2013, 1-10. <http://dx.doi.org/10.1155/2013/279726>

Roldos, V., Nakayama, H., Rolón, M., Montero-Torres, A., Trucco, F., Torres, S., Vega, C., Marrero-Ponce, Y., Heguaburu, V., Yaluff, G., Gómez-Barrio, A., Sanabria, L., Ferreira,

M.E., Rojas de Arias, A., Pandolfi, E., 2008. Activity of a hydroxybinenzyl bryophyte constituent against *Leishmania* spp and *Trypanosoma cruzi*: In-silico, in vitro and in-vivo activity studies. *European Journal of Medicinal Chemistry*. 43, 1797-1807.
<https://doi.org/10.1016/j.ejmech.2007.11.007>

Rolón, M., Seco, E.M., Vega, C., Nogal, J.J., Escario, J.A., Gómez-Barrio, A., Malpartida, F., 2006. Selective activity of polyene macrolides produced by genetically modified *Streptomyces* on *Trypanosoma cruzi*. *International Journal of Antimicrobial Agents*. 28, 104-109. <https://doi.org/10.1016/j.ijantimicag.2006.02.025>

Romagnolo, M.B., Souza, M.C., 2006. O gênero *Eugenia* L. (Myrtaceae) na planície de alagável do Alto Rio Paraná, Estados de Mato Grosso do Sul e Paraná, Brasil. *Acta Botânica Brasiliaca*. 20, 529-548. <http://dx.doi.org/10.1590/S0102-33062006000300004>.

Ronquillo, E.C.C., Galarza, T.S.C., 2016. Evaluación de la actividad antioxidante bioautográfica de 5 variedades de aceites esenciales amazónicos (*Ocotea quixos*, *Psidium guajava*, *Eugenia stipitata*, *Piper auritum*, *Piper imperiale*). Dissertação (Mestrado em Biotecnología de Recursos Naturais) – Sede Quito, Universidad Politecnica Salesiana, Quito.

Santoro, G.F.; Cardoso, M.G.; Guimarães, L.G.; Freire, J.M.; Soares, M.J., 2007. Anti-proliferative effect of the essential oil of *Cymbopogon citratus* (DC) Stapf (lemongrass) on intracellular amastigotes, bloodstream trypomastigotes and culture epimastigotes of *Trypanosoma cruzi* (Protozoa: Kinetoplastida). *Parasitology*. 134, 1649–1656.
<https://doi.org/10.1017/S0031182007002958>

Santos, A.O., Ueda-Nakamura, T., Dias Filho, B.P., Veiga Júnior, V.F., Pinto, A.C., Nakamura, C.V., 2008. Effect of Brazilian copaiba oils on *Leishmania amazonensis*. *Journal of Ethnopharmacology*. 120, 204–208. <https://doi.org/10.1016/j.jep.2008.08.007>

Santos, K.K.A.; Matias, E.F.F.; Tintino, S.R.; Souza, C.E.S.; Braga, M.F.B.M.; Guedes, G.M.M.; Rólón, M.; Vega, C.; Arias, A.R.; Costa, J.G.M.; Menezes, I.R.A.; Coutinho, H.D.M., 2012a. Anti-*Trypanosoma cruzi* and cytotoxic activities of *Eugenia uniflora* L. *Experimental Parasitology*. 131, 130-132. <https://doi.org/10.1016/j.exppara.2012.02.019>

Santos, K.K.A., Matias, E.F.F., Tintino, S.R., Souza, C.E.S., Braga, M.F.B.M., Guedes, G.M.M., Rolón, M., Vega, C., Arias, A.R., Costa, J.G.M., Menezes, I.R.A., Coutinho, H.D.M., 2012b. Cytotoxic, trypanocidal, and antifungal activities of *Eugenia jambolana* L. *Journal of Medicinal Food*. 15, 66-70. <https://doi.org/10.1089/jmf.2010.0298>

Sauter, I.P., Santos, J.C., Apel, M.A., Cibulski, S.P., Roehe, P.M., Von Poser, G.L., Rott, M.B., 2011. Amoebicidal activity and chemical composition of *Pterocaulon polystachyum* (Asteraceae) essential oil. *Parasitology Research*. 109, 575-580.
<http://dx.doi.org/10.1590/S1516-05722013000400016>

Sharifi-Rad, J., Sureda, A; Tenore, G.C., Daghia, M., Sharifi-Rad, M., Valussi, M., Tundis, R., Sharifi-Rad, M., Loizzo, M.R., Ademiluyi, A.O., Sharifi-Rad, R., Ayatollahi, S.A., Iriti, M., 2017. Biological activities of essential oils: from plant chemoecology to traditional healing systems. *Molecules*. 22, 70. <http://dx.doi.org/10.3390/molecules22010070>.

Silva, A.G., Alves, R.C.C., Bezerra Filho, C.M., Bezerra-Silva, P.C., Santos, L.M.M. D.,

- Foglio, M.A., Navarro, D.M.A.F., Da Silva, M.V., Correia, M.T.D.S., 2015. Chemical composition and larvicidal activity of the essential oil from leaves of *Eugenia brejoensis* Mazine (Myrtaceae). Journal of Essential Oil Bearing Plants. 18, 1441-1447. <https://doi.org/10.1080/0972060X.2014.1000390>
- Silva, W.A.G., Linhares, K.V., Campos, A.A., 2011. Plano de ação nacional para a conservação do Soldadinho-do-Araripe. Brasília, Instituto Chico Mendes de Conservação da Biodiversidade. 15, 14-19.
- Silva Júnior, E.N., Jardim, G.A.M., Menna-Barreto, R.F.S., Castro, S.L., 2014. Anti Trypanosoma cruzi compounds: our contribution for the evaluation and insights on the mode of action of naphthoquinones and derivatives. Journal of the Brazilian Chemical Society. 25, 1780–1798. <http://dx.doi.org/10.5935/0103-5053.20140180>
- Souza, L.I.O., Bezzera-Silva, P.C., Navarro, D.M.D.A.F., Da Silva, A.G., Correia, M. T.D.S., Da Silva, M.V., De Figueiredo, R.C.B.Q., 2017. The chemical composition and trypanocidal activity of volatile oils from Brazilian Caatinga plants. Biomedicine & Pharmacotherapy. 96, 1055-1064. <https://doi.org/10.1016/j.biopha.2017.11.121>
- Susen, V.P., Frank, F.M., Cazorla, S.I., Anesini, C.A., Malchiodi, E.L., Freixa, B., Vila, R., Muschietti, L.V., Martino, V.S., 2008. Trypanocidal and Leishmanicidal Activities of Sesquiterpene Lactones from Ambrosia tenuifolia Sprengel (Asteraceae). Antimicrob Agents Chemother. 52, 2415-2419. DOI: 10.1128/AAC.01630-07
- Tariku, Y., Hymete, A., Hailu, A., Rohloff, J., 2010. Essential-oil composition, antileishmanial, and toxicity study of Artemisia abyssinica and Satureja punctata ssp. punctata from Ethiopia. Chemistry and Biodiversity. 7, 1009–1018. <https://doi.org/10.1002/cbdv.200900375>
- Tasdemir, D., Kaiser, M., Brun, R., Yardley, V., Schmidt, T.J., Tosun, F., Rüedi, P., 2006. Antitrypanosomal and leishmanicidal activities of flavonoids and their analogues: in vitro, in vivo, structure-activity relationship, and quantitative structure-activity relationship studies. Antimicrobial Agents and Chemotherapy. 50, 1352–1364. DOI: 10.1128/AAC.50.4.1352-1364.2006
- Tenfen, A., Siebert, D.A., Yamanaka, C.N., Córdova, C.M.M., Scharf, D.R., Simionatto, E.L., Alberton, M.D., 2016. Chemical composition and evaluation of the antimicrobial activity of the essential oil from leaves of *Eugenia platysema*. Natural Product Research. 30, 2007-2011. <https://doi.org/10.1080/14786419.2015.1107056>
- Urbina, J. A., 2002. Chemotherapy of Chagas disease. Current Pharmaceutical Design, 8, 287–295.
- Vega, C., Rolón, M., Martínez-Fernández, A. R., Escario, J. Á., Gómez-Barrio, A., 2005. A new pharmacological screening assay with Trypanosoma cruzi epimastigotes expressing beta-galactosidase. Parasitology Research. 95, 296-298. DOI: 10.1007/s00436-005-1300-3
- Victoria, F.N., Lenardão, E.J., Savegnago, L., Perin, G., Jacob, R.G., Alves, D., Da Silva, W.P., Da Motta, A.S., Nascente, P.S., 2012. Essential oil of the leaves of Eugenia uniflora L.: antioxidant and antimicrobial properties. Food and Chemical Toxicology. 50, 2668-2674.

<https://doi.org/10.1016/j.fct.2012.05.002>

WHO, 2008. Les Chagas disease: control and elimination.

WHO, 2018a. Neglected Tropical Diseases. Disponível em:
http://www.who.int/neglected_diseases/diseases/en. Acesso em: 04 mar. 2018.

WHO, 2018b. Leishmaniasis. Disponível em:
<http://www.who.int/mediacentre/factsheets/fs375/en>. Acesso em: 04 mar. 2018.

Yap, P.S.X., Yiap, B.C., Ping, H.C., Lim, S.H.E., 2014. Essential oils, a new horizon in combating bacterial antibiotic resistance. *The Open Microbiology Journal*. 8, 6-14.
<https://doi.org/10.2174/1874285801408010006>

Zatelli, G. A., Zimath, P., Tenfen, A., Cordova, C. M. M., Scharf, D. R., Simionatto, E. L., Alberton, M. D., Falkenberg, M., 2016. Antimycoplasmic activity and seasonal variation of essential oil of *Eugenia hiemalis* Cambess (Myrtaceae). *Natural Product Research*. 30, 1961-1964. <https://doi.org/10.1080/14786419.2015.1091455>

5 CONCLUSÃO GERAL

Os resultados obtidos nesse estudo permitem concluir que:

- O OE de *Eugenia stipitata* tem atividade Leishmanicida;
- O OE não apresentou efeito antiparasitário relevante frente as formas epimastigotas de *T. cruzi*;
- O OE da pesquisa apresentou moderada citotoxicidade frente a fibroblastos;
- Foi o primeiro relato de atividade antiparasitária do OE extraído das folhas de *Eugenia stipitata* e a primeira caracterização química do óleo dessa espécie encontrada no Brasil;
- Os constituintes químicos mais abundantes do OE das folhas de *Eugenia stipitata* extraídas do bioma Caatinga foram: β -eudesmol (15.28%), γ -eudesmol (10.85%), Elemol (10.21%), caryophyllene oxide (6.5%), Clovane (6.18%) e Espanulenol (6.14%).

