

UNIVERSIDADE FEDERAL DE PERNAMBUCO
CENTRO DE CIÊNCIAS BIOLÓGICAS
PROGRAMA DE PÓS-GRADUAÇÃO EM GENÉTICA

STÊNIO ÍTALO ARAÚJO FOERSTER

DIVERSIDADE GENÉTICA E PADRÕES FILOGEOGRÁFICOS DO ESCORPIÃO
JAGUAJIR ROCHAE (BORELLI, 1910) EM UM GRADIENTE DE
ESTABILIDADE CLIMÁTICA DA CAATINGA

Recife

2020

STÊNIO ÍTALO ARAÚJO FOERSTER

**Diversidade genética e padrões filogeográficos do escorpião *Jaguajir
rochae* (Borelli, 1910) em um gradiente de estabilidade climática da Caatinga**

Dissertação apresentada ao Programa de
Pós-Graduação em Genética da
Universidade Federal de Pernambuco como
parte dos requisitos exigidos para obtenção
do título de Mestre em Genética.

Orientador: Dr. Valdir de Queiroz Balbino

Recife

2020

Catálogo na fonte
Elaine C Barroso
(CRB4 1728)

Foerster, Stênio Ítalo Araújo

Diversidade genética e padrões filogeográficos do escorpião *Jaguajir rochae* (Borelli, 1910) em um gradiente de estabilidade climática da Caatinga/ Stênio Ítalo Araújo Foerster– 2020.

121 f.: il., fig., tab.

Orientador: Valdir de Queiroz Balbino

Dissertação (mestrado) – Universidade Federal de Pernambuco. Centro de Biociências. Programa de Pós-Graduação em Genética, 2020.
Inclui referências e anexo.

1. Escorpião 2. Biogeografia 3. Invertebrados I. Balbino, Valdir de Queiroz (orient.) II. Título

595.46

CDD (22.ed.)

UFPE/CB – 2020- 132

STÊNIO ÍTALO ARAÚJO FOERSTER

**Diversidade genética e padrões filogeográficos do escorpião *Jaguajir
rochae* (Borelli, 1910) em um gradiente de estabilidade climática da Caatinga**

Aprovado em: 07/04/2020

Banca Examinadora

**Dr. Valdir de Queiroz Balbino
Universidade Federal de Pernambuco**

**Dr. Rodrigo Augusto Torres
Universidade Federal de Pernambuco**

**Dr. Ricardo Pinto-da-Rocha
Universidade de São Paulo**

AGRADECIMENTOS

Agradeço e dedico este trabalho aos meus queridos amigos, alguns de longa data, outros nem tanto, mas todos de fundamental importância para minha vida pessoal e profissional. Minha querida prima Dra. Theresa Liberal, obrigado por ter me trazido para o fascinante mundo da genética, o qual, eu adentrei graças ao meu prezado orientador e amigo, Dr. Valdir Balbino, a quem devo imensurável gratidão pela confiança e suporte durante a minha estadia no PPGG/UFPE. Bárbara Natieli, uma profissional excepcional e uma amiga incrível, obrigado por me ensinar absolutamente tudo que foi necessário na bancada. Ao Dr. André Lira, meu brother de longa data, muito obrigado pela ajuda em campo sempre regada a risadas e boas discussões. Sou igualmente grato aos meus colegas José Rivaldo, Aleson Silva, Hugo Rodrigo, Welton Dionísio, Hugo Leonardo e Thayna Rhayane (Hugo-Thayna s2) pela ajuda na obtenção de material biológico. Não posso deixar de agradecer ao Dr. Márcio Bernardino daSilva (UFPB) pela disponibilidade e boa vontade em me auxiliar durante algumas coletas. De fundamental importância foi o meu amigo Raul Azevedo, que me deu todo o suporte necessário para o trabalho de campo no Ceará, sempre com muita disposição e boa vontade. Também devo lembrar das importantes contribuições prestadas pela equipe do Instituto Agrônomo de Pernambuco (especialmente ao Sr. Félix) que se prestou a dar todo o suporte necessário para nossas coletas em Caruaru. Agradeço a Dra. Flávia Lanna (UFRN/Ohio State University) por se dispor a discutir os resultados do presente estudo, sempre com muita atenção e dedicação, e também a minha companheira, namorada e amiga de toda vida, Valdeane G. Silva, não apenas pela ajuda em campo, mas principalmente pela enorme paciência para lidar com todos os meus estresses. Finalmente, sou grato ao Programa de Pós-Graduação em

Genética da UFPE e a todos os seus docentes, bem como ao Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) pela oportunidade de me graduar em um excelente Programa de Pós-Graduação, e ao mesmo tempo, contribuir para o conhecimento da história biogeográfica da Caatinga.

RESUMO

O domínio das Caatingas representa o maior núcleo contínuo de florestas tropicais sazonalmente secas do novo mundo, entretanto, a história evolutiva da fauna destes ambientes tem sido pouco investigada. Assim, o presente estudo buscou avaliar a atuação de potenciais núcleos de estabilidade climática como refúgios e / ou centros de diversificação da Caatinga, utilizando como modelo, o escorpião *Jaguajir rochae*. Diferentes estratégias de agrupamento e datação molecular foram empregadas em busca de possíveis compatibilidades cronológicas entre os processos de diversificação de linhagens mitocondriais de *J. rochae*, e os principais eventos climáticos e geomorfológico que moldaram as paisagens da Caatinga. Em adição, nove modelos de migração foram aplicados para elucidar os padrões de fluxo gênico entre populações distribuídas ao redor do Planalto da Borborema, avaliando assim, o efeito desta formação geológica na diferenciação genética e isolamento geográfico de populações de *J. rochae*. Nossos resultados indicam altos níveis de diversidade genética em populações situadas em áreas de Caatinga historicamente estáveis, e que os processos de diversificação intrapopulacional ocorreram após o soerguimento do Planalto da Borborema. Ainda, foi demonstrado que o Planalto da Borborema constitui uma barreira semipermeável ao fluxo gênico de *J. rochae*, e um padrão dinâmico de migração é proposto para a referida espécie. Finalmente, os processos que levaram a atual distribuição espacial da diversidade genética em populações de *J. rochae* são discutidos sob a luz da Teoria dos Gradientes Ecológicos.

Palavras-chave: Filogeografia. Biogeografia. SDTF. Invertebrados. Modelagem ecológica.

ABSTRACT

The Caatinga domain represent the largest and continuous nucleus of Seasonally Dry Tropical Forest in the New World, however, the evolutive story of its fauna is poorly known. Thus, we use the scorpion *Jaguajir rochae* as a biological model to investigate the role of potential nuclei of climatic stability, as diversification hotspots within the Caatinga formations. Different clustering strategies and molecular dating were applied to infer possible chronological associations among the processes of mitochondrial lineage diversification in *J. rochae*, and the major geoclimatic events that shaped the Caatinga landscapes. In addition, nine migration models were tested to elucidate the patterns of gene flow among populations distributed around the Borborema Plateau, evaluating its effects on the genetic differentiation and geographic isolation among populations of *J. rochae*. Our results indicate high levels of genetic diversity in populations distributed in historically stable areas of Caatinga, with intraspecific diversification events occurring after the uplift of the Borborema Plateau. We also demonstrated that the Borborema Plateau is a semipermeable barrier to the gene flow of *J. rochae*, and a dynamic pattern of gene flow is proposed for this species. Finally, the processes behind the current distribution of the genetic diversity among populations of *J. rochae* are discussed under the light of the Ecological Gradient Theory.

Keywords: Phylogeography. Biogeography. Caatinga. SDTF. Invertebrates. Ecological modelling.

LISTA DE ILUSTRAÇÕES

Referencial Teórico

Figura 1 -	Ecorregiões propostas para a Caatinga	23
Figura 2 -	Representação das áreas de estabilidade e instabilidade climática da Caatinga gerada a partir de modelos de circulação atmosférica	24
Figura 3 -	Representação esquemática da estrutura do genoma mitocondrial do escorpião <i>Mesobuthus martensii</i> (Karsch, 1879)	31
Figura 4 -	Estrutura do DNA ribossômico (rDNA) comumente encontrado no genoma nuclear de eucariotos	33
Figura 5 -	Distribuição geográfica das espécies do gênero <i>Jaguajir</i> na América do Sul	38
Figura 6 -	Espécime adulto de <i>Jaguajir rochae</i>	39
Figure 1 -	Geographical distribution of sampling locations of <i>Jaguajir rochae</i> , and genetic structure patterns around the Borborema Plateau	82
Figure 2 -	Migration models implemented in Migrate-n to test for nine gene flow schemes among populations of <i>Jaguajir rochae</i> distributed around the Borborema Plateau	83
Figure 3 -	Bayesian dated phylogeny inferred for non-redundant haplotypes of <i>Jaguajir rochae</i>	84
Figure 4 -	Binary maps of the potential distribution, and spatial changes in niche suitability of <i>Jaguajir rochae</i> through late-Quaternary	85
Figure S1 -	Pair plot depicting patterns of autocorrelation among bioclimatic variables used to construct ecological niche models for <i>Jaguajir rochae</i>	87

LISTA DE TABELAS

Table 1 -	Sampling locations of <i>Jaguajir rochae</i> , depicting the output of genetic diversity estimations and neutrality tests	78
Table 2 -	Summary results of the Analysis of Molecular Variance (AMOVA) computed from the mitochondrial data set of <i>Jaguajir rochae</i>	79
Table 3 -	Migrate-n model comparisons for different migration schemes among populations of <i>Jaguajir rochae</i>	79
Table S1 -	List of zoological collections (institutions) accessed to retrieve occurrence points of <i>Jaguajir rochae</i> used to construct ecological niche models	86
Table S2 -	Numerical results of model (D) and (F) estimated in Migrate-n using the mitochondrial data set of <i>Jaguajir rochae</i>	86