REFERÊNCIAS

- ABE, F.; NAGAFUJI, S.; YAMAUCHI, T.; OKABE, H.; MAKI, J.; HIGO, H.; AKAHANE, H.; AGUILAR, A.; JIMÉNEZ-ESTRADA, M.; REYES-CHILPA, R. Trypanocidal Constituents in Plants 1. Evaluation of Some Mexican Plants for Their Trypanocidal Activity and Active Constituents in Guaco, Roots of *Aristolochia taliscana*. **Biol. Pharm. Bull.**, v. 25, n. 9, p. 1188–1191, 2002.
- ALBERNAZ, L. C. Activités antiparasitaires et antifongiques des plantes du Cerrado: *Spiranthera odoratissima* et *Diospyros hispida*. [s.n.]. [S.l.], v. 1, p. 27-201, 2010.
- ALBUQUERQUE, U. P.; MONTEIRO, J. M.; RAMOS, M. A.; AMORIM, E. L. C. Medicinal and magic plants from a public market in northeastern Brazil. **Journal of Ethnopharmacology**, v. 110, n. 1, p. 76-91, 2007.
- ALIZADEH, B.H.; FOROUMADI, A.; ARDESTANI, S.K.; POORRAJAB, F.; SHAFIEE, A. Leishmanicidal evaluation of novel synthetic chromenes. **Arch Pharm**, v. 341, p. 787–793, 2008.
- ALMASSY JÚNIOR, A. A.; LOPES, R. C.; ARMOND, C.; SILVA, F.; CASALI, V. W. D. Folhas de chá: plantas medicinais na terapêutica humana. **Viçosa: UFV**, p. 233, 2005.
- ALMEIDA, C. F. C. B. R.; ALBUQUERQUE, U. P. Uso e conservação de plantas e animais medicinais no estado de Pernambuco (Nordeste do Brasil): um estudo de caso. **Interciencia**, v. 27, n. 6, 2002.
- ALVES, E. S.; TRESMONDI, F.; LONGUI, E. L. Análise estrutural de folhas de *Eugenia uniflora* L. (Myrtaceae) coletadas em ambientes rural e urbano, SP, Brasil. **Acta Botânica Brasil**, v.22, p.241-248, 2008.
- ALVES, W. F.; MOTA, A. S.; LIMA, R. A. A.; BELLEZONI, R.; VASCONCELLOS, A. Termites as Bioindicators of habitat Quality in the caatinga, Brazil: Is There agreement Between Structural habitat variables and the Sampled assemblages?. **Neotropical Entomology**, v. 40, n. 1, p. 39-46, 2011.
- AMORIM, A. C.; LIMA, C. K.; HOVELL, A. M.; MIRANDA, A. L.; REZENDE, C. M. Antinociceptive and hypothermic evaluation of the leaf essential oil and isolated terpenoids from *Eugenia uniflora* L. (Brazilian Pitanga). **Phytomedicine**, v. 16, n. 10, p. 923-928, 2009.
- ANDRADE, J. S.; RIBEIRO, F. C. F.; ARAGÃO, C. G.; FERREIRA, S. A. N. Adequação tecnológica de frutos da Amazônia: licor de araçá-boi (*Eugenia stipitata*) McVaugh. **Acta Amazonica**, v. 27, n. 4, p. 273-278, 1997.
- ANDRADE-LIMA, D. The caatinga dominium. **Revista Brasileira de Botânica**, v.4, p.149-153, 1981.
- ANDRADE, Z.A.; BARRAL-NETTO, M. *Trypanosoma cruzi e doença de Chagas*. 2.ed. Rio de Janeiro, **Guanabara Koogan**, p. 153-169, 2000.
- APEL, M. A.; SOBRAL, M.; ZUANAZZI, J. A.; HENRIQUES, A. T. Essential oil composition of four *Plinia* species (Myrtaceae). **Flavor and Fragrance Journal**, v. 21, p.

565-567, 2006.

AZEREDO, C. M. O.; SANTOS, T. G.; MAIA, B. H. L. N. S.; SOARES, M. J. In vitro biological evaluation of eight different essential oils against *Trypanosoma cruzi* with emphasis on *Cinnamomum verum* essential oil. **Complementary & Alternative Medicine**, 2014.

BADKE, M. R.; BUDÓ, M. D. L. D.; DA SILVA, F. M.; RESSEL, L. B. Plantas medicinais: o saber sustentado na prática do cotidiano popular. **Escola Anna Nery Revista de Enfermagem**, v. 15, n. 1, p. 132-139, 2011.

BAKKALI, F.; AVERBECK, S.; AVERBECK, D.; IDAOMAR, M. Biological effects of essential oils—a review. **Food and Chemical Toxicology**, v. 46, n. 2, p. 446-475, 2008.

BARROSO, G. M.; PEIXOTO, A. L.; COSTA, C. G.; ICHASO, C. L.; LIMA, H. C.; **Sistemática das Angiospermas do Brasil, Myrtaceae**. 2. ed. Viçosa: Universidade Federal de Viçosa, 377p. 1984.

BASANO, S. A; CAMARGO, L. M. A. Leishmaniose tegumentar americana: histórico, epidemiologia e perspectivas de controle. **Rev bras epidemiol**, v.7, n.3, p. 328-337, 2004.

BASSO, L. A.; DA SILVA, L. H.; FETT-NETO, A. G.; AZEVEDO-JUNIOR, W. F.; MOREIRA, I. S.; PALMA, M. S.; CALIXTO, J. B.; ASTOLFI-FILHO, S.; DOS SANTOS R. R.; SOARES, M. B.; SANTOS, D. S. The use of biodiversity as source of new chemical entities against defined molecular targets for treatment of malaria, tuberculosis, and T-cell mediated diseases: a review. **Memórias do Instituto Oswaldo Cruz**, v. 100, n. 6, p. 475-506, 2005.

BASTING, R. T.; NISHIJIMA, C.M.; LOPES, J.A.; SANTOS, R.C.; LUCENA, P. L.; LAUFER, S.; BAUER, S.; COSTA, M.F.; SANTOS, L.C.; ROCHA, L.R.; VILEGAS, W; SANTOS, A.R.; SANTOS, C; HIRUMA-LIMA, C.A. Antinociceptive, anti-inflammatory and gastroprotective effects of a hydroalcoholic extract from the leaves of *Eugenia punicifolia* (Kunth) DC. in rodents. **Journal of ethnopharmacology**, v. 157, p. 257-267, 2014.

BEZERRA, I. C. F.; RAMON, R. T. M.; FERREIRA, M. R. A.; SOARES, L. A. L. Chromatographic profiles of extractives from leaves of *Eugenia uniflora*. **Revista Brasileira de Farmacognosia**, v. 28, n. 1, 2017.

BITTENCOURT, S. C.; CAPONI, S.; FALKENBERG, M. B. O uso das plantas medicinais sob prescrição médica: pontos de diálogo e controvérsias com o uso popular. **Revista Brasileira de farmacognosia**, v. 12, p. 89-91, 2002.

BONNEY, K. M.; ENGMAN, D. M. Chagas heart disease pathogenesis: One mechanism or many. **NIH Public Access.**, v. 8, n. 6, p. 510–518, 2010.

BRASIL. Ministério da Saúde. Secretaria de Vigilância em Saúde. **Manual de vigilância da leishmaniose tegumentar americana**, 2 ed., p.184, 2007.

CARNEIRO, N. S.; ALVES, J. M.; ALVES, C. C. F.; ESPERANDIM, V. R.; MIRANDA, M. L. D. ÓLEO ESSENCIAL DAS FLORES DE *Eugenia klotzschiana* (MYRTACEAE):

SUA COMPOSIÇÃO QUÍMICA E ATIVIDADES TRIPANOCIDA E CITOTÓXICA IN VITRO. **Revista Virtual de Química**, v. 9, n. 3, 2017.

CHÁVES FLORES, W. B.; CLEMENTE, C. R. Considerações sobre o araçá-boi (*Eugenia stipitata* McVaugh, Myrtaceae) na Amazônia Brasileira. In: **CONGRESSO BRASILEIRO DE FRUTICULTURA**. Florianópolis, 1984. *Anais...* Florianópolis: SBF, 1984. p. 167-177.

CORREIA, V. C. D. S.; LIMA, N. O.; OLIVEIRA, F. A. D. S.; SANTOS, A. P. D. A. D.; TELES, C. B. G.; OLIVEIRA JÚNIOR, W. P. D. Evaluation of the antiplasmodial and leishmanicidal potential of *Myrciaria dubia* (Myrtaceae) extract. **Revista da Sociedade Brasileira de Medicina Tropical**, v. 49, n. 5, p. 586-592, 2016.

COSTA, T. R.; FERNANDES, O. F.; SANTOS, S. C.; OLIVEIRA, C. M.; LIÃO, L. M.; FERRI, P. H.; PAULA, J. R.; FERREIRA, H. D.; SALES, B. H. N.; MARIA DO ROSÁRIO, R. S. Antifungal activity of volatile constituents of *Eugenia dysenterica* leaf oil. **Journal of ethnopharmacology**, v. 72, n. 1-2, p. 111-117, 2000.

DAMETTO, A. C.; AGUSTONI, D.; MOREIRA, T. F.; PLAZA, C. V.; PRIETO, A. M.; SILVA, T. G. A.; SOUZA, F. O.; BORALLE, N.; SORBO, J. M.; SILVA, D. H. S.; SOARES, C. P. Chemical composition and in vitro chemoprevention assessment of *Eugenia jambolana* Lam. (Myrtaceae) fruits and leaves. **Journal of Functional Foods**, v. 36, p. 490-502, 2017.

D'ANGELIS, A. S. R.; NEGRELLE, R. R. B. Pimenta pseudocaryophyllus (Gomes) Landrum: aspectos botânicos, ecológicos, etnobotânicos e farmacológicos. **Rev. bras. plantas med**, v. 16, n. 3, p. 607-617, 2014.

DE ARAÚJO-JORGE, T. C.; DE CASTRO, S. L. **Doença de Chagas: manual para experimentação animal**. SciELO-Editora FIOCRUZ, p. 368, 2000.

DHIFI, W.; BELLILI, S.; JAIZI, S.; BAHLOUL, N.; MNIF, W. Essential oils' chemical characterization and investigation of some biological activities: a critical review. **Medicines**, v. 3, n. 4, p. 25, 2016.

DI STASI, L. C.; HIRUMA-LIMA, C. A. **Plantas medicinais na Amazônia e Mata Atlântica**. 2. ed. São Paulo: UNESP, p. 323-331. 2002.

DONATO, A. M.; MORRETES, B. L. Anatomia foliar de *Eugenia brasiliensis* Lam. (Myrtaceae) proveniente de áreas de restinga e de floresta. **Revista Brasileira de Farmacognosia**, v. 17, p. 426, 2007.