LISTA DE ABREVIATURAS, SIGLAS E SÍMBOLOS

Referencial Teórico

Item	Definição
FTSSs	Florestas Tropicais Sazonalmente Secas
HRP	Hipótese dos Refúgios Pleistocênicos
NC	Núcleo Caatinga
HAP	Hipótese do Arco-Pleistocênico
MDEs	Modelos de Distribuição de Espécies
mtDNA	DNA mitocondrial
nDNA	DNA nuclear
rnDNA	DNA ribossômico nuclear
COI	Citocromo c oxidase I
SDTFs	Seasonally Dry Tropical Forests
CD	Caatinga Domain
BOP	Borborema Plateau
ICMBio	Instituto Chico Mendes de Preservação da Biodiversidade
COI	Cytochrome c oxidase I
SER	Serra Talhada
h	Number of haplotypes
Hd	Haplotype diversity
π	Nucleotide diversity
AMOVA	Analysis of Molecular Variance
BIC	Bayesian Information Criterion
LBF	Log Bayes Factor
MCMC	Markov Chain Monte Carlo
ESS	Effective Sample Size
M	Mutation-scaled migration rate
Θ	Mutation-scaled effective population size
ENMs	Ecological Niche Models
LIG	Last Interglacial
LGM	Last Glacial Maximum
VIF	Variance Inflation Factor
AUC_{TEST}	Area under the receiver operating characteristic curve for the testing data
AUC_{DIFF}	The difference between the area under the receiver operating characteristic curve calculated for testing, and training data
TSS	True Skill Statistics

OR0	0% Omission Rate
OR10	10% Omission Rate
CAR	Caruaru
CUM	Cumaru
AGU	Água Branca
AFO	Afogados da Ingazeira
CAE	Caetés
PAR	Parnamirim
LIM	Limoeiro
HPD	Highest Posterior Density

SUMÁRIO

1	INTRODUÇÃO	14
1.1	Objetivo geral	16
1.2	Objetivos específicos	16
2	REFERENCIAL TEÓRICO	17
2.1	Aspectos biogeográficos da Caatinga	17
2.2	Núcleos de estabilidade climática da Caatinga	22
2.3	A Filogeografia e os modelos de distribuição de espécies na identificação de centros de diversificação	27
2.4	Seleção do organismo modelo	34
3	Insights on the role of historical habitat stability and geological barriers on lineage diversification processes in Caatinga environments revealed by landscape genetics of the scorpion <i>Jaguajir rochae</i> (Scorpiones: Buthidae)	40
4	DISCUSSÃO GERAL	88
5	CONCLUSÕES	90
	REFERÊNCIAS BIBLIOGRÁFICAS	91
	ANEXO I. NORMAS DE FORMATAÇÃO EXIGIDAS PELO PERIÓDICO <i>JOURNAL OF BIOGEOGRAPHY</i>	110
	CURRICULUM VITAE (LATTES)	121

1 INTRODUÇÃO

A identificação de refúgios naturais é uma etapa fundamental para a preservação da biodiversidade. Esses refúgios são áreas que propiciam condições favoráveis de habitat, facilitando a permanência das espécies em escala de tempo evolutiva, ao mesmo tempo em que preservam a diversidade genética da biota, favorecendo os eventos de diversificação de espécies e/ou linhagens. Nesse contexto, análises filogeográficas e inferências de diversidade genética, podem auxiliar na identificação e delimitação de refúgios (áreas de estabilidade climática) e centros de diversificação de espécies, além de expor potenciais barreiras ao fluxo gênico de populações.

No presente estudo, nós buscamos compreender e utilizar os padrões filogeográficos do escorpião *Jaguaíra rochae* (Borelli, 1910), como ferramenta para testar a atuação de áreas de alta estabilidade climática como refúgios históricos, importantes para promover diversificação da biodiversidade em ambientes de Caatinga. Nós também avaliamos a atuação do Planalto da Borborema como o principal elemento orográfico da paisagem que limita o fluxo gênico entre populações de *J. rochae* em escala regional. Sob um ponto de vista histórico, nós apresentamos novas perspectivas sobre as inter-relações entre a história geoclimática da Caatinga e os processos de diversificação e estruturação de populações que ocorrem nestes ambientes. A notória escassez de informações sobre os processos evolutivos responsáveis pelos padrões atuais de distribuição espacial da diversidade genética em táxons de Caatinga foi uma das principais motivações para a condução deste estudo. Em adição, a assertiva geral de que a Caatinga não apresenta barreiras orográficas suficientes para impor resistência ao fluxo gênico entre populações, nos motivou a testar a hipótese de que o Planalto

da Borborema constitui um importante elemento na estruturação genética de espécies com baixa capacidade de dispersão.

Com isso, nós acreditamos que o presente estudo agrega novas informações que são relevantes para o progressivo desenvolvimento do conhecimento acerca da história biogeográfica da Caatinga, gerando um conteúdo que certamente fomentará discussões em âmbito acadêmico. Do ponto de vista prático, o nosso estudo ilustra a evolução da diversidade genética em linhagens mitocondriais de *J. rochae* ao longo do tempo, provendo detalhes sobre como a distribuição espacial desta diversidade respondeu aos ciclos climáticos do Pleistoceno em escala regional. Como os eventos climáticos mencionados possuem um comportamento cíclico, os nossos resultados podem fornecer subsídios para possíveis previsões a respeito dos efeitos das mudanças climáticas sobre a diversidade genética de animais com baixa capacidade de dispersão. Em adição, uma outra implicação imediata é de que a estabilidade histórica da Caatinga deve ser levada em consideração durante o delineamento de estratégias de preservação da biodiversidade, como por exemplo, a criação de unidades de conservação.

1.1 Objetivo geral

Obter estimativas de diversidade genética e elucidar padrões filogeográficos em populações do escorpião *J. rochae* em escala regional, buscando identificar potenciais refúgios e/ou centros de diversificação em ambientes de Caatinga, bem como possíveis barreiras ao fluxo gênico da espécie, utilizando para isso, modelos de distribuição de espécies e análises moleculares do gene mitocondrial citocromo C oxidase I (COI).

1.2 Objetivos específicos

- 1 Verificar se populações de *J. rochae* distribuídas em áreas de Caatinga historicamente estáveis apresentam maiores níveis de diversidade genética e estabilidade demográfica quando comparadas às populações de áreas instáveis;
- 2 Avaliar a possível influência do Planalto da Borborema no fluxo gênico e estruturação genética de populações de *J. rochae*;
- 3 Investigar a existência de possíveis associações cronológicas entre os eventos de diversificação intraespecífica em *J. rochae*, e a história geoclimática da região do Planalto da Borborema;
- 4 Inferir potenciais mudanças espaciais no nicho climático de *J. rochae* na região do Planalto da Borborema, ocorridas a partir do último período interglacial.

2 REFERENCIAL TEÓRICO

2.1 Aspectos biogeográficos da Caatinga

As florestas tropicais sazonalmente secas (FTSSs), são caracterizadas pelo predomínio de vegetação decídua, ocorrendo em regiões com regimes pluviométricos irregulares, e inferiores a 1.800 mm por ano. (Pennington *et al.*, 2009). Durante muito tempo, as formações de vegetação aberta da América do Sul (Caatinga, Cerrado e Chaco) foram classificados em um único bloco, representando a diagonal (ou corredor) de vegetação aberta que se estende da região nordeste do Brasil, até a região centro-norte da Argentina (Vanzolini, 1974; Bucher, 1982; Werneck, 2011). Essa classificação foi sustentada por diversos estudos envolvendo vertebrados (Short, 1975; Mares *et al.*, 1985; Vitt, 1991) e invertebrados (Camargo & Becker, 1999).

Nos dias atuais a Caatinga, Cerrado e Chaco são regiões ecológicas distintas, resultantes de diferentes padrões geomorfológicos, climáticos e biogeográficos (Pennington *et al.*, 2000, 2009; Motta *et al.*, 2002; Spichiger *et al.*, 2004; Werneck, 2011; Queiroz *et al.*, 2018). Consequentemente, os diferentes processos biogeográficos ocorridos nas referidas regiões ecológicas foram responsáveis por moldar a distribuição da biodiversidade atual, influenciando a história demográfica das populações, e promovendo eventos de especiação e/ou diversificação de linhagens (Werneck *et al.*, 2011; Apgaua *et al.*, 2014; Oliveira *et al.*, 2015; Bartoletti *et al.*, 2017; Leal *et al.*, 2018).

Dentre as florestas sazonalmente secas da região Neotropical, a Caatinga é uma região ecológica situada, em sua maior parte, no semiárido Brasileiro, abrangendo uma área de 912.529 km² (Silva *et al.*, 2018). Trata-se, portanto, do maior núcleo contínuo de FTSS do Novo Mundo (Queiroz *et al.*, 2018). Em termos

florísticos, os padrões de distribuição da vegetação da Caatinga são fortemente influenciados por fatores climáticos, edáficos e geomorfológicos da paisagem, que resultam na distinção de dois grupos fitofisionômicos da Caatinga – a Caatinga cristalina, e a Caatinga sedimentar (Moro *et al.*, 2015; Queiroz *et al.*, 2018).