DUKE, J. **Dr. Duke's phytochemical and ethnobotanical databases**. Disponível em: <<https://phytochem.nal.usda.gov/phytochem/search>> Acesso em: 27 de dezembro de 2017 .

DUSCHAK, V. G.; COUTO, A. S. An insight on targets and patented drugs for chemotherapy of Chagas disease. **Recent Patents on Anti-Infective Drug Discovery**, v. 2, n. 1, p. 19-51, 2007.

DUTRA, R. C.; BRAGA, F. G.; COIMBRA, E. S.; SILVA, A. D.; BARBOSA, N. R. Atividades antimicrobiana e leishmanicida das sementes de *Pterodon emarginatus* Vogel. **Rev. bras. farmacogn**, v. 19, n. 2a, p. 429-435, 2009.

DUTRA, P. M. L.; COUTO, L. C.; LOPES, A. H. C. S.; MEYER-FERNANDES, J. R.; Characterization of ecto-phosphatase activities of *Trypanosoma cruzi*: A comparative study between Colombiana and Y strains. **Acta Tropica**, v. 100, p. 88-95, 2006.

EMBRAPA. **Avaliação do desempenho do araçá-boi (*Eugenia stipitata* McVaugh) na região de Manaus, AM**, 1996. Disponível em: <<https://www.embrapa.br/busca-de-publicacoes/-/publicacao/665438/avaliacao-do-desempenho-do-araca-boi-eugenia-stipitata-mcvaugh-na-regiao-de-manaus-am>> Acesso em: 27 de dezembro de 2017.

ESTEVAM, E. B. B.; ALVES, C. C. F.; ESPERANDIM, V. R.; CAZAL, M. D. L.; SOUZA, A. F.; MIRANDA, M. L. D. Chemical composition, anti-*Trypanosoma cruzi* and cytotoxic activities of the essential oil from green fruits of *Protium ovatum* (BURSERACEAE). **Revista Brasileira de Fruticultura**, v. 40, n. 1, 2018.

FERNANDES, T. G. **Efeito sinérgico do extrato aquoso das folhas de *Psidium guineense* Swartz em associação com agentes antimicrobianos frente a cepas de *Staphylococcus aureus* multidrogas resistente**. 2011. Dissertação (Mestrado em Ciências Farmacêuticas) – Centro de Ciências da Saúde, Universidade Federal de Pernambuco, Recife.

FERNÁNDEZ-TRUJILLO, J. P.; HERNÁNDEZ, M. S.; CARRILLO, M.; BARRERA, J. Arazá (*Eugenia stipitata* McVaugh). In: YAHIA, E. M. (Org.). **Postharvest Biology and Technology of Tropical and Subtropical Fruits**. England: Woodhead Publishing, 2011. p. 98-117.

FERRAZ, M. L.; GAZZINELLI, R. T.; ALVES, R. O.; URBINA, J. A.; ROMANHA, A. J. The anti-*Trypanosoma cruzi* activity of posaconazole in a murine model of acute Chagas' Disease is less dependent on gamma interferon than that of benznidazole. **Antimicrob. Agents Chemother.**, v. 51, p. 1359-1364, 2007.

FERREIRA, E. I. Planejamento de fármacos na área de doença de chagas: avanços e desafios. **Revista Virtual de Química**, v.4, p. 225, 2012.

FIGUEIREDO, A. C.; BARROSO, J. G.; PEDRO, L. G.; SCHEFFER, J. C. Factors affecting secondary metabolite production in plants: volatile components and essential oils. **Flavour and Fragrance Journal**, v. 23, n. 4, p. 213-226, 2008.

FORZZA, R. C.; BAUMGARTZ, J. F.; COSTA, A.; HOPKINS, M. J. G.; LEITMAN, P. M.; LOHMANN, L. G.; MARTINELLI, G.; MENEZES, M.; MORIM, M. P.; NADRUZ-COELHO, M.; PEIXOTO, A. L.; PIRANI, J. R.; QUEIROZ, L. P.; STEHMANN, J. R.; WALTER, B. M. T.; ZAPPI, D. C. As angiospermas do Brasil. In: FORZZA et al. (Org.). **Catalogo de Plantas e Fungos do Brasil**. Rio de Janeiro: Andrea Jakobsson Estudio/ Instituto de Pesquisas Jardim Botanico do Rio de Janeiro, 2010. p. 78-89.

GALENO, D. M. L.; CARVALHO, R. P.; BOLETI, A. P.; LIMA, A. S.; OLIVEIRA, P. D. A.; PACHECO, C. C.; PEREIRA, S. T.; LIMA, E. S. Extract from *Eugenia punicifolia* is an antioxidant and inhibits enzymes related to metabolic syndrome. **Applied biochemistry and biotechnology**, v. 172, n. 1, p. 311-324, 2014.

GALHEIGO, M. R. U.; PRADO, L. C. D. S.; MUNDIN, A. M. M.; GOMES, D. O.;

CHANG, R.; LIMA, A. M. C.; CANABRAVA, H. A. N.; BISPO-DA-SILVA, L. B. Antidiarrhoeic effect of *Eugenia dysenterica* DC (Myrtaceae) leaf essential oil. **Natural product research**, v. 30, n. 10, p. 1182-1185, 2016.

GIULIETTI, A. M.; HARLEY, R. M.; QUEIROZ, L. P.; BARBOSA, M. R. V.; BOCAGENETA, A. L.; FIGUEIREDO, M. A. Espécies endêmicas da caatinga. In: SAMPAIO, E. V. S. B.; GIULIETTI, A. M.; VIRGÍNIO, J.; GAMARRA-ROJAS, C. F. L. (Eds.). **Vegetação e flora da caatinga**. Recife: Associação Plantas do Nordeste e Centro Nordestino de Informação sobre Plantas, 2002. p. 103-105.

GOMES, E. C. S.; VILAR, F. C. R.; PEREZ, J. O.; BARBOSA, J.; VILAR, R. C.; FREIRE, J. L. O.; LIMA, A. N.; DIAS, T. J. Plantas da caatinga de uso terapêutico: levantamento etnobotânico. **Engenharia Ambiental**, v. 5, n. 2, p. 74- 85, 2008.

GONTIJO, B.; CARVALHO, M. L. R. Leishmaniose tegumentar americana. **Revista Sociedade Brasileira de Medicina Tropical**, v. 36, n. 1, p. 71-80, 2003.

HALBERSTEIN, R. A. Medicinal plants: historical and cross-cultural usage patterns. **Annals of epidemiology**, v. 15, n. 9, p. 686-699, 2005.

KONDRAVIN, A.V.; BARANOVA, A.M.; MOROZOVA, L.F.; STEPANOVA, E.V. Global trends in malaria control. **Progress and topical tasks in ma-laria control programs Med Parazitol**, v. 3, n. 8, 2011.

LAGO, J. H. G.; SOUZA, E. D.; MARIANE, B.; PASCON, R.; VALLIM, M. A.; MARTINS, R. C. C.; BAROLI, A. A.; CARVALHO, B. A.; SOARES, M. G.; SANTOS, R. T.; SARTORELLI, P. Chemical and biological evaluation of essential oils from two species of Myrtaceae - *Eugenia uniflora* L. and *Plinia trunciflora* (O. Berg) Kausel. **Molecules**, v.16, p. 9827-9837, 2011.

LEAL, I. R.; DA SILVA, J. M. C.; TABARELLI, M.; LACHER-JUNIOR, T. E. Changing the course of biodiversity conservation in the Caatinga of northeastern Brazil. **Conservation Biology**, v. 19, n. 3, p. 701-706, 2005.

LEITE, P. E. C.; LIMA, A. K. G.; FRANÇA, G. R.; LAGROTA, C. J.; SANTOS, W. C.; QUIRICO, S. T. Implant of polymer containing pentacyclic triterpenes from *Eugenia puniceifolia* inhibits inflammation and activates skeletal muscle remodeling. **Archivum immunologiae et therapiae experimentalis**, v. 62, n. 6, p. 483-491, 2014.

LI, Y.; XU, J; YUAN, C; MA, H.; LIU, T.; LIU, F.; SEERAM, N. P.; MU, Y.; HUANG, X.; LI, L. Chemical composition and anti-hyperglycaemic effects of triterpenoid enriched *Eugenia jambolana* Lam. berry extract. **Journal of Functional Foods**, v. 28, p. 1-10, 2017.

LIMA, F. M.; OLIVEIRA, P.; MORTARA, R. A.; SILVEIRA, J. F.; BAHIA, D. The challenge of Chagas' disease: Has the human pathogen, *Trypanosoma cruzi*, learned how to modulate signaling events to subvert host cells? **New biotechnology**, v. 27, n. 6, p. 837–843, 2010.

LÔBO, K. M. S.; ATHAYDE, A. C. R. ; SILVA, A. M. A. ; RODRIGUES, F. F. G. ; LÔBO, I. S.; BEZERRA, D. A. C.; COSTA, J. G. M. Avaliação da atividade antibacteriana e

prospecção fitoquímica de *Solanum paniculatum Lam.* e *Operculina hamiltonii* (G. Don) D. F. Austin & Staples, do semi-árido paraibano. **Rev. Bras. Pl. Med.**, Botucatu, v.12, n.2, p.227-233, 2010.

LUCHESSI, A. D.; MARÇAL, B. F.; ARAÚJO, G. F.; ULIANA, L. Z.; ROCHA, M. R. G.; PINTO, T. J. A. Monitoração de propaganda e publicidade de medicamentos: âmbito de São Paulo. **Revista Farmacêutica de Ciências Farmacêuticas**, v. 41, n. 3, 2005.

MARGARET, E.; SHAILAJA, A. M.; RAO, V. V. Evaluation of antioxidant activity in different parts of *Syzygium cumini* L. **International Journal of Current Microbiology and Applied Sciences**, v. 4, p. 372-379, 2015.

MATOS, F.J. A. **Farmácias vivas**. 4. ed. Fortaleza, UFC/SEBRAE. 2002.

MCKERROW, J. H.; DOYLE, P. S.; ENGEL, J. C.; PODUST, L. M.; ROBERTSON, S. A.; FERREIRA, R.; SAXTON, T.; ARKIN, M.; KERR, I.D.; BRINEN, L.S.; CRAIK, C.S. Two approaches to discovering and developing new drugs for Chagas disease. **Memórias do Instituto Oswaldo Cruz**, v. 104, p. 263-269, 2009.

MCVAUGH, R. Tropical American Myrtaceae. **Fieldiana Botany**, v. 29, n. 3, p. 145-228, 1956.

MEDEIROS, J. R.; MEDEIROS, N.; MEDEIROS, H.; DAVIN, L. B.; LEWIS, N. G. Composition of the bioactive oils from the leaves of *Eugenia stipitata* McVaugh ssp. *sororia* from the Azores. **Journal of Essential Oil Research**, v. 15, p. 293-295, 2003.