As conformações do relevo e os diferentes tipos de solo estão entre os principais filtros ambientais que permitem, ou restringem, o estabelecimento de comunidades vegetais na Caatinga (Tabarelli *et al.*, 2003; Santos *et al.*, 2012; Moro *et al.*, 2016). A Caatinga cristalina, por exemplo, possui ampla distribuição, ocorrendo em solos rasos, porém ricos em nutrientes, originados a partir de rochas cristalinas, enquanto que a Caatinga sedimentar, restringe-se aos solos sedimentares, profundos, mas geralmente com baixo teor de nutrientes (Moro *et al.*, 2015, 2016; Queiroz *et al.*, 2018). Em adição, diversos estudos têm demonstrado as complexas relações de similaridade e divergência entre as comunidades vegetais da Caatinga cristalina e da Caatinga sedimentar, evidenciando padrões distintos de riqueza e distribuição de plantas nesses dois ambientes (Gomes *et al.*, 2006; Cardoso & Queiroz, 2007; Costa *et al.*, 2015).

De acordo com Ab'Sáber (1974), as fitofisionomias típicas de Caatinga, definidas pelo autor como “verdadeiros sertões”, ocorrem nas depressões interplanálticas do nordeste do Brasil, originadas a partir de aplanções neogênicas, isto é, ocorridas na segunda metade do Cenozoico, que compreende o final do período Terciário e início do Quaternário. Ainda segundo o referido autor, as fitofisionomias da Caatinga não sofreram grandes alterações em decorrência das oscilações climáticas do Quaternário, a ponto de serem substituídas por outras floras. Ao invés disso, a Caatinga se expandiu durante as épocas mais secas do Quaternário, por meio de corredores formados entre a Bacia Amazônica e o

Planalto Central, alcançando áreas que atualmente são ocupadas pelo Cerrado (Tricart, 1958; Ab'Sáber, 1974, 1977). Os modelos de expansão da Caatinga durante os períodos secos do Quaternário foram elaborados em concordância com os ensaios paleoclimáticos formalmente propostos por Damuth & Fairbridge (1970), os quais, apontam a persistência de climas áridos e semiáridos na região Sul-Americana equatorial durante o Quaternário, mostrando ainda, que os padrões de deslocamento vertical da célula de alta pressão do Atlântico Sul foram responsáveis por controlar o clima do Brasil durante o Quaternário, propiciando o estabelecimento de períodos secos durante as glaciações, e períodos mais úmidos em intervalos interglaciais.

Após os estudos realizados pelo geógrafo Aziz Ab'Sáber na década de 1970, diversos autores buscaram avaliar as possíveis relações entre as oscilações históricas dos biomas Neotropicais de vegetação aberta, e os padrões atuais de distribuição da biodiversidade (Werneck, 2011), utilizando como modelo, diversas espécies de vertebrados (Mares *et al.*, 1985; Carnaval & Bates, 2007; Carnaval & Moritz, 2008; Werneck *et al.*, 2015; Thomé *et al.*, 2016; Lima *et al.*, 2017; Costa *et al.*, 2018; Castro *et al.*, 2019), e invertebrados (Brown *et al.*, 1974; Magalhães *et al.*, 2014; Miranda *et al.*, 2017; Peretolchina *et al.*, 2018), bem como elementos da flora (Prado & Gibbs, 1993; Oliveira *et al.*, 1999; Santos *et al.*, 2007; Werneck *et al.*, 2011; Menezes *et al.*, 2016; Souza *et al.*, 2018).

Considerando os potenciais efeitos das variações climáticas ocorridas na região Neotropical no final do Plioceno e durante o Pleistoceno, Haffer (1969), propôs a hipótese dos refúgios pleistocênicos (HRP), para elucidar as possíveis origens da diversidade de aves Amazônicas. A HRP prediz que a fragmentação e a substituição espaço-temporal das florestas úmidas neotropicais, por

fitofisionomias abertas (Savanas), promoveram eventos de especiação alopátrica (por vicariância) em diferentes táxons (Haffer *et al.*, 1969; Wüster *et al.*, 2005). Subsequentemente, a HRP tem sido debatida por muitos autores (Hoorn *et al.*, 2010; Werneck, 2011; Werneck *et al.*, 2011; Turchetto-Zolet *et al.*, 2013), gerando inúmeras controvérsias fundamentadas em inconsistências nas premissas postuladas pela HRP. Provavelmente, a extensão que os biomas de vegetação aberta da região Neotropical alcançaram em seu ápice de expansão durante o Quaternário, e o tempo de divergência entre espécies e linhagens de diversos táxons neotropicais, correspondem às principais fontes de controvérsias relacionadas à HRP (Moritz, 2000; Colinvaux *et al.*, 2001).

A partir de dados palinológicos, Colinvaux *et al.* (2001) enfatizaram que as oscilações climáticas do Quaternário não foram suficientes para promover a substituição da floresta Amazônica por outras fitofisionomias, implicando apenas em mudanças populacionais nas comunidades vegetais do referido bioma. Adicionalmente, Cheng *et al.* (2013), por meio de datação absoluta de isótopos de oxigênio em espeleotemas, forneceram indícios de maiores concentrações pluviométricas no oeste da Amazônia durante o último período glacial, enquanto que na face leste, o clima seco foi mais pronunciado. Ainda segundo os autores, a estabilidade climática foi responsável por manter a biodiversidade do oeste da Amazônia, contrariando o princípio da fragmentação predita pela HRP.

Um segundo aspecto controverso da HRP ainda muito discutido em estudos atuais, refere-se ao tempo de diversificação de linhagens e/ou espécies. Se consideramos estritamente as premissas da HRP estabelecidas por Haffer (1969), estamos assumindo que os eventos de especiação ocorreram em um intervalo de tempo que abrange o Terciário-Quaternário, que em outras palavras, significa que

a diversificação da fauna amazônica é um fenômeno relativamente recente. Há evidências, no entanto, de que a cladogênese em diversos grupos taxonômicos que ocorrem em florestas úmidas da América do Sul precede essa janela de tempo (Moritz *et al.*, 2000; Rull, 2006; Couvreur *et al.*, 2011; Fritz *et al.*, 2011; Patel *et al.*, 2011; Álvarez-Presas *et al.*, 2014; Kozak *et al.*, 2015; Gehara *et al.*, 2017). Provavelmente, os exemplos mais explícitos de incompatibilidade cronológica entre a diversificação da fauna neotropical e as premissas postuladas pela HRP, são ilustrados por Moritz *et al.* (2000), em que apenas 7 de 125 eventos de diversificação de pequenos mamíferos da América do Sul ocorreram em uma escala de tempo que coincide com Pleistoceno, o que também foi observado em 18 de 64 eventos de especiação em grupos de aves Sul-Americanas. Em adição, processos de diversificação ocorridos antes do período Quaternário também são reportados para grupos taxonômicos de florestas secas da América do Sul (ex. Pennington *et al.*, 2009; Pirie *et al.*, 2009; De-Nova *et al.*, 2012).

É importante lembrar, que a HRP apresentada por Haffer (1969), foi elaborada unicamente a partir de dados de populações atuais de aves, desconsiderando, portanto, informações paleoecológicas (Bush & Oliveira, 2006), e a atuação de rios e outros corpos d'água como potenciais barreiras ao fluxo gênico das populações distribuídas na região Amazônica (Connor, 1986). Assim, não existe uma harmonia consensual entre as premissas estabelecidas pela HRP (*stricto sensu*), e a história biogeográfica de diversos táxons distribuídos na região Neotropical (Knapp & Mallet, 2003).

2.2 Núcleos de estabilidade climática da Caatinga

As FTSSs presentes na América do Sul possuem um padrão de distribuição disjunta, em que as maiores unidades de vegetação contínua são representadas por três grandes blocos: o núcleo Caatinga (NC), situado majoritariamente no nordeste do Brasil; o núcleo Misiones, localizado ao longo das bacias hidrográficas dos rios Paraná-Paraguai; e o núcleo Piedmont, distribuído em uma região que compreende o noroeste da Argentina e sudoeste da Bolívia (Prado & Gibbs, 1993; Prado, 2000; Werneck *et al.*, 2011). O núcleo Caatinga representa o mais extenso bloco contínuo de FTSS da região Neotropical (Prado & Gibbs, 1993; Pennington *et al.*, 2000, 2009; Werneck *et al.*, 2011). Historicamente, modelos de distribuição de espécies e análises palinológicas, indicam um padrão dinâmico de distribuição da vegetação contida no NC, o que significa dizer que apesar de sua grande extensão contínua, existem subnúcleos dentro do NC que podem ser distinguidos entre si de acordo com diferentes níveis de estabilidade climática (Werneck *et al.*, 2011).

Apesar de ter sido formulada sob a perspectiva das florestas úmidas neotropicais, a HRP proposta por Haffer (1969) e discutida no tópico anterior, está indiretamente associada com dinâmica da distribuição das FTSSs. Esta associação torna-se evidente se consideramos que as oscilações climáticas ocorridas entre o final do período Terciário (atualmente dividido nos períodos Paleógeno e Neógeno) e o período Quaternário, foram fundamentais para os processos de alternância entre as florestas úmidas e as florestas de vegetação aberta na região Neotropical (Haffer, 1969, Webb, 1978; Behling, 1998; Prado *et al.*, 2012).

Partindo da premissa de que as mudanças climáticas ocorridas ao longo do Quaternário foram responsáveis por promover eventos de expansão e retração das

FTSSs da América do Sul, Werneck et al. (2011) puderam então definir áreas de estabilidade climática situadas no NC, a partir de predições de ocorrência de ambientes favoráveis para estabelecimento de FTSSs em diferentes épocas do período Quaternário, as quais, foram validadas por meio de dados palinológicos. As áreas de estabilidade climática do NC estabelecidas por Werneck et al. (2011), condizem com a classificação das ecorregiões da Caatinga propostas por Velloso et al. (2002) (Figura 1), situando-se, na maior parte da Depressão Sertaneja Setentrional, Complexo Ibiapaba-Araripe, e Complexo da Chapada Diamantina. Há também, duas grandes áreas de estabilidade climática na Depressão Sertaneja Meridional, intercaladas por uma área instável, representada por uma diagonal que se estende desde a região centro-leste da Depressão Sertaneja Meridional, até a região centro-sul do Planalto da Borborema (Figura 2).

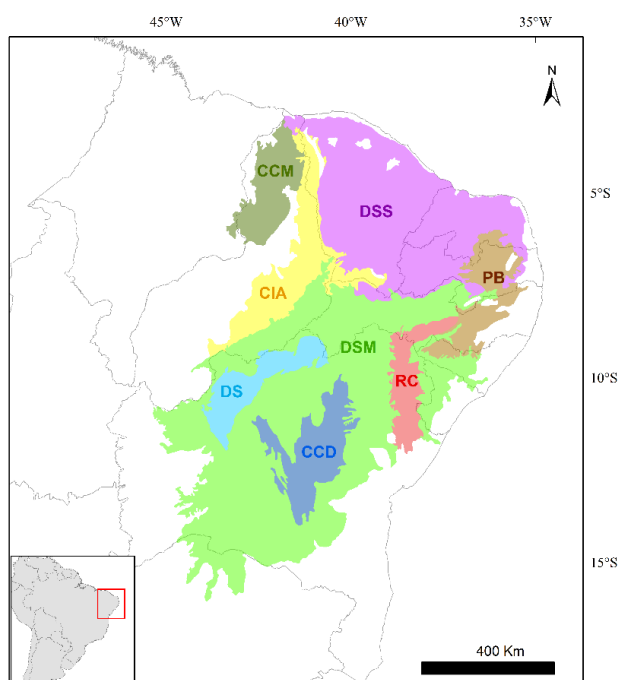


Figura 1. Ecorregiões propostas para a Caatinga: Complexo Campo Maior (CCM); Complexo da Chapada Diamantina (CCD); Complexo do Ibiapaba-Araripe (CIA); Depressão Sertaneja Meridional (DSM); Depressão Sertaneja Setentrional (DSS); Dunas do São Francisco (DS); Planalto da Borborema (PB); e Raso da Catarina (RC). Adaptado de Velloso et al. (2002).

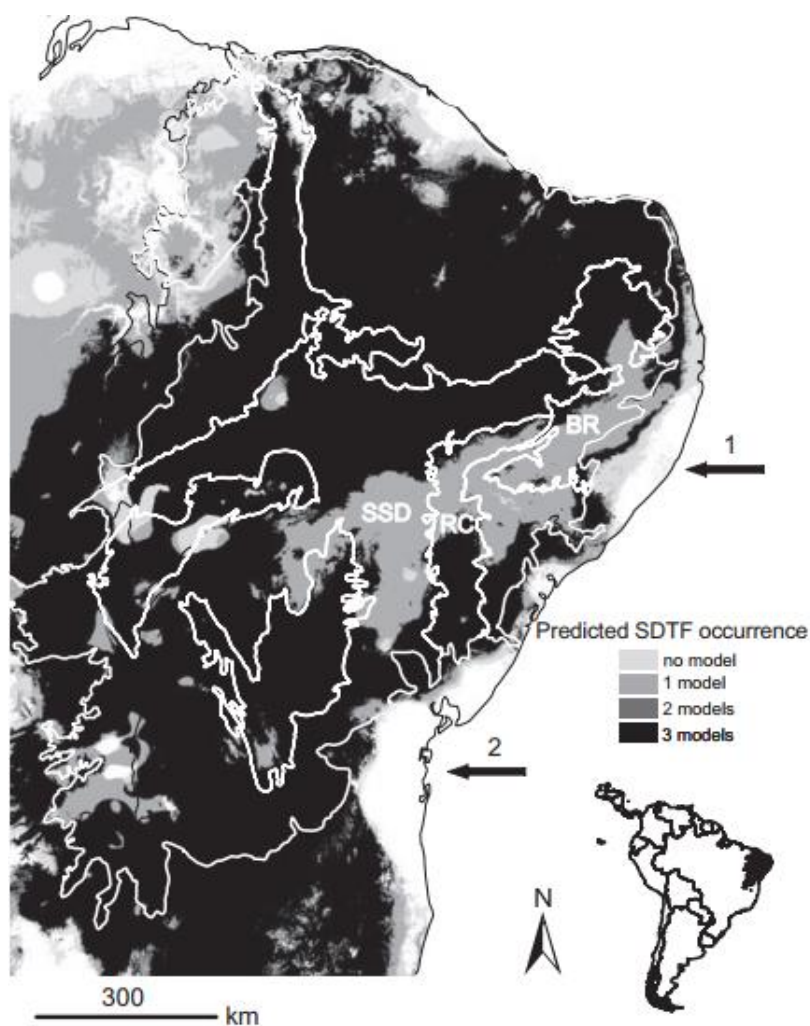


Figura 2. Representação das áreas de estabilidade (preto) e instabilidade (tons de cinza) climática da Caatinga gerada a partir de modelos de circulação atmosférica. As setas representam as áreas de alta estabilidade climática da Mata Atlântica, as quais, coincidem com os centros de diversidade Pernambuco (1), e Bahia (2). Nestas áreas, nenhum dos modelos indicaram a ocorrência de FTSSs. As linhas brancas delimitam as ecorregiões da Caatinga: Planalto da Borborema (BP), Raso da Catarina (RC), Depressão Sertaneja Meridional (SSD). Fonte: Werneck et al. (2011).

As inúmeras discussões sobre a origem e os padrões de distribuição da biodiversidade Sul-Americana no espaço e no tempo são comumente associadas a hipóteses formuladas sob a perspectiva das florestas úmidas (ex. Haffer, 1969; Nelson *et al.*, 1990; Fernandes *et al.*, 2012, 2013; Leite & Rogers, 2013). De fato, uma grande parcela do conhecimento sobre a biogeografia dos táxons da América

do Sul é oriunda de estudos com espécies típicas de florestas úmidas, como a floresta Amazônica (Morrone, 2000; Aleixo & Rossetti, 2007; Alfaro *et al.*, 2011; Dagosta & Pinna, 2017) e a mata Atlântica (Costa *et al.*, 2000; Cabanne *et al.*, 2008; Martins, 2011; Saraiva & da Silva, 2016) ou ainda, de remanescentes de florestas úmidas distribuídas dentro dos domínios das florestas de vegetação aberta, como os brejos de altitude (Costa, 2003). Por outro lado, a história biogeográfica da biota distribuída nas florestas de vegetação aberta da América do Sul tem recebido pouca atenção (Prado & Gibbs, 1993; Wüster *et al.*, 2005; Werneck *et al.*, 2011, 2012; Bueno *et al.*, 2016).

Uma das principais hipóteses biogeográficas referentes aos biomas de vegetação aberta da América do Sul é a hipótese do arco pleistocênico (HAP), formulada por Prado & Gibbs (1993). A HAP assume que o padrão atual de distribuição disjunta das florestas de vegetação aberta da América do Sul, resulta da fragmentação de uma área ancestral contínua, que se estendeu, em sentido diagonal, do nordeste do Brasil, até o noroeste da Argentina (Prado & Gibbs, 1993). O ápice da expansão das florestas de vegetação aberta teria ocorrido durante o Pleistoceno, mais especificamente, em um intervalo de tempo que corresponde ao último máximo glacial (21.000 – 12.000 anos antes do presente), quando o clima frio e seco propiciou a expansão das florestas de vegetação aberta, ao mesmo tempo em que as florestas úmidas se retraíram (Ab'Sáber, 1977; Prado & Gibbs, 1993).

Em casos específicos, a HAP tem sido adotada para explicar a distribuição de determinados táxons em diferentes escalas de tempo e espaço (ex. Werneck & Colli, 2006; Caetano *et al.*, 2008), porém, há também exemplos em que a história biogeográfica de muitas espécies não condiz com as premissas

estabelecidas pela HAP (ex. Magalhães *et al.*, 2014; Côrtes *et al.*, 2015, Bueno *et al.*, 2016). Em nível de paisagem, diversos estudos florísticos têm refutado a HAP (Oliveira *et al.*, 1999; Pessenda *et al.*, 2010; Werneck *et al.*, 2011, 2012; Sobral-Souza *et al.*, 2015; Leite *et al.*, 2016; Arruda *et al.*, 2018). Por exemplo, Arruda *et al.* (2018), a partir de modelos de distribuição ecológica e uso de dados palinológicos, estimaram que durante o último máximo glacial, a expansão dos biomas de vegetação aberta do Brasil se restringiu às áreas ecotonais. Os modelos propostos por Arruda *et al.* (2018) apontam ainda um clima frio e úmido na maior parte do Brasil, refutando uma das asserções da HAP, que prediz um clima frio e seco na referida região (Ab'Sáber, 1977; Prado & Gibbs, 1993). Similarmente, Werneck *et al.* (2011) demonstraram que os modelos de distribuição das FTSSs neotropicais ao longo do Quaternário são incompatíveis com as premissas propostas pela HAP, denotando que as condições climáticas do último máximo glacial não foram favoráveis para a expansão das FTSSs, e que ao invés disso, a distribuição das FTSSs fora ainda mais disjunta nesse período. Um padrão semelhante foi também observado em áreas de Cerrado, que segundo Werneck *et al.* (2012), atingiu seu ápice de retração durante o último máximo glacial.