MELO, R. R. **Perfil fitoquímico, avaliação da atividade antimicrobiana e biocompatibilidade de *Syzygium malaccense* (L.) Merr. & L. M. Perry (Myrtaceae)**. 2009. Dissertação (Mestrado em Ciências Farmacêuticas) – Centro de Ciências da Saúde, Universidade Federal de Pernambuco, Recife.

MICHELIN, D. C. Estudo químico-farmacológico de *Operculina macrocarpa* (L.) urb. (Convolvulaceae). Tese (doutorado) - Universidade Estadual Paulista, Faculdade de Ciências Farmacêuticas, 2008.

MIRANDA, M. L. D.; ALVES, C.; ALVES, J.; ESPERANDIM, V.; CARNEIRO, N. Óleo essencial das flores de *Eugenia klotzschiana* (Myrtaceae): sua composição química e atividades tripanocida e citotóxica in vitro. **Revista Virtual de Química**, v. 9, n. 3, 2017.

MITROPOULOS, P.; KONIDAS, P.; DURKIN-KONIDAS, M. New World cuta-neous leishmaniasis: updated review of current and future diagnosisand treatment. **J am Acad Dermatol**, v. 63, p. 309–322, 2010.

MORENO, M. A.; ZAMPINI, I. C.; COSTAMAGNA, M. S.; SAYAGO, J. M.; ORDÓÑEZ, R. M.; ISLA, M. I. Phytochemical composition and antioxidant capacity of *Psidium guajava* fresh fruits and flour. **Food and Nutrition Science**, v. 5, 2014.

MUKHERJEE, P. K. et al. Screening of anti-diarrhoeal profile of some plant extracts of a specific region of West Bengal, India. **Journal of Ethnopharmacology**, v. 60, p. 85-89, 1998.

MUÑOZ-SARAIVA, S. G.; HABERLAND, A.; WALLUKAT, G.; SCHIMKE, I. Chronic Chagas' heart disease: a disease on its way to becoming a worldwide health problem: epidemiology, etiopathology, treatment, pathogenesis and laboratory medicine. **Heart Failure Review**, v. 17, n. 1, p. 45-64, 2010.

NERI-NUMA, I.A.; CARVALHO-SILVA, L. B.; MORALES, J. P.; MALTA, L. G.L.; MURAMOTO, M. T.; FERREIRA, J. E. M.; CARVALHO, J. E.; RUIZ, A. L. T. G.; MARÓSTICA JÚNIOR, M. R. M.; PASTORE, G. M. Evaluation of the antioxidant, antiproliferative and antimutagenic potential of araçá-boi fruit (*Eugenia stipitata* Mc Vaugh—Myrtaceae) of the Brazilian Amazon Forest. **Food Research International**, v. 50, n. 1, p. 70-76, 2013.

NIETO, G. Biological Activities of Three Essential Oils of the Lamiaceae Family. **Medicines**, v. 4, n. 3, p. 63, 2017.

OGUNWANDE, I. A.; OLAWORE, N. O.; EKUNDAYO, O.; WALHER, T. M.; SETZER, W. N. Studies on the essential oils composition, antibacterial and cytotoxicity of *Eugenia uniflora* L. **International Journal of Aromatherapy**, v. 15, n. 3, p. 147-152, 2005.

OLIVEIRA, M. F.; NAGAO-DIAS, A. T.; PONTES, V. M. O.; SOUSA JÚNIOR, A. S.; COELHO, H. L. L.; COELHO, I. C. B. Tratamento etiológico da Doença de Chagas no Brasil. **Revista de Patologia Tropical** 2008, 37, 209.

OLIVEIRA, P. S.; CHAVES, V. C.; BONA, N. P.; SOARES, M. S. P.; CARDOSO, J. S.; VASCONCELLOS, F. A.; TAVARES, R. G.; VIZZOTTO, M.; SILVA, L. M. C. D.; GRECCO, F. B.; GAMARO, G. D.; SPANEVELLO, R. M.; LENCINA, C. L.; REGINATTO, F. H.; STEFANELLO, F. M. *Eugenia uniflora* fruit (red type) standardized extract: a potential pharmacological tool to diet-induced metabolic syndrome damage management. **Biomedicine & Pharmacotherapy**, v. 92, p. 935-941, 2017.

PALMEIRO, M. R.; ROSALINO, C. M. V.; QUINTELLA, L. P.; MORGADO, F. N.; MARTINS, A. C. C.; MOREIRA, J.; CONCEIÇÃO-SILVA, F. Leishmaniose gengival em um paciente HIV-negativo. **Cirurgia Oral, Medicina Oral, Patologia Oral, Radiologia Oral e Endodontologia**, v. 104, n. 6, p. e12-e16, 2007.

PEIXOTO, L. F. **Avaliação do efeito protetor do extrato bruto hidroalcoólico das folhas de *Eugenia dysenterica* DC. sobre a neurotoxicidade induzida pelo alumínio**. 2015. Dissertação (Mestrado em Ciências Biológicas) – Instituto de Ciências Biológicas, Universidade Federal de Goiás, Goiânia.

PEREIRA, N. L. F.; AQUINO, P. E. A.; JÚNIOR, J. G. A. S.; CRISTO, J. S.; VIEIRA FILHO, M. A.; MOURA, F. F.; FERREIRA, N. M. N.; SILVA, M. K. N.; NASCIMENTO, E. M.; NASCIMENTO, E. M.; CORREIA, F. M. A.; CUNHA, F. A. B.; BOLIGON, A. A.; COUTINHO, H. D. M.; RIBEIRO-FILHO, J.; MATIAS, E. F. F.; GUEDES, M. I. F. Antibacterial activity and antibiotic modulating potential of the essential oil obtained from *Eugenia jambolana* in association with led lights. **Journal of Photochemistry and Photobiology B: Biology**, v. 174, p. 144-149, 2017.

PEREIRA, P.; CEBOLA, M. J.; OLIVEIRA, M. C.; GIL, M. G. B. Antioxidant capacity and

identification of bioactive compounds of *Myrtus communis* L. extract obtained by ultrasound-assisted extraction. **Journal of Food Science and Technology**, v. 54, n. 13, p. 4362-4369, 2017.

PRADO, L. C. D. S.; SILVA, D. B.; DE OLIVEIRA-SILVA, G. L.; HIRAKI, K. R. N.; CANABRAVA, H. A. N.; BISPO-DA-SILVA, L. B. The gastroprotective effects of *Eugenia dysenterica* (Myrtaceae) leaf extract: the possible role of condensed tannins. **Biological and Pharmaceutical Bulletin**, v. 37, n. 5, p. 722-730, 2014.

QUEIROZ, J. M. G.; SUZUKI, M. C. M.; MOTTA, A. P. R.; NOGUEIRA, J. M. R.; CARVALHO, E. M. Aspectos populares e científicos do uso de espécies de *Eugenia* como fitoterápico. **Revista Fitoterápicos Eletrônica**, v. 9, n. 2, p. 87-100, 2015.

RASSI JR, A.; RASSI, A.; LITTLE, W. C.; XAVIER, S. S.; RASSI, S. G.; RASSI, A. G.; RASSI, G. G.; HASSLOCHER-MORENO, A.; SOUSA, A. S.; SCANAVACCA, M. I. Development and validation of a risk score for predicting death in Chagas' heart disease. **New England Journal of Medicine**, v. 355, n. 8, p. 799-808, 2006.

REBOLLO, J.; OLIVERO-VERBEL, J.; REYES, N. New agents with potential leishmanicidal activity identified by virtual screening of chemical databases: New agents with potential leishmanicidal activity. **Revista de la Universidad Industrial de Santander. Salud**, v. 45, n. 1, p. 33-40, 2013.

RODRIGUES, K.A.; AMORIM, L.V.; DIAS, C.N.; MORAES, D.F.; CARNEIRO, S.M.; CARVALHO, F.A. Syzygium cumini (L.) Skeels essential oil and its major constituent alpha-pinene exhibit anti-Leishmania activity through immunomodulation in vitro. **J Ethnopharmacol**, v. 160, p. 32–40, 2015.

ROMAGNOLO, M. B.; SOUZA, M. C. O gênero *Eugenia* L. (Myrtaceae) na planície de alagável do Alto Rio Paraná, Estados de Mato Grosso do Sul e Paraná, Brasil. **Acta Botânica Brasiliaca**, v. 20, n. 3, p. 529-548, 2006.

ROMERO, E. L.; MORILLA, M. J. Nanotechnological approaches against Chagas disease. **Advanced drug delivery reviews**, v. 62, n. 4-5, p. 576–588, 2010.

ROSA, M. S. S.; MENDONÇA-FILHO, R. R.; BIZZO, H. R.; DE ALMEIDA RODRIGUES, I.; SOARES, R. M. A.; SOUTO-PADRÓN, T.; LOPES, A. H. C. Antileishmanial activity of a linalool-rich essential oil from Croton cajucara. **Antimicrob. Agents Chemother.**, v. 47, v.6, p.1895-1901, 2003.

SACRAMENTO, C. K.; BARRETTO, W. S.; FARIA, J. C. Araçá-boi: uma alternativa para agroindústria. **Bahia Agrícola**, v. 8, n. 2, p. 22-24, 2008.

SAHA, S.; SUBRAHMANYAM, E. V. S.; CHANDRASHEKAR, K.; SHUBHASH, C. M.; SHASHIDARA, C. S. Evaluation of antinociceptive and anti-inflammatory activities of extract and fractions of *Eugenia jambolana* root bark and isolation of phytoconstituents. **Revista Brasileira de Farmacognosia**, v. 23, n. 4, p. 651-661, 2013.

SALES, D. S.; CARMONA, F.; AZEVEDO, B. C.; TALEB-CONTINI, S. H.; BARTOLOMEU, A. C.; HONORATO, F. B.; MARTINEZ, E. Z.; PEREIRA, A. M. *Eugenia puncticifolia* (Kunth) DC. as an Adjuvant Treatment for Type-2 Diabetes Mellitus: A non-

Controlled, Pilot Study. **Phytotherapy research**, v. 28, n. 12, p. 1816-1821, 2014.

SAMPAIO, E. V. S. B. Overview of the brazilian caatinga. In: BULLOCK, S. H.; MOONEY, H. A.; MEDINA, E. (Eds.). **Seasonally Tropical Dry Forests**. Cambridge: Cambridge University Press, 1995.

SANTOS, K. K. A.; ROLÓN, M.; VEGA, C.; ARIAS, A. R.; COSTA, J. G. M. D.; COUTINHO, H. D. M. Atividade leishmanicida in vitro de Eugenia uniflora e Momordica charantia. **Revista de Ciências Farmacêuticas Básica e Aplicada**, v. 34, n. 1, p. 47-50, 2013.