Como pode ser observado, a distribuição histórica dos biomas de vegetação aberta da América do Sul é um tema relativamente bem abordado na literatura (ex. Werneck, 2011; Werneck *et al.*, 2011; 2012; Arruda *et al.*, 2018). No entanto, o principal objetivo desses estudos tem sido a categorização e a delimitação dos referidos biomas neotropicais ao longo do espaço e do tempo. Enquanto isso, os efeitos da variação espaço-temporal desses biomas sobre a biota que os habitam ainda é um tema pouco explorado (Turchetto-Zolet *et al.*, 2013). Da mesma forma, são escassos os estudos que buscam validar as áreas de alta estabilidade climática

previamente preditas (ex. Werneck *et al.*, 2011, 2012; Arruda *et al.*, 2018) como refúgios históricos e/ou centros de diversificação em ambientes de vegetação aberta (Werneck *et al.*, 2011, 2012; Turchetto-Zolet *et al.*, 2013), sobre tudo em áreas de Caatinga (Werneck *et al.*, 2011, 2015).

2.3 A Filogeografia e os modelos de distribuição de espécies na identificação de centros de diversificação

Nas últimas décadas, o desenvolvimento de novas ferramentas computacionais contribuiu substancialmente para o progresso da biogeografia em escala global, especialmente quando se trata de modelos de distribuição de espécies (Guisan & Thuiller, 2005; Thuiller *et al.*, 2005; Dormann, 2007; Ficetola *et al.*, 2007; Sunday *et al.*, 2012; Mainali *et al.*, 2015). Conceitualmente, os modelos de distribuição de espécies (MDEs) correspondem ao conjunto de técnicas e algoritmos implementados para predizer a potencial distribuição de um táxon em diferentes escalas de tempo e espaço (Guisan & Thuiller, 2005; Elith & Leathwick, 2009).

Utilizando registros de ocorrência georreferenciados (presença de espécies) e um conjunto pré-selecionado de variáveis bióticas e abióticas, os MDEs são capazes de predizer a potencial distribuição de um determinado táxon em áreas não amostradas (Guisan & Zimmermann, 2000; Guisan & Thuiller, 2005). Esses modelos podem ser aplicados para elucidar inúmeras questões na biologia, como por exemplo, para predizer e / ou explicar padrões atuais de distribuição da biodiversidade (Pineda & Lobo, 2009; Raes *et al.*, 2009; Pérez & Font, 2012; Moraes *et al.*, 2014), estimar áreas de ocorrência de espécies raras (Williamns *et al.*, 2009; Marini *et al.*, 2010; Sousa-Silva *et al.*, 2014; Tronstad *et al.*, 2018),

ameaçadas (Crawford & Hoagland, 2010; Chunco *et al.*, 2013; Sousa-Silva *et al.*, 2014), e/ou invasoras (Young *et al.*, 2013); inferir possíveis respostas da biodiversidade frente às mudanças climáticas (Bond *et al.*, 2011; Padonou *et al.*, 2015), e delimitar ambientes prioritários para conservação (Syfert *et al.*, 2014; Padonou *et al.*, 2015; Angelieri *et al.*, 2016; Heuner *et al.*, 2016; Kaky & Gilbert, 2016; Fois *et al.*, 2018), dentre outras aplicações (uma revisão detalhada pode ser consultada em Guisan & Thuiller, 2005).

Um dos principais recursos dos MDEs é a possibilidade de projetar modelos de distribuição baseados em variáveis ambientais do período presente, sob condições ambientais de períodos passados ou futuros. Na literatura, essa técnica se chama transferabilidade (Randin *et al.*, 2006; Fitzpatrick & Hargrove, 2009), e tem sido utilizada principalmente em estudos de conservação, em que a intenção principal é estimar a distribuição de áreas favoráveis para a ocorrência de determinados táxons no presente, e também em cenários futuros hipotéticos (Franklin, 2013; Porfirio *et al.*, 2014), auxiliando na compreensão dos possíveis efeitos das mudanças climáticas sobre composição e distribuição espaço-temporal da biodiversidade (Sinclair *et al.*, 2010; Hamann & Aitken, 2012).

Os MDEs são ferramentas praticamente indispensáveis em estudos de biogeografia histórica, utilizados especialmente para formular hipóteses a priori (Werneck *et al.*, 2011, 2012, Sobral-Souza *et al.*, 2015; Arruda *et al.*, 2018), ou para testar hipóteses resultantes de outros tipos de análises (Porto *et al.*, 2012; Bryson Jr *et al.*, 2014; Oke *et al.*, 2014). Em ambos os casos, é extremamente importante incorporar aos MDEs, informações sobre a história natural e a ecologia das espécies-alvo, evitando assim, uma análise puramente matemática (“data-driven

analysis”, Mellert *et al.*, 2011) e com pouco significado biológico (Dormann *et al.*, 2011).

Em estudos biogeográficos, os MDEs são comumente utilizados em paralelo com outros tipos de análises, como por exemplo, em inferências filogeográficas (Richards *et al.*, 2007), fundamentadas na teoria da coalescência (Kingman, 1982a, 1982b). A filogeografia é considerada uma disciplina intimamente vinculada à biogeografia, que integra métodos de disciplinas voltadas à microevolução (Etologia, Demografia, Genética de Populações), e macroevolução (Geografia Histórica, Paleontologia, Filogenia), para tratar dos princípios e processos que governam a distribuição geográfica de linhagens genealógicas no espaço e no tempo, especialmente em espécies filogeneticamente próximas (Avice, 2000, 2009; Templeton, 2004). Assim, a filogeografia permite o surgimento de novas perspectivas sobre a evolução das espécies, a partir da compreensão das interações entre a ecologia e a história natural das espécies-alvo, bem como dos eventos geológicos e influências ambientais e geográficas da paisagem (Knowles, 2009). Apesar de sua origem recente (meados da década de 1980), o número cada vez maior de publicações no decorrer dos anos é um reflexo fidedigno do notório interesse global pela filogeografia (Avice, 2009; Turchetto-Zolet *et al.*, 2013; Tu *et al.*, 2016).

Os fundamentos teóricos da filogeografia estão intrinsicamente entrelaçados com os princípios da teoria da coalescência. De forma simplificada, a teoria da coalescência assume que, em um modelo neutralista de evolução, é possível traçar a história genealógica de um conjunto de linhagens atuais, “retrocedendo” até o seu ancestral comum mais próximo (Nordborg, 2007). O processo de coalescência é a descoberta do ancestral comum mais próximo de um número qualquer de

linhagens. Em outras palavras, dizemos que um número n de linhagens coalesce, quando alcançamos o ancestral comum mais próximo dessas linhagens (Rosenberg & Nordborg, 2002).

O modelo neutro comumente adotado em simulações de coalescência é o de Wright-Fisher, amplamente utilizado em genética de populações e análises evolutivas de variações em loci gênicos ao longo do tempo (Gory *et al.*, 2018). Teoricamente, o tamanho constante das populações, a ausência de gerações sobrepostas, a panmixia e a ausência de seleção, constituem as principais conjecturas estabelecidas pelo modelo de Wright-Fisher (Hudson, 2002; Parida, 2010, 2012). Adicionalmente, ao adotar o referido modelo, assume-se que as novas gerações são formadas a partir de amostragens randômicas (com reposição) de um número qualquer de indivíduos parentais da geração atual (Nordborg, 2007). Considerando que as premissas propostas pelo modelo de Wright-Fisher podem parecer pouco realísticas quando aplicadas às comunidades reais, existem diversas derivações que se adequam às situações encontradas em populações naturais (Parida, 2012), incluindo modelos que consideram a ação de seleção balanceadora e direcional, a presença de gerações sobrepostas, e as flutuações no tamanho populacional ao longo do tempo, além de algoritmos capazes de operar com loci recombinantes (Hein *et al.*, 2005).

Por se tratar de linhagens genealógicas, o modelo de Wright-Fisher, e consequentemente, a teoria da coalescência, foram originalmente elaborados para analisar dados moleculares de natureza haplotípica (Kingman, 1982a). Por conta disso, os marcadores moleculares de linhagem podem ser vistos como o carro-chefe das análises filogeográficas (Avise, 2000, 2009). Dentre essa classe de marcadores, o DNA mitocondrial (mtDNA) é, sem dúvidas, um dos recursos mais

utilizados na filogeografia de espécies animais (Rahman *et al.*, 2010; Miraldo *et al.*, 2012; Turchetto-Zolet *et al.*, 2013). Na maioria dos animais, o mtDNA é uma molécula de estrutura circular, com tamanho médio de aproximadamente 16,5 kb (Bernt *et al.*, 2013), e de simples organização estrutural (Figura 3). Em artrópodes por exemplo, o mtDNA é organizado em uma molécula de DNA circular, usualmente contendo regiões que codificam 13 proteínas, 22 RNAs transportadores, dois RNAs ribossômicos, e uma região controladora (replicação/transcrição) rica em bases A+T (Chen *et al.*, 2011).

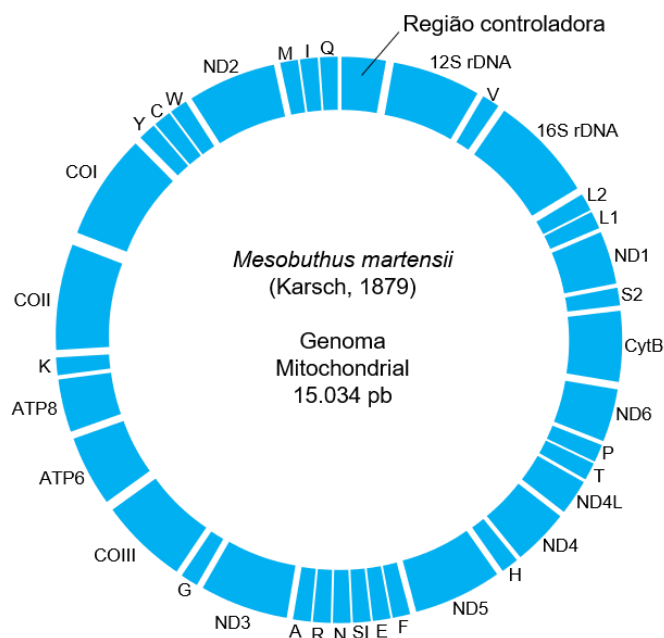


Figura 3. Representação esquemática da estrutura do genoma mitocondrial do escorpião *Mesobuthus martensii* (Karsch, 1879), mostrando a posição relativa de cada gene, e também da região controladora. Os genes que codificam as subunidades da NADH desidrogenase são indicados de maneira respectiva à cada subunidade (ND1-6, 4L), o mesmo foi feito para os genes responsáveis pelas subunidades da citocromo C oxidase (COI, COII, COIII), e para os genes das subunidades 6 e 8 da ATPase (ATP6, ATP8). Os genes de RNAs transportadores são indicados por letras referentes aos seus respectivos aminoácidos, seguindo o código da IUPAC; citocromo b (Cytb). Adaptado de Choi *et al.* (2007).

Além da estrutura simples, o mtDNA de animais possui algumas outras propriedades desejáveis em estudos de filogeografia. A transmissão uniparental do mtDNA assegura a natureza haplotípica da informação genética, requerida como input em análises filogeográficas (Avise, 2009). Uma vez que a genealogia das linhagens é comumente representada por árvores topológicas, marcadores haplotípicos são desejáveis para gerar tais topologias, isto porque, regiões autossômicas podem ser transmitidas por ambos os sexos, e as análises com essas sequências normalmente resultariam em um diagrama de rede, ao invés de uma topologia (Parida, 2012). A ausência de recombinação no mtDNA de animais (com raras exceções – ver Piganeau *et al.*, 2004; Tsaousis *et al.*, 2005) também é uma importante propriedade a ser considerada, garantindo que a origem da variação encontradas nas sequências do mtDNA decorre apenas de mutações acumuladas ao longo do tempo (Avise, 2009).

Com o uso adequado de modelos evolutivos e estimativas confiáveis de calibração (ex. registros fósseis, eventos geológicos, taxas de mutação), as variações no mtDNA podem ser aplicadas em simulações de relógio molecular, resultando em uma filogenia datada, que indica o tempo de divergência em cada nó presente na árvore (Rutschmann, 2006). Finalmente, a alta taxa de substituição de nucleotídeos presente no mtDNA possibilita a realização de inferências filogeográficas em nível intraespecífico, e o elevado número de cópias por célula (com raros casos de heteroplasmia) garante, do ponto de vista técnico, uma maior facilidade de obtenção do material genético (Avise, 2009).

Regiões informativas do DNA nuclear (nDNA) também podem ser incorporadas em análises filogeográficas. Normalmente, os *loci* de cópias únicas do nDNA são os mais desejáveis em inferências filogeográficas (Avise, 2009). No

entanto, por apresentar propriedades biológicas distintas, o nDNA não pode ser analisado sob a mesma perspectiva do mtDNA. Por exemplo, a taxa de substituição de nucleotídeos do nDNA é muito inferior à encontrada no mtDNA de animais (Hellberg, 2006; Fischer *et al.*, 2013). Há também, uma certa dificuldade em isolar haplótipos em regiões de nDNA de organismos diploides, além dos possíveis ruídos que a recombinação dessas regiões pode causar nas análises (Avice, 2009). Apesar disso, existem estratégias metodológicas para contornar a problemática dos marcadores de nDNA em inferências filogeográficas. Dentre elas está a própria seleção da região de interesse, e o uso de ferramentas adequadas de bioinformática para analisá-las. Nesse sentido, o DNA ribossômico nuclear (rnDNA) pode ser útil em estudos de filogeografia (Ji *et al.*, 2003). Em eucariotos, o rnDNA está organizado em blocos repetidos em tandem (Figura 4), compostos por uma região não transcrita, seguida de uma região externa transcrita, e dos genes ribossômicos 18S, 5.8S e 28S, intercalados pelas regiões internas transcritas 1 (entre os genes 18S e 5.8S) e 2 (entre os genes 5.8S e 28S) (Hillis & Dixon, 1991).

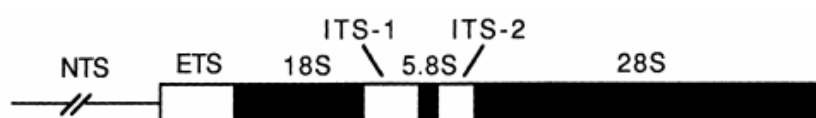


Figura 4. Estrutura do DNA ribossômico (rnDNA) comumente encontrado no genoma nuclear de eucariotos, mostrando uma unidade repetitiva composta pelas seguintes regiões: região não transcrita (NTS), espaçador externo transcrito (ETS), espaçadores internos transcritos 1 e 2 (ITS1, ITS2), e genes das subunidades ribossômicas 18S, 5.8S, e 28S. Fonte: Hillis & Dixon (1991).

Por se tratar de uma região repetitiva, o rnDNA ocorre com maior frequência no genoma, facilitando sua amplificação (Hwang *et al.*, 1998). Além disso, o rnDNA está sujeito a evolução em concerto, o que acaba levando à homogeneização

dessas regiões em nível intraespecífico ao mesmo tempo em que as divergências interpesecíficas se acentuam ao longo do tempo (Hillis & Dixon, 1991), tornando mais viável a identificação de haplótipos a partir de regiões do rDNA.

Considerando que o rDNA é herdado por ambos os sexos, a sua incorporação em análises filogeográficas pode reduzir o viés de uma possível dispersão assimétrica relacionada ao sexo, o que poderia levar a diferentes padrões filogeográficos entre machos e fêmeas de uma mesma espécie / população, se analisadas unicamente a partir de marcadores de mtDNA (Palumbi & Baker, 1994). Em adição, a implementação de marcadores de nDNA pode reduzir substancialmente os intervalos de confiança de inúmeros tipos de análises (ex. tamanho populacional, fluxo gênico, crescimento populacional, e tempo de divergência), levando a resultados mais fidedignos e com maior significado biológico (Zink *et al.*, 2005). Finalmente, quando usados em conjunto, as diferentes taxas de substituição de nucleotídeos encontradas em marcadores de mtDNA e rDNA, permitem uma resolução mais refinada em filogenias datadas (Ojanguren-Affilastro *et al.*, 2017), assim como a construção de redes de haplótipos com diferentes padrões de aglomeração, que variam conforme o nível de conservação dos marcadores utilizados.

2.4 Seleção do organismo modelo

Eventos geológicos e ciclos climáticos são comumente interpretados como os motores que impulsionam as mudanças na paisagem ao longo do tempo, modelando os padrões de biodiversidade (Gillespie & Roderick, 2014). Em nível populacional, tanto os eventos geológicos quanto os ciclos climáticos deixam marcas no DNA dos indivíduos. Por exemplo, alterações de paisagem provocadas

pelos ciclos climáticos geralmente são rápidas (sob a perspectiva do tempo geológico), podendo deixar traços detectáveis de efeito gargalo ou efeito fundador, os quais, reduzem a diversidade genética das populações (Álvarez-Presas *et al.*, 2011). Por outro lado, as mudanças na paisagem ocorridas em decorrência de eventos geológicos, normalmente ocorrem ao longo de uma larga escala de tempo, permitindo a adaptação progressiva das espécies, a estabilização do tamanho populacional, e o acúmulo de variação genética intrapopulacional (Hewitt, 2004; Álvarez-Presas *et al.*, 2011). Essa série de fatores é frequentemente observada em populações que habitam áreas com alta estabilidade climática (ex. Hugall *et al.*, 2002; Carnaval *et al.*, 2009; Yannic *et al.*, 2013; Faye *et al.*, 2016).

Apesar das alterações da paisagem influenciarem efetivamente a dinâmica das populações, não existe um mecanismo único de resposta a essas alterações que englobe todas as espécies. Entretanto, espécies especialistas e com baixa capacidade de dispersão acabam sendo preferíveis em estudos de filogeografia, uma vez que os padrões de distribuição e diversidade desses táxons são fortemente correlacionados com a estabilidade histórica de seus habitats (Hewitt, 2004; Avise, 2009; Álvarez-Presas *et al.*, 2011; Silva *et al.*, 2017). Assim, sob condições desfavoráveis, espera-se que as espécies estenotópicas persistam apenas em áreas estáveis dentro de sua distribuição geográfica geral (refúgios), e que na ausência de mecanismos de longa dispersão, as populações de tais espécies estão mais propícias às flutuações demográficas (Silva *et al.*, 2017).