SERAFIM, C. **Estudo da composição química e das propriedades biológicas das partes aéreas de *Plinia golmerata***. 2006. Dissertação (Mestrado em Ciências Farmacêuticas) – Universidade do Vale do Itajaí, Santa Catarina.

SILVA, A. C. O.; ALBUQUERQUE, U. P. Woody medicinal plants of the caatinga in the state of Pernambuco (Northeast Brazil). **Acta botânica brasílica**, v. 19, n. 1, p. 17-26, 2005.

SILVA, C. F.; MEUSER, M. B.; DE SOUZA, E. M.; MEIRELLES, M. N. L.; STEPHENS, C. E.; SOM, P.; BOYKIN, D. W.; SOEIRO, M. N. C. Cellular Effects of Reversed Amidines on Trypanosoma cruzi. **Antimicrobial agents and chemotherapy**, v. 51, n. 11, p. 3803–3809, 2007.

SILVA, F.; CASALI, V. W. D. Plantas medicinais e aromáticas: pós-colheita e óleos essenciais. Viçosa – MG, 2000, 135p.

SILVA, P. D. Estudo fitoquímico e avaliação das atividades antimicrobianas e antiparasitárias dos flavonóides isolados de Myrcia hiemalis (Myrtaceae). **Repositório Institucional-UFBA**, p.92, 2007.

SILVA, R. A. Caracterização da flora apícola e do mel produzido por Apis mellifera L., 1758 (Hymenoptera: Apidae) no estado da Paraíba. Tese (Doutorado em Zootecnia) – Centro de Ciências Agrárias. Universidade Federal da Paraíba, Areia, 99 f.: il. 2006.

SOARES, E. C. **Caracterização de aditivos para secagem de araçá-boi (*Eugenia stipitata McVaugh*) em leito de espuma**. 2009. 89 f. Dissertação (Mestrado em Engenharia de Alimentos) – Universidade Estadual do Sudoeste da Bahia, Bahia. 2009.

SOEIRO, M. N. C.; DANTAS, A. P.; DALIRY, A.; DA SILVA, C. F.; BATISTA, D. G.; SOUZA, E. M.; OLIVEIRA, G. M.; SALOMÃO, K.; BATISTA, M. M.; PACHECO, M.; SILVA, P. B.; SANTA-RITA, R. M.; MENNA-BARRETO, R. F. S.; BOYKIN, D. W.; DE CASTRO, S. L. Experimental chemotherapy for Chagas disease: 15 years of research contributions from in vivo and in vitro studies. **Memórias do Instituto Oswaldo Cruz**, v. 104, p. 301-310, 2009.

SOUZA, C. E. S. SILVA, A. R. P.; ROCHA, J. E.; GOMES, M. C. V.; RÓLOM, M.; CORONEL, C.; COSTA, J. G. M.; NETTO, M. L. C.; ROLIM, L. A.; COUTINHO, H. D. M. LC-MS characterization, anti-kinetoplastide and cytotoxic activities of natural products from *Eugenia jambolana* Lam. and *Eugenia uniflora*. **Asian Pacific Journal of Tropical Biomedicine**, v. 7, n. 9, p. 836-841, 2017.

TEIXEIRA, A. R.; NITZ, N.; GUIMARO, M. C.; GOMES, C.; SANTOS-BUCH, C. A. Chagas disease. **Postgrad. Med. J.**, v. 82, p. 788-798, 2006.

TOLEDO, A.C.O.; HIRATA, L.H.; BUFFON, M.C.M.; MIGUEL. M. D.; MIGUEL, O.G. Fitoterápicos: uma abordagem farmacotécnica. **Revista Lecta**, Bragança Paulista, v. 21, n. 1/2, p. 7-13, 2003.

URBINA, J. A. Ergosterol biosynthesis and drug development for Chagas disease. **Memórias do Instituto Oswaldo Cruz**, v. 104, p. 311-318, 2009.

VANDESMET, L. C. S. Etnobotânica de plantas medicinais no bioma da Caatinga. Dissertação (Mestrado em Bioprospecção Molecular) – Universidade Regional do Cariri, Ceará. 2015.

_____. Estudo do efeito hepático e das atividades citoprotetora, leishmanicida e tripanocida do extrato hidroalcóolico da casca de *Stryphnodendron rotundifolium* Mart. Dissertação (Mestrado em Bioprospecção Molecular) – Universidade Regional do Cariri, Ceará. 2014.

VEBER, J.; PETRINI, L. A.; ANDRADE, L. B.; SIVIERO, J. Determinação dos compostos fenólicos e da capacidade antioxidante de extratos aquosos e etanólicos de Jambolão (*Syzygium cumini* L.). **Revista Brasileira de Plantas Medicinais**, v. 17, p. 267-273, 2015.

VICTORIA, F. N.; LENARDÃO, E. J.; SAVEGNAGO, L.; PERIN, G.; JACOB, R. G.; ALVES, D.; SILVA, W. P.; MOTTA, A. S.; NASCENTE, P. S. Essential oil of the leaves of *Eugenia uniflora* L.: antioxidant and antimicrobial properties. **Food and Chemical Toxicology**, v. 50, n. 8, p. 2668-2674, 2012.

VITEK, R.; DE NOVAIS, L. M.; TORQUATO, H. F.; PAREDES-GAMERO, E. J.; DE CARVALHO, M. G.; SOUSA-JUNIOR, P. T.; JACINTO, M. J.; DA SILVA, V. C. Chemical constituents and antileukemic activity of *Eugenia dysenterica*. **Natural product research**, v. 31, n. 16, p. 1930-1934, 2017.

WALDRON, L.S.; FERRARI, B.C.; CHEUNG-KWOK-SANG, C.; BEGGS, P. J.; STEPHENS, N.; POWER, M.L. Molecular epidemiology and spatial distributionof a waterborne cryptosporidiosis outbreak in Australia. **ApplEnviron Microbiol**, v. 77, p. 7766–7771, 2011.

WERNECK, G. L. Geographic spread of visceral leishmaniasis in Brazil. **Cadernos de Saúde Pública**, v. 26, n. 4, p. 644-645, 2010.

WILSON, P. G.; O'BRIEN, M. M.; GADEK, P. A.; QUINN, C. J. Myrtaceae revisited: a reassessment of interfamilial groups. **American Journal of Botany**, v. 88, p. 2013-2025, 2001.

WILSON, P. G.; O'BRIEN, M. M.; HELESWOOD, M. M.; QUINN, C. J. Relationships within Myrtaceae sensu lato based on a matK phylogeny. **Plant Systematic and Evolution**, v. 251, p. 3-19, 2005. CAPÍTULO 1.

ANEXO A – Normas da Revista



FOOD AND CHEMICAL TOXICOLOGY

**AUTHOR
INFORMATION
PACK**



DESCRIPTION

Food and Chemical Toxicology (FCT), an internationally renowned journal, that publishes original research articles and reviews on **toxic effects**, in animals and humans, of natural or synthetic chemicals occurring in the human environment with particular emphasis on **food, drugs, and chemicals, including agricultural and industrial safety**, and **consumer product safety**. Areas such as safety evaluation of **novel foods and ingredients, biotechnologically-derived** products, and **nanomaterials** are included in the scope of the journal. FCT also encourages submission of papers on **inter-relationships between nutrition and toxicology** and on *in vitro* techniques, particularly those fostering the **3 Rs**.

The principal aim of the journal is to publish high impact, scholarly work and to serve as a multidisciplinary forum for research in toxicology. Papers submitted will be judged on the basis of scientific originality and contribution to the field, quality and subject matter. **Studies should address at least one of the following:** Adverse physiological/biochemical, or pathological changes induced by **specific defined** substances New techniques for assessing potential toxicity, including molecular biology Mechanisms underlying toxic phenomena Toxicological examinations of specific chemicals or consumer products, both those showing adverse effects and those demonstrating safety, that meet current standards of scientific acceptability

Authors must **clearly and briefly identify what novel toxic effect (s) or toxic mechanism (s)** of the chemical are being reported and what their **significance** is in the abstract. Furthermore, sufficient doses should be included in order to provide information on NOAEL/LOAEL values.

Manuscripts describing research involving the following areas will not be considered: materials/substances of only local interest materials/substances for which the chemical composition is not clearly defined only pharmacological properties, or potentially beneficial effects using *in vitro* or *in vivo* systems chemical analyses of toxins in foods without addressing the toxic implication to humans [risk assessment should be included] unrealistic human doses, inappropriate route of exposure, or *in vitro* experiments that do not reflect serum levels in humans

FCT is committed to the highest standards. Only papers that have not been previously published, that fit in the above mentioned scope, and that have been reviewed by experts in the field prior to publication will be accepted. Cover letters must state that the manuscript is new and original and not under consideration for publication elsewhere. Co-authors should be individuals who have contributed substantially to the content of the papers. All authors must declare any potential conflict of interest and all financial support.

Benefits to authors

We provide many author benefits, such as free PDFs, a liberal copyright policy, special discounts on Elsevier publications and much more. Please click [here](#) for more information on our [author services](#).

Please see the [Guide for Authors](#) for information on article submission. If you require further information or help, please visit our [Support Center](#)

AUDIENCE

Food scientists, toxicologists, chemists and researchers working in the pharmaceutical industry.

IMPACT FACTOR

2016: 3.778 © Clarivate Analytics Journal Citation Reports 2017

GUIDE FOR AUTHORS

Your Paper Your Way

We now differentiate between the requirements for new and revised submissions. You may choose to submit your manuscript as a single Word or PDF file to be used in the refereeing process. Only when your paper is at the revision stage, will you be requested to put your paper in to a 'correct format' for acceptance and provide the items required for the publication of your article.

To find out more, please visit the Preparation section below.

INTRODUCTION

Food and Chemical Toxicology (FCT), an internationally renowned journal, aspires to publish original research articles and reviews on **toxic effects**, in animals or humans, of natural or synthetic chemicals occurring in the human environment with particular emphasis on **food, drugs, and chemicals, including agricultural and industrial safety**, and **consumer product safety**. Areas such as safety evaluation of **novel foods and ingredients, biotechnologically-derived products**, and **nanomaterials** are included in the scope of the journal. FCT also encourages submission of papers on **inter-relationships between nutrition and toxicology** and on *in vitro* techniques, particularly those fostering the **3 Rs**.

The principal aim of the journal is to publish high impact, scholarly work and to serve as a multidisciplinary forum for research in toxicology. Papers submitted will be judged on the basis of scientific originality and contribution to the field, quality and subject matter. Studies should address at least one of the following: Physiological, biochemical, or pathological changes induced by specific substances Techniques for assessing potential toxicity, including molecular biology Mechanisms underlying toxic phenomena Toxicological examinations of specific chemicals or consumer products, both those showing adverse effects and those demonstrating safety, that meet current standards of scientific acceptability

Manuscripts concerning materials/substances of only local interest for which the chemical composition of the material/substance is **not clearly defined** will **not** be considered. Manuscripts addressing only pharmacological properties, or only potentially beneficial effects using *in vitro* or *in vivo* systems, are not within the scope of the journal.