Em termos práticos, muitos escorpiões reúnem características ecológicas desejáveis em estudos de filogeografia, incluindo baixa capacidade de dispersão (Polis *et al.*, 1985; Habel *et al.*, 2012; Bryson Jr *et al.*, 2013a, 2016; Štundlová *et al.*, 2019), e significativa especificidade de micro-habitat (Bryson Jr *et al.*, 2013a,

2013b, Monod *et al.*, 2013; Ojanguren-Affilastro *et al.*, 2016; Esposito & Prendini, 2019). Em escorpiões da família Buthidae C.L. Koch, 1837 presume-se que a baixa capacidade de dispersão, e consequentemente, o limitado fluxo gênico entre as populações, atuam como mecanismos fundamentais em processos de especiação (Husemann *et al.*, 2012). Teoricamente, quanto mais limitada for a dispersão e mais generalista for o táxon ancestral (assumindo os escorpiões como exemplo), maior será a riqueza esperada de espécies em um ambiente mutável com barreiras geográficas recorrentes (Lourenço *et al.*, 2016).

Até 2014, eram registradas 28 espécies de escorpiões em áreas de Caatinga (Porto *et al.*, 2014). Com o avanço dos estudos sobre taxonomia dos escorpiões (Esposito *et al.*, 2017; Lira *et al.*, 2017; Santos-da-Silva *et al.*, 2017), a estimativa atual, é de que pelo menos 33 espécies de escorpiões ocorram na Caatinga. Dentre essas espécies, o gênero *Jaguajir* Esposito, Yamaguti, Souza, Pinto-da-Rocha & Prendini, 2017, reúne duas espécies com distribuição em áreas de Caatinga e Cerrado, e uma terceira, distribuída nas savanas do extremo norte do Brasil.

Os escorpiões do gênero *Jaguajir* são caracterizados pelo seu grande porte (5-11 cm), e pela presença de estruturas estridulatórias situadas nos pentes, e no terceiro esternito (detalhes em Esposito *et al.*, 2017). Estima-se que o gênero *Jaguajir* tenha se originado na América do Sul (assim como o ancestral de todas as espécies da subfamília Centruroidinae Kraus, 1955) há cerca de 34,5 milhões de anos atrás, em um evento cladogenético que separou dois grandes clados, o primeiro contendo o ancestral dos atuais gêneros *Jaguajir*, *Troglophalurus* Lourenço, Baptista & Giupponi, 2004, *Rhopalurus* Thorell, 1876, *Physoctonus* Mello-Leitão, 1934, e *Ischnotelson* Esposito, Yamaguti, Souza, Pinto-da-Rocha & Prendini, 2017, todos com distribuição restrita à América do Sul; e o segundo clado,

que comporta os gêneros *Centruroides* Marx, 1890, e *Heteroctenus* Pocock, 1893, distribuídos na América Central e América do Norte (Esposito & Prendini, 2019). Embora não haja citação direta, os resultados apresentados por Esposito & Prendini (2019) sobre a história evolutiva da subfamília Centruroidinae, possivelmente corroboram a hipótese postulada por Lourenço (1990, 1994), de que o Escudo Brasileiro representa um possível centro de dispersão para a família Buthidae.

O gênero *Jaguajir* ocorre unicamente em formações de vegetação aberta da América do Sul (Lourenço, 1986, 1994). A espécie *J. rochae* ocorre quase que exclusivamente em áreas de Caatinga, com registros também para áreas ecotonais (Caatinga-Cerrado, Caatinga-Mata Atlântica), ao passo em que *Jaguajir agamemnom* (C.L. Koch, 1839) se distribui majoritariamente em áreas de Cerrado. A distribuição geográfica da terceira espécie do gênero, *Jaguajir pinto* (Mello-Leitão, 1932), está restrita às áreas de vegetação aberta situadas no extremo norte do Brasil, abrangendo ainda uma pequena parte do sudeste da Venezuela e oeste da Guiana (Figura 5) (Esposito *et al.*, 2017).

Dentre as espécies que ocorrem na Caatinga, *J. rochae* pode ser facilmente identificada pelo seu grande porte (60-72 mm), e também pela sua coloração característica, em tons de amarelo-claro (Lenarducci *et al.*, 2005; Esposito *et al.*, 2017) (Figura 6), além da presença de um aparelho estridulatório capaz de emitir sons audíveis (Prendini *et al.*, 2009).

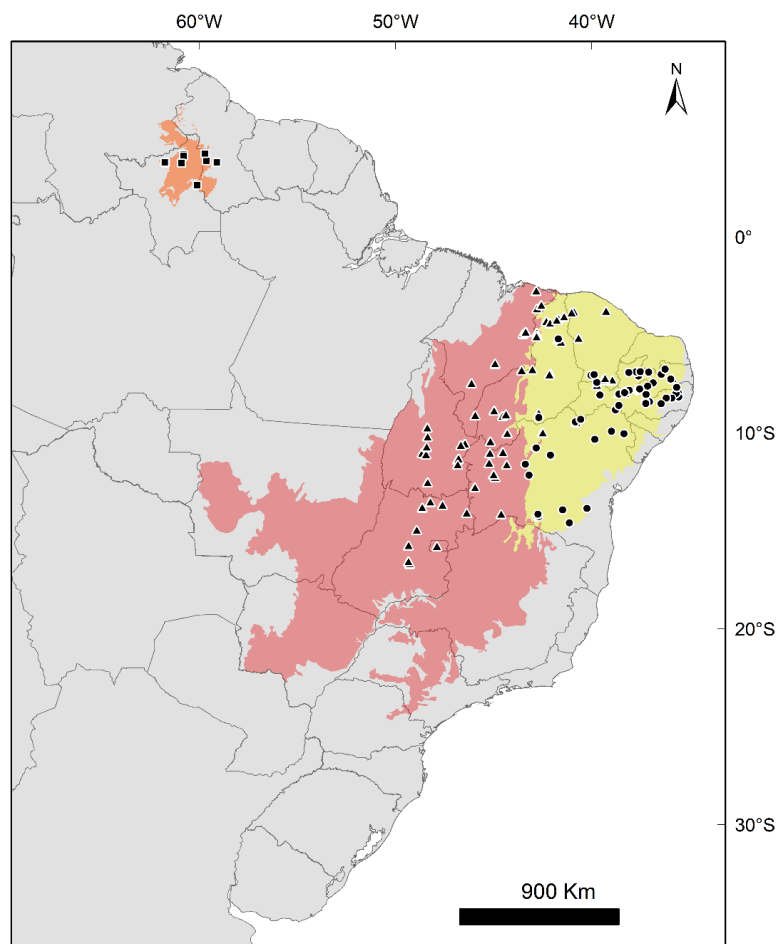


Figura 5. Distribuição geográfica das espécies do gênero *Jaguarir*: *J. rochae* (círculos), *J. agamemnom* (triângulos), e *J. pinto* (quadrados). As áreas coloridas representam as delimitações da Caatinga (amarelo), e Cerrado (vermelho) de acordo com as projeções do Instituto Brasileiro de Geografia e Estatística (IBGE, 2004). A área em laranja, denota as formações abertas do tipo Savana, situadas no extremo norte do Brasil (Olson et al., 2001), em que são registradas ocorrências de *J. pinto*. Adaptado de Esposito et al. (2017).

Do ponto de vista prático, a facilidade de identificação, a elevada abundância em inventários de fauna conduzidos na Caatinga (Carmo et al., 2013; Lira et al., 2018), e o porte robusto dos espécimes de *J. rochae*, facilitam o processo de obtenção e análise de material genético em análises filogeográficas. Além disso, a ampla distribuição de *J. rochae* em áreas de Caatinga (Figura 5) permite que esse táxon possa ser utilizado para inferir a atuação de áreas de estabilidade climática

da Caatinga como centros de diversificação de linhagens. Adicionalmente, a presença dessa espécie em áreas ecotonais, permite também, a comparação direta entre os padrões de história demográfica das populações que habitam a região central da distribuição da espécie, com as populações das áreas ecotonais, produzindo interpretações que podem ser relevantes no contexto da dinâmica das paisagens (ex. biomas, ecorregiões).



Figura 6. Espécime adulto de *Jaguajir rochae* (fêmea), fotografado no Parque Estadual Mata da Pimenteira, uma unidade de conservação da Caatinga situada na região central de Pernambuco (fevereiro, 2016).

Por fim, inúmeros estudos têm demonstrados que os escorpiões são excelentes modelos em estudos de biogeografia e filogeografia (Gantenbein & Largiadèr, 2003; Sousa *et al.*, 2012; Graham *et al.*, 2012, 2013a, 2013b, 2014; Bryson Jr *et al.*, 2013a, 2013b, 2014, 2016; Miller *et al.*, 2014; Ceccarelli *et al.*, 2016, 2017; Ojanguren-Affilastro *et al.*, 2017; Esposito & Prendini, 2019), contudo, os estudos dessa natureza são ainda incipientes na América do Sul, e restritos às espécies que ocorrem no sul e na costa oeste da América do Sul (ex. Ceccarelli *et al.*, 2016, 2017; Ojanguren-Affilastro *et al.*, 2017).

3 Insights on the role of historical habitat stability and geological barriers on lineage diversification processes in Caatinga environments revealed by landscape genetics of the scorpion *Jaguajir rochae* (Scorpiones: Buthidae)

Stênio Ítalo A. Foerster^{1*}, Bárbara Natieli S. Pereira¹, André Felipe de A. Lira², Juliana F. Lima³, Ricardo Pinto-da-Rocha³ and Valdir de Queiroz Balbino¹

¹ Departamento de Genética, Universidade Federal de Pernambuco, Avenida da Engenharia, S/N, CEP 50740-580, Recife, Brazil.