FCT is committed to the highest standards. Only papers that have not been previously published, that fit in the above mentioned scope, and that have been reviewed by experts in the field prior to publication will be accepted. Cover letters must state that the paper is new and original and not under consideration for publication elsewhere. Papers pending in other journals will not be considered. Co- authors should be individuals who have contributed substantially to the content of the papers.

Types of paper

The Journal's main purpose is the publication of papers reporting and interpreting original unpublished toxicological research, particularly studies promoting an understanding of the mechanisms underlying toxic effects or improvements in methods for predicting adverse effects. Papers reporting the toxicological examination of specific foods, chemicals or consumer products will be published, irrespective of the positive or negative nature of the

results, provided the tests and reporting meet current standards of acceptability. In addition, Short Communications will also be considered, as will concise interpretative Reviews of toxicological topics of contemporary significance. Letters to the Editor will be limited to comments on contributions already published in the journal; if a letter is accepted, a response (for simultaneous publication) will be invited from the authors of the original contribution. All Letters to the Editor should be submitted to the Editor in Chief, Jose L. Domingo through the online submission system of the Journal.

Submission checklist

You can use this list to carry out a final check of your submission before you send it to the journal for review. Please check the relevant section in this Guide for Authors for more details.

Ensure that the following items are present:

One author has been designated as the corresponding author with contact details:

- E-mail address
- Full postal address

All necessary files have been uploaded:

Manuscript:

- Include keywords
 - All figures (include relevant captions)
 - All tables (including titles, description, footnotes)
 - Ensure all figure and table citations in the text match the files provided
 - Indicate clearly if color should be used for any figures
- inprint Graphical Abstracts / Highlights files (where applicable) Supplemental files (where applicable)*

Further considerations

- Manuscript has been 'spell checked' and 'grammar checked'
- All references mentioned in the Reference List are cited in the text, and viceversa
- Permission has been obtained for use of copyrighted material from other sources (including the Internet)
- A competing interests statement is provided, even if the authors have no competing interests to declare
- Journal policies detailed in this guide have been reviewed
- Referee suggestions and contact details provided, based on journal requirements

For further information, visit our [Support Center](#).

BEFORE YOU BEGIN

Ethics in publishing

Please see our information pages on [Ethics in publishing](#) and [Ethical guidelines for journal publication](#).

Human and animal rights

If the work involves the use of human subjects, the author should ensure that the work described has been carried out in accordance with [The Code of Ethics of the World Medical Association \(Declaration of Helsinki\)](#) for experiments involving humans; [Uniform Requirements for manuscripts submitted to Biomedical journals](#). Authors should include a statement in the manuscript that informed consent was obtained for experimentation with human subjects. The privacy rights of human subjects must always be observed.

All animal experiments should comply with the [ARRIVE guidelines](#) and should be carried out in accordance with the U.K. Animals (Scientific Procedures) Act, 1986 and associated guidelines, [EU Directive 2010/63/EU for animal experiments](#), or the National Institutes of

Health guide for the care and use of Laboratory animals (NIH Publications No. 8023, revised 1978) and the authors should clearly indicate in the manuscript that such guidelines have been followed.

Conflict of interest

Food and Chemical Toxicology follows the ICMJE recommendations regarding conflict of interest disclosures. All authors are required to report the following information with each submission: All third- party financial support for the work in the submitted manuscript. All financial relationships with any entities that could be viewed as relevant to the general area of the submitted manuscript. All sources of revenue with relevance to the submitted work who made payments to you, or to your institution on your behalf, in the 36 months prior to submission. Any other interactions with the sponsor of outside of the submitted work should also be reported. Any relevant patents or copyrights (planned, pending, or issued). Any other relationships or affiliations that may be perceived by readers to have influenced, or give the appearance of potentially influencing, what you wrote in the submitted work.

As a general guideline, it is usually better to disclose a relationship than not. This information will be acknowledged at publication in a Transparency Document. Additional information on the ICMJE recommendations can be found at: <http://www.icmje.org>. The form for conflict of interest disclosure can be downloaded [here](#), or at http://www.icmje.org/coi_disclosure.pdf (if this link does not display properly in your browser, please right-click the link and select "Save Target As..." or "Save Link as..." from the popup menu.).

Submission declaration

Submission of an article implies that the work described has not been published previously (except in the form of an abstract, a published lecture or academic thesis, see '[Multiple, redundant or concurrent publication](#)' for more information), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright- holder.

Submission declaration and verification

Submission of an article implies that the work described has not been published previously (except in the form of an abstract or as part of a published lecture or academic thesis or as an electronic preprint, see <http://www.elsevier.com/postingpolicy>), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically without the written consent of the copyright-holder. To verify originality, your article may be checked by the originality detection service CrossCheck <http://www.elsevier.com/editors/plagdetect>.

Each manuscript must also be accompanied by a cover letter outlining the basic findings of the paper and their significance. Furthermore, it is understood that with submission of this article the authors have complied with the institutional policies governing the humane and ethical treatment of the experimental subjects (i.e. animals and human subjects), and that they are willing to share the original data and materials if so requested.

Preprints

Please note that [preprints](#) can be shared anywhere at any time, in line with Elsevier's [sharing policy](#). Sharing your preprints e.g. on a preprint server will not count as prior publication (see '[Multiple, redundant or concurrent publication](#)' for more information).

Changes to authorship

Authors are expected to consider carefully the list and order of authors **before** submitting their manuscript and provide the definitive list of authors at the time of the original submission. Any addition, deletion or rearrangement of author names in the authorship list should be made only **before** the manuscript has been accepted and only if approved by the journal Editor. To request such a change, the Editor must receive the following from the **corresponding author**: (a) the reason for the change in author list and (b) written confirmation (e-mail, letter) from all authors that they agree with the addition, removal or rearrangement. In the case of addition or removal of authors, this includes confirmation from the author being added or removed.

Only in exceptional circumstances will the Editor consider the addition, deletion or rearrangement of authors **after** the manuscript has been accepted. While the Editor considers the request, publication of the manuscript will be suspended. If the manuscript has already been published in an online issue, any requests approved by the Editor will result in a corrigendum.

Article transfer service

This journal is part of our Article Transfer Service. This means that if the Editor feels your article is more suitable in one of our other participating journals, then you may be asked to consider transferring the article to one of those. If you agree, your article will be transferred automatically on your behalf with no need to reformat. Please note that your article will be reviewed again by the new journal. [More information](#).

Copyright

Upon acceptance of an article, authors will be asked to complete a 'Journal Publishing Agreement' (see [more information](#) on this). An e-mail will be sent to the corresponding author confirming receipt of the manuscript together with a 'Journal Publishing Agreement' form or a link to the online version of this agreement.

Subscribers may reproduce tables of contents or prepare lists of articles including abstracts for internal circulation within their institutions. [Permission](#) of the Publisher is required for resale or distribution outside the institution and for all other derivative works, including compilations and translations. If excerpts from other copyrighted works are included, the author(s) must obtain written permission from the copyright owners and credit the source(s) in the article. Elsevier has [preprinted forms](#) for use by authors in these cases.

For gold open access articles: Upon acceptance of an article, authors will be asked to complete an 'Exclusive License Agreement' ([more information](#)). Permitted third party reuse of gold open access articles is determined by the author's choice of [user license](#).

Author rights

As an author you (or your employer or institution) have certain rights to reuse your work. [More information](#).

Elsevier supports responsible sharing

Find out how you can [share your research](#) published in Elsevier journals.

Role of the funding source

You are required to identify who provided financial support for the conduct of the research and/or preparation of the article and to briefly describe the role of the sponsor(s), if any, in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the article for publication. If the funding source(s) had no such involvement then this should be stated. Please see <http://www.elsevier.com/funding>.

Funding body agreements and policies

Elsevier has established a number of agreements with funding bodies which allow authors to

comply with their funder's open access policies. Some funding bodies will reimburse the author for the gold open access publication fee. Details of [existing agreements](#) are available online.

Open access

This journal offers authors a choice in publishing their research:

Subscription

- Articles are made available to subscribers as well as developing countries and patient groups through our [universal access programs](#).
- No open access publication fee payable by authors.
- The Author is entitled to post the [accepted manuscript](#) in their institution's repository and make this public after an embargo period (known as green Open Access). The [published journal article](#) cannot be shared publicly, for example on ResearchGate or Academia.edu, to ensure the sustainability of peer-reviewed research in journal publications. The embargo period for this journal can be found below. **Gold open access**
- Articles are freely available to both subscribers and the wider public with permitted reuse.
- A gold open access publication fee is payable by authors or on their behalf, e.g. by their research funder or institution.

Regardless of how you choose to publish your article, the journal will apply the same peer review criteria and acceptance standards.

For gold open access articles, permitted third party (re)use is defined by the following [Creative Commons user licenses](#):

Creative Commons Attribution (CC BY)

Lets others distribute and copy the article, create extracts, abstracts, and other revised versions, adaptations or derivative works of or from an article (such as a translation), include in a collective work (such as an anthology), text or data mine the article, even for commercial purposes, as long as they credit the author(s), do not represent the author as endorsing their adaptation of the article, and do not modify the article in such a way as to damage the author's honor or reputation.

Creative Commons Attribution-NonCommercial-NoDerivs (CC BY-NC-ND)

For non-commercial purposes, lets others distribute and copy the article, and to include in a collective work (such as an anthology), as long as they credit the author(s) and provided they do not alter or modify the article.

The gold open access publication fee for this journal is **USD 2800**, excluding taxes. Learn more about Elsevier's pricing policy: <https://www.elsevier.com/openaccesspricing>.

Green open access

Authors can share their research in a variety of different ways and Elsevier has a number of green open access options available. We recommend authors see our [green open access page](#) for further information. Authors can also self-archive their manuscripts immediately and enable public access from their institution's repository after an embargo period. This is the version that has been accepted for publication and which typically includes author-incorporated changes suggested during submission, peer review and in editor-author communications. Embargo period: For subscription articles, an appropriate amount of time is needed for journals to deliver value to subscribing customers before an article becomes freely available to the public. This is the embargo period and it begins from the date the article is formally published online in its final and fully citable form. [Find out more](#).

This journal has an embargo period of 12 months.

Elsevier Researcher Academy

[Researcher Academy](#) is a free e-learning platform designed to support early and mid-career

researchers throughout their research journey. The "Learn" environment at Researcher Academy offers several interactive modules, webinars, downloadable guides and resources to guide you through the process of writing for research and going through peer review. Feel free to use these free resources to improve your submission and navigate the publication process with ease.

Language (usage and editing services)

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Authors who feel their English language manuscript may require editing to eliminate possible grammatical or spelling errors and to conform to correct scientific English may wish to use the [English Language Editing service](#) available from Elsevier's WebShop.

Submission

Our online submission system guides you stepwise through the process of entering your article details and uploading your files. The system converts your article files to a single PDF file used in the peer- review process. Editable files (e.g., Word, LaTeX) are required to typeset your article for final publication. All correspondence, including notification of the Editor's decision and requests for revision, is sent by e-mail.

Referees

The Editors require submissions by the authors of the names and addresses of 4 potential reviewers for this submission. The institutional address and e-mail address are required. At least 2 of the referees should be from a different country to the corresponding author's. The Editors reserve the right to use these or other reviewers.

PREPARATION

NEW SUBMISSIONS

Submission to this journal proceeds totally online and you will be guided stepwise through the creation and uploading of your files. The system automatically converts your files to a single PDF file, which is used in the peer-review process.

As part of the Your Paper Your Way service, you may choose to submit your manuscript as a single file to be used in the refereeing process. This can be a PDF file or a Word document, in any format or lay- out that can be used by referees to evaluate your manuscript. It should contain high enough quality figures for refereeing. If you prefer to do so, you may still provide all or some of the source files at the initial submission. Please note that individual figure files larger than 10 MB must be uploaded separately.

References

There are no strict requirements on reference formatting at submission. References can be in any style or format as long as the style is consistent. Where applicable, author(s) name(s), journal title/book title, chapter title/article title, year of publication, volume number/book chapter and the pagination must be present. Use of DOI is highly encouraged. The reference style used by the journal will be applied to the accepted article by Elsevier at the proof stage. Note that missing data will be highlighted at proof stage for the author to correct.

Formatting requirements

There are no strict formatting requirements but all manuscripts must contain the essential elements needed to convey your manuscript, for example Abstract, Keywords, Introduction, Materials and Methods, Results, Conclusions, Artwork and Tables with Captions. If your article includes any Videos and/or other Supplementary material, this should be included in your initial submission for peer review purposes.

Divide the article into clearly defined sections.

Please ensure the text of your paper is double-spaced— this is an essential peer review requirement.

Figures and tables embedded in text

Please ensure the figures and the tables included in the single file are placed next to the relevant text in the manuscript, rather than at the bottom or the top of the file. The corresponding caption should be placed directly below the figure or table.

Peer review

This journal operates a single blind review process. All contributions will be initially assessed by the editor for suitability for the journal. Papers deemed suitable are then typically sent to a minimum of two independent expert reviewers to assess the scientific quality of the paper. The Editor is responsible for the final decision regarding acceptance or rejection of articles. The Editor's decision is final. [More information on types of peer review](#).

REVISED SUBMISSIONS

Use of word processing software

Regardless of the file format of the original submission, at revision you must provide us with an editable file of the entire article. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing the article. The electronic text should be prepared in a way very similar to that of conventional manuscripts (see also the [Guide to Publishing with Elsevier](#)). See also the section on Electronic artwork.

To avoid unnecessary errors you are strongly advised to use the 'spell-check' and 'grammar-check' functions of your word processor.

Article structure

Subdivision - numbered sections

Divide your article into clearly defined and numbered sections. Subsections should be numbered

1.1 (then 1.1.1, 1.1.2, ...), 1.2, etc. (the abstract is not included in section numbering). Use this numbering also for internal cross-referencing: do not just refer to 'the text'. Any subsection may be given a brief heading. Each heading should appear on its own separate line.

Introduction

State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

Material and methods

Provide sufficient details to allow the work to be reproduced by an independent researcher. Methods that are already published should be summarized, and indicated by a reference. If quoting directly from a previously published method, use quotation marks and also cite the source. Any modifications to existing methods should also be described.

Results

Results should be clear and concise.

Discussion

This should explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is often appropriate. Avoid extensive citations and discussion of published literature.

Conclusions

The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion or Results and Discussion section.

Essential title page information

- **Title.** Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.
- **Author names and affiliations.** Please clearly indicate the given name(s) and family name(s) of each author and check that all names are accurately spelled. You can add your name between parentheses in your own script behind the English transliteration. Present the authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lower-

case superscript letter immediately after the author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name and, if available, the e-mail address of each author.

- **Corresponding author.** Clearly indicate who will handle correspondence at all stages of refereeing and publication, also post-publication. This responsibility includes answering any future queries about Methodology and Materials. **Ensure that the e-mail address is given and that contact details are kept up to date by the corresponding author.**
- **Present/permanent address.** If an author has moved since the work described in the article was done, or was visiting at the time, a 'Present address' (or 'Permanent address') may be indicated as a footnote to that author's name. The address at which the author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes.

Abstract

A concise and factual abstract is required. The abstract should state briefly the purpose of the research, the principal results and major conclusions. An abstract is often presented separately from the article, so it must be able to stand alone. For this reason, References should be avoided, but if essential, then cite the author(s) and year(s). Also, non-standard or uncommon abbreviations should be avoided, but if essential they must be defined at their first mention in the abstract itself.

Graphical abstract

Although a graphical abstract is optional, its use is encouraged as it draws more attention to the online article. The graphical abstract should summarize the contents of the article in a concise, pictorial form designed to capture the attention of a wide readership. Graphical abstracts should be submitted as a separate file in the online submission system. Image size: Please provide an image with a minimum of 531 × 1328 pixels (h × w) or proportionally more. The image should be readable at a size of 5 × 13 cm using a regular screen resolution of 96 dpi. Preferred file types: TIFF, EPS, PDF or MS Office files. You can view [Example Graphical Abstracts](#) on our information site.

Authors can make use of Elsevier's [Illustration Services](#) to ensure the best presentation of their images and in accordance with all technical requirements.

Highlights

Please amend your research highlights so that they consist of 3 to 5 brief bullet points which convey the core findings of your work. Please ensure EACH bullet point does NOT exceed 125 characters (including spaces). An example is given below:

RESEARCH HIGHLIGHTS EXAMPLE:

* Research highlights are a mandatory field of a submitted paper & therefore should not exceed 85 characters including spaces.

Keywords

Immediately after the abstract, provide a maximum of 6 keywords, using British spelling and avoiding general and plural terms and multiple concepts (avoid, for example, 'and', 'of'). Be sparing with abbreviations: only abbreviations firmly established in the field may be eligible. These keywords will be used for indexing purposes.

Abbreviations

Abbreviations should be used sparingly; they should be defined when first used in the paper but also listed in alphabetical order under *Abbreviations* as a footnote to the title page (see above).

Acknowledgements

Collate acknowledgements in a separate section at the end of the article before the references and do not, therefore, include them on the title page, as a footnote to the title or otherwise. List here those individuals who provided help during the research (e.g., providing language

help, writing assistance or proof reading the article, etc.).

Nomenclature and units

All measurements should be expressed in metric, preferably SI, units. Test chemicals and enzymes must be clearly identified, IUPAC and CAS names being used, wherever possible with the aid of CAS Registry and EC numbers. Pesticides should be referred to by their ISO names and human and veterinary drugs by their INNs.

Footnotes

Footnotes should be used sparingly. Number them consecutively throughout the article. Many word processors build footnotes into the text, and this feature may be used. Should this not be the case, indicate the position of footnotes in the text and present the footnotes themselves separately at the end of the article.

Artw

ork

Electr

onic

artwo

rk

Gene

ral

points

- Make sure you use uniform lettering and sizing of your original artwork.
- Preferred fonts: Arial (or Helvetica), Times New Roman (or Times), Symbol, Courier.
- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Indicate per figure if it is a single, 1.5 or 2-column fitting image.
- For Word submissions only, you may still provide figures and their captions, and tables within a single file at the revision stage.
- Please note that individual figure files larger than 10 MB must be provided in separate source files. A detailed [guide on electronic artwork](#) is available.

You are urged to visit this site; some excerpts from the detailed information are given here.

Formats

Regardless of the application used, when your electronic artwork is finalized, please 'save as' or convert the images to one of the following formats (note the resolution requirements for line drawings, halftones, and line/halftone combinations given below):

EPS (or PDF): Vector drawings. Embed the font or save the text as 'graphics'.

TIFF (or JPG): Color or grayscale photographs (halftones): always use a minimum of 300 dpi. TIFF (or JPG): Bitmapped line drawings: use a minimum of 1000 dpi.

TIFF (or JPG): Combinations bitmapped line/half-tone (color or grayscale): a minimum of 500 dpi is required.

Please do not:

- Supply files that are optimized for screen use (e.g., GIF, BMP, PICT, WPG); the resolution is too low.
- Supply files that are too low in resolution.
- Submit graphics that are disproportionately large for the content.

Color artwork

Please make sure that artwork files are in an acceptable format (TIFF (or JPEG), EPS (or PDF), or MS Office files) and with the correct resolution. If, together with your accepted article, you submit usable color figures then Elsevier will ensure, at no additional charge, that these figures will appear in color online (e.g., ScienceDirect and other sites) regardless of whether or not these illustrations are reproduced in color in the printed version. **For color reproduction in print, you will receive information regarding the costs from Elsevier after receipt of your accepted article.** Please indicate your preference for color: in print or online only. [Further information on the preparation of electronic artwork](#).

Figure captions

Ensure that each illustration has a caption. A caption should comprise a brief title (**not** on the figure itself) and a description of the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

Tables

Please submit tables as editable text and not as images. Tables can be placed either next to the relevant text in the article, or on separate page(s) at the end. Number tables consecutively in accordance with their appearance in the text and place any table notes below the table body. Be sparing in the use of tables and ensure that the data presented in them do not duplicate results described elsewhere in the article. Please avoid using vertical rules and shading in table cells.

References

Citation in text

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these references are included in the reference list they should follow the standard reference style of the journal and should include a substitution of the publication date with either 'Unpublished results' or 'Personal communication'. Citation of a reference as 'in press' implies that the item has been accepted for publication.

Reference links

Increased discoverability of research and high quality peer review are ensured by online links to the sources cited. In order to allow us to create links to abstracting and indexing services, such as Scopus, CrossRef and PubMed, please ensure that data provided in the references are correct. Please note that incorrect surnames, journal/book titles, publication year and pagination may prevent link creation. When copying references, please be careful as they may already contain errors. Use of the DOI is encouraged.

A DOI can be used to cite and link to electronic articles where an article is in-press and full citation details are not yet known, but the article is available online. A DOI is guaranteed never to change, so you can use it as a permanent link to any electronic article. An example of a citation using DOI for an article not yet in an issue is: VanDecar J.C., Russo R.M., James D.E., Ambeh W.B., Franke M. (2003). Aseismic continuation of the Lesser Antilles slab beneath northeastern Venezuela. *Journal of Geophysical Research*, <https://doi.org/10.1029/2001JB000884>. Please note the format of such citations should be in the same style as all other references in the paper.