² Departamento de Morfologia e Fisiologia Animal, Universidade Federal Rural de Pernambuco, Rua Dom Manoel de Medeiros, S/N, CEP 52171-900, Recife, Brazil.

³ Departamento de Zoologia, Universidade de São Paulo, CEP 05508-090, São Paulo, Brazil.

* Author for correspondence: stenioit@gmail.com

Manuscrito a ser submetido ao Periodico *Journal of Biogeography* (Qualis A2)

Abstract

Aim The Caatinga domain is the largest refugium of Seasonally Dry Tropical Forest (SDTF) on the Neotropics, and recent studies improved our knowledge on the historical biogeography of Caatinga taxa at broad scales. Although important, such conclusions may be masking deterministic processes related to the genesis and maintenance of genetic diversity at regional or local scales. We performed a series of phylogeographical analysis on the Caatinga scorpion *Jaguajir rochae*, to evaluate the potential role of geological barriers and SDTF stability of the genetic structuring processes at fine spatial scales.

Location Northeastern Brazil.

Taxon The Caatinga scorpion *Jaguajir rochae*.

Methods We sequenced a partial fragment of the mitochondrial gene cytochrome c oxidase I for 77 specimens from 11 sampling localities. Genetic diversity and historical demographic changes were accessed by several metrics. Bayesian methods were applied to infer genetic clusters, divergence dating, and migration models among populations, whereas niche suitability through the late-Quaternary was estimated by ecological niche models with optimized settings.

Results Populations from historically stable areas of SDTF display high levels of genetic diversity, whereas those from changeable environments are genetically undifferentiated. Most mitochondrial lineages diverged after the uplift of the Borborema Plateau (Pleistocene), but this massif persists as a prominent barrier to the gene flow of *J. rochae*. The climatic niche suitability of *J. rochae* remained relatively stable since the Last Interglacial, but spatial shifts were detected at local scale, agreeing with the proposed migration model.

Main conclusions For arid-adapted taxa with low dispersal capabilities, such as the scorpion *J. rochae*, historical refugia, and geomorphological features within the Caatinga domain are prominent forces acting under the genesis, and maintenance of genetic diversity.

Keywords SDTF, Phylogeography, Biogeography, invertebrates, Caatinga

Introduction

The historical stability of the South American open vegetation biomes becomes a convergence point among recent investigations (Werneck, Costa, Colli, Prado, & Sites, 2011; Werneck, Gamble, Colli, Rodrigues, & Sites, 2012; Sobral-Souza, Lima-Ribeiro, & Solferini, 2015; Costa et al., 2017; Arruda, Schafer, Fonseca, Solar, & Fernandes-Filho, 2018). In Seasonally Dry Tropical Forests (SDTFs) for example, paleoecological niche modelling and palynological data support the hypothesis of multiple stable nuclei of SDTFs on the neotropics (Werneck et al., 2011). Accordingly, many authors agree that most part of the spatial extent of neotropical SDTFs remained stable during the Quaternary (Sobral-Souza et al., 2015; Arruda et al., 2018, but see Costa et al., 2017), and a weak interplay with adjacent rainforests (Amazon forest and Atlantic forest) was proposed, but restricted to ecotonal zones (Arruda et al., 2018).

Behind the discussions concerning the dynamics of the spatial extent of SDTFs over time, which has been stressed in the literature (e.g. Pennington, Prado, & Pendry, 2000; Pennington, Lavin, & Oliveira-Filho, 2009; Werneck, 2011; Werneck et al., 2011), the impacts of these changes on the structure of communities and/or populations has received increasing attention in the last years (e.g. Bartoletti, Peres, Fontes, da Silva, & Solferini, 2018; Oliveira et al., 2018; Foerster, Lira, & deSouza, 2019; Lanna et al., 2019). The expressive geographical extent of neotropical SDTFs, combined with its geomorphological features and habitat heterogeneity, certainly arouse the interest of naturalists to study the patterns and process behind the genesis of biodiversity in such environments. The Caatinga domain (CD) for example, offers a singular opportunity to explore the effects of niche and climate stability on the genetic structure of specific taxa, improving our knowledge on the historical modifications of the landscape promoted by cyclical climatic oscillations.

The CD is a geographically disjunct unity of SDTF, comprising a set of complex vegetation types – from cactus scrub to tall forests, but mostly represented by tree-dominated landscapes of (semi)deciduous species (Werneck et al., 2011). The strong seasonality of the rain season plays a pivotal role on the vegetation dynamics of the CD, impacting the aboveground net primary productivity (Moura & Lee, 2004; Menezes, Sampaio, Giongo, & Pérez-Marin, 2012; Salimon & Anderson,

2018), and the life cycles of its biodiversity (Vieira, Santana, & Arzabe, 2009; Rodrigues & Silva, 2014; Cavalcanti, Paiva, & França, 2016; Gonçalves, Cerqueira, Brasil, & Santos, 2017). The structure of biological communities of the CD derives from synergistic processes involving a strong niche conservatism and the effects of environmental conditions at local scale, resulting in complex landscapes marked by expressive levels of endemism and β -diversity (Pennington et al., 2009; Queiroz, Cardoso, Fernandes, & Moro, 2018).

Werneck et al. (2011) have demonstrated the presence of historically unstable regions within the CD, matching the ecoregions of Borborema Plateau (BOP), Raso da Catarina, and the Southern Sertaneja Depression (Velloso, Sampaio, & Pareyn, 2002). More specifically, the study of Werneck et al. (2011) proposes that the BOP is one of the major divisors between stable and unstable areas of SDTF, which is predicted to occur at western and eastern sides of the BOP, respectively. This longitudinal gradient is geographically close to the Pernambuco Atlantic forest refugium (Carnaval & Moritz, 2008), and the climatic fluctuations of the Quaternary may be related to the possible interplay (vegetation shifts) between Caatinga and Atlantic forest (Werneck et al., 2011).

Considering that habitat stability is a predictor of demographic immutability, genetic and taxonomic diversity, and endemism (Carnaval, Hickerson, Haddad, Rodrigues, & Moritz, 2009; Faye et al., 2016; Gutiérrez-Rodríguez, Barbosa, & Martínez-Solano, 2017; Camps, Martínez-Meyer, Verga, Sérsic, & Cosacov, 2018; Nogueras, Cordero, & Ortego, 2018; Lima-Rezende et al., 2019), it is reasonable to think that historically stable areas within the CD act like a refugia, harboring demographically stable populations with high levels of genetic diversity (Werneck et al., 2011; Binks, Gibson, Ottewill, Macdonald, & Byrne, 2019) and several exclusive haplotypes. In this sense, phylogeographical structure of low-dispersal taxa can be a valuable resource to test hypotheses regarding the landscape evolution and its effects on geographical patterns of genetic diversity (Avise, 2009; Álvarez-Presas, Carbayo, Rozas, & Riutort, 2011; Silva et al., 2017). For such purposes, scorpions can be seen as a good biological model (Ceccarelli et al., 2016; Ceccarelli, Pizarro-Araya, & Ojanguren-Affilastro, 2017; Ojanguren-Affilastro, Adilardi, Mattoni, Ramírez, & Ceccarelli, 2017; Esposito & Prendini, 2019) because they are relatively stenotopic animals (Bryson Jr, Savary, & Prendini, 2013; Ojanguren-Affilastro et al., 2016), with low dispersal capabilities (Polis, McReynolds, & Ford, 1985; Bryson Jr,

Savary, Zellmer, Bury, & McCormack, 2016; Štundlová, Šmíd, Nguyen, & Šťáhlavský, 2019). Thus, it could be expected that the historical processes relating the evolution of biological traits in scorpions (i.e. physiological tolerances, population demography, geographical distribution, taxonomic and genetic diversity) can shed light on the biogeographic steps that shape the current biodiversity patterns at regional and local scales.

Among the 33 described species of scorpions that occur in the CD (Porto, Carvalho, de Souza, Oliveira, & Brescovit, 2014; Lira, Pordeus, & Albuquerque, 2017; Esposito, Yamaguti, Souza, Pinto-da-Rocha, & Prendini, 2017; Santos-da-Silva, Carvalho, & Brescovit, 2017), the genus *Jaguajir* comprises three species, of which, *Jaguajir rochae* (Borelli, 1910) is a typical element of Caatinga environments (Esposito et al., 2017), while the other two species, *Jaguajir agamemnom* (C.L. Koch, 1839) and *Jaguajir pinto* (Mello-Leitão, 1932), are mainly distributed in Cerrado formations of central and northern regions of Brazil, respectively (Teruel & Tietz, 2008; Lourenço, 2008; Esposito et al., 2017). The geographical distribution of *J. rochae* extends across most part of the CD, comprising the entire extent of the longitudinal gradient of habitat stability mentioned above. In this study, therefore, we explore the genetic population structure of *J. rochae* under a phylogeographical framework, integrated with paleodistribution modelling, to test for genetic clustering of spatially structured haplogroups in historically stable and unstable areas within the CD.

We are motivated by the hypothesis that the BOP can be a physical barrier responsible for structuring populations of species adapted to dry climatic conditions, such as those present in Caatinga environments. We derive this main goal into five pertinent questions: I) are populations of *J. rochae* structured around the BOP? II) Are within-lineage diversification events occurred into the time slice comprising the late Quaternary climatic oscillations, or BOP uplift? III) Is the BOP a stringent barrier to the gene flow of *J. rochae* populations? IV) Are populations from historically stable areas (sensu Werneck et al., 2011) more genetically diverse and demographically stable than those from changeable environments? VI) Has niche suitability of *J. rochae* changed considerably