Data references

This journal encourages you to cite underlying or relevant datasets in your manuscript by citing them in your text and including a data reference in your Reference List. Data references should include the following elements: author name(s), dataset title, data repository, version (where available), year, and global persistent identifier. Add [dataset] immediately before the reference so we can properly identify it as a data reference. The [dataset] identifier will not appear in your published article.

Reference management software

Most Elsevier journals have their reference template available in many of the most popular reference management software products. These include all products that support [Citation Style Language styles](#), such as [Mendeley](#) and Zotero, as well as EndNote. Using the word processor plug-ins from these products, authors only need to select the appropriate journal template when preparing their article, after which citations and bibliographies will be automatically formatted in the journal's style. If no template is yet available for this journal,

please follow the format of the sample references and citations as shown in this Guide.

Users of Mendeley Desktop can easily install the reference style for this journal by clicking the following link:

<http://open.mendeley.com/use-citation-style/food-and-chemical-toxicology>

When preparing your manuscript, you will then be able to select this style using the Mendeley plug- ins for Microsoft Word or LibreOffice.

Reference formatting

There are no strict requirements on reference formatting at submission. References can be in any style or format as long as the style is consistent. Where applicable, author(s) name(s), journal title/book title, chapter title/article title, year of publication, volume number/book chapter and the pagination must be present. Use of DOI is highly encouraged. The reference style used by the journal will be applied to the accepted article by Elsevier at the proof stage. Note that missing data will be highlighted at proof stage for the author to correct. If you do wish to format the references yourself they should be arranged according to the following examples:

Reference style

Text: All citations in the text should refer to:

1. *Single author:* the author's name (without initials, unless there is ambiguity) and the year of publication;
2. *Two authors:* both authors' names and the year of publication;
3. *Three or more authors:* first author's name followed by 'et al.' and the year of publication. Citations may be made directly (or parenthetically). Groups of references should be listed first alphabetically, then chronologically.

Examples: 'as demonstrated (Allan, 2000a, 2000b, 1999; Allan and Jones, 1999).

Kramer et al. (2010) have recently shown'

List: References should be arranged first alphabetically and then further sorted chronologically if necessary. More than one reference from the same author(s) in the same year must be identified by the letters 'a', 'b', 'c', etc., placed after the year of publication.

Examples:

Reference to a journal publication:

Van der Geer, J., Hanraads, J.A.J., Lupton, R.A., 2010. The art of writing a scientific article. *J. Sci. Commun.* 163, 51–59.

Reference to a book:

Strunk Jr., W., White, E.B., 2000. *The Elements of Style*, fourth ed. Longman, New York. Reference to a chapter in an edited book:

Mettam, G.R., Adams, L.B., 2009. How to prepare an electronic version of your article, in: Jones, B.S., Smith , R.Z. (Eds.), *Introduction to the Electronic Age*. E-Publishing Inc., New York, pp. 281–304.

Reference to a website:

Cancer Research UK, 1975. Cancer statistics reports for the UK.

<http://www.cancerresearchuk.org/> aboutcancer/statistics/cancerstatsreport/ (accessed 13 March 2003).

Reference to a dataset:

[dataset] Oguro, M., Imahiro, S., Saito, S., Nakashizuka, T., 2015. Mortality data for Japanese oak wilt disease and surrounding forest compositions. Mendeley Data, v1. <https://doi.org/10.17632/xwj98nb39r.1>.

Journal abbreviations source

Journal names should be abbreviated according to the [List of Title Word Abbreviations](#).

Video

Elsevier accepts video material and animation sequences to support and enhance your scientific research. Authors who have video or animation files that they wish to submit with their article are strongly encouraged to include links to these within the body of the article.

This can be done in the same way as a figure or table by referring to the video or animation content and noting in the body text where it should be placed. All submitted files should be properly labeled so that they directly relate to the video file's content. In order to ensure that your video or animation material is directly usable, please provide the file in one of our recommended file formats with a preferred maximum size of 150 MB per file, 1 GB in total. Video and animation files supplied will be published online in the electronic version of your article in Elsevier Web products, including [ScienceDirect](#). Please supply 'stills' with your files: you can choose any frame from the video or animation or make a separate image. These will be used instead of standard icons and will personalize the link to your video data. For more detailed instructions please visit our [video instruction pages](#). Note: since video and animation cannot be embedded in the print version of the journal, please provide text for both the electronic and the print version for the portions of the article that refer to this content.

AudioSlides

The journal encourages authors to create an AudioSlides presentation with their published article. AudioSlides are brief, webinar-style presentations that are shown next to the online article on ScienceDirect. This gives authors the opportunity to summarize their research in their own words and to help readers understand what the paper is about. [More information and examples are available](#). Authors of this journal will automatically receive an invitation e-mail to create an AudioSlides presentation after acceptance of their paper.

Data visualization

Include interactive data visualizations in your publication and let your readers interact and engage more closely with your research. Follow the instructions [here](#) to find out about available data visualization options and how to include them with your article.

Supplementary material

Supplementary material such as applications, images and sound clips, can be published with your article to enhance it. Submitted supplementary items are published exactly as they are received (Excel or PowerPoint files will appear as such online). Please submit your material together with the article and supply a concise, descriptive caption for each supplementary file. If you wish to make changes to supplementary material during any stage of the process, please make sure to provide an updated file. Do not annotate any corrections on a previous version. Please switch off the 'Track Changes' option in Microsoft Office files as these will appear in the published version.

Research data

This journal encourages and enables you to share data that supports your research publication where appropriate, and enables you to interlink the data with your published articles. Research data refers to the results of observations or experimentation that validate research findings. To facilitate reproducibility and data reuse, this journal also encourages you to share your software, code, models, algorithms, protocols, methods and other useful materials related to the project.

Below are a number of ways in which you can associate data with your article or make a statement about the availability of your data when submitting your manuscript. If you are sharing data in one of these ways, you are encouraged to cite the data in your manuscript and reference list. Please refer to the "References" section for more information about data citation. For more information on depositing, sharing and using research data and other relevant research materials, visit the [research data](#) page.

Data linking

If you have made your research data available in a data repository, you can link your article directly to the dataset. Elsevier collaborates with a number of repositories to link articles on ScienceDirect with relevant repositories, giving readers access to underlying data that gives

them a better understanding of the research described.

There are different ways to link your datasets to your article. When available, you can directly link your dataset to your article by providing the relevant information in the submission system. For more information, visit the [database linking page](#).

For [supported data repositories](#) a repository banner will automatically appear next to your published article on ScienceDirect.

In addition, you can link to relevant data or entities through identifiers within the text of your manuscript, using the following format: Database: xxxx (e.g., TAIR: AT1G01020; CCDC: 734053; PDB: 1XFN).

Mendeley Data

This journal supports Mendeley Data, enabling you to deposit any research data (including raw and processed data, video, code, software, algorithms, protocols, and methods) associated with your manuscript in a free-to-use, open access repository. Before submitting your article, you can deposit the relevant datasets to *Mendeley Data*. Please include the DOI of the deposited dataset(s) in your main manuscript file. The datasets will be listed and directly accessible to readers next to your published article online.

For more information, visit the [Mendeley Data for journals page](#).

Data in Brief

You have the option of converting any or all parts of your supplementary or additional raw data into one or multiple data articles, a new kind of article that houses and describes your data. Data articles ensure that your data is actively reviewed, curated, formatted, indexed, given a DOI and publicly available to all upon publication. You are encouraged to submit your article for *Data in Brief* as an additional item directly alongside the revised version of your manuscript. If your research article is accepted, your data article will automatically be transferred over to *Data in Brief* where it will be editorially reviewed and published in the open access data journal, *Data in Brief*. Please note an open access fee of 500 USD is payable for publication in *Data in Brief*. Full details can be found on the [Data in Brief website](#). Please use [this template](#) to write your Data in Brief.

MethodsX

You have the option of converting relevant protocols and methods into one or multiple MethodsX articles, a new kind of article that describes the details of customized research methods. Many researchers spend a significant amount of time on developing methods to fit their specific needs or setting, but often without getting credit for this part of their work. MethodsX, an open access journal, now publishes this information in order to make it searchable, peer reviewed, citable and reproducible. Authors are encouraged to submit their MethodsX article as an additional item directly alongside the revised version of their manuscript. If your research article is accepted, your methods article will automatically be transferred over to MethodsX where it will be editorially reviewed. Please note an open access fee is payable for publication in MethodsX. Full details can be found on the MethodsX website. Please use [this template](#) to prepare your MethodsX article.

Data statement

To foster transparency, we encourage you to state the availability of your data in your submission. This may be a requirement of your funding body or institution. If your data is unavailable to access or unsuitable to post, you will have the opportunity to indicate why during the submission process, for example by stating that the research data is confidential. The statement will appear with your published article on ScienceDirect. For more information, visit the [Data Statement page](#).

AFTER ACCEPTANCE

Online proof correction

Corresponding authors will receive an e-mail with a link to our online proofing system, allowing annotation and correction of proofs online. The environment is similar to MS Word: in addition to editing text, you can also comment on figures/tables and answer questions from the Copy Editor. Web-based proofing provides a faster and less error-prone process by allowing you to directly type your corrections, eliminating the potential introduction of errors. If preferred, you can still choose to annotate and upload your edits on the PDF version. All instructions for proofing will be given in the e-mail we send to authors, including alternative methods to the online version and PDF.

We will do everything possible to get your article published quickly and accurately. Please use this proof only for checking the typesetting, editing, completeness and correctness of the text, tables and figures. Significant changes to the article as accepted for publication will only be considered at this stage with permission from the Editor. It is important to ensure that all corrections are sent back to us in one communication. Please check carefully before replying, as inclusion of any subsequent corrections cannot be guaranteed. Proofreading is solely your responsibility.

Offprints

The corresponding author will, at no cost, receive a customized [Share Link](#) providing 50 days free access to the final published version of the article on [ScienceDirect](#). The Share Link can be used for sharing the article via any communication channel, including email and social media. For an extra charge, paper offprints can be ordered via the offprint order form which is sent once the article is accepted for publication. Both corresponding and co-authors may order offprints at any time via Elsevier's [Webshop](#). Corresponding authors who have published their article gold open access do not receive a Share Link as their final published version of the article is available open access on ScienceDirect and can be shared through the article DOI link.

AUTHOR INQUIRIES

Visit the [Elsevier Support Center](#) to find the answers you need. Here you will find everything from Frequently Asked Questions to ways to get in touch.

You can also [check the status of your submitted article](#) or find out [when your accepted article will be published](#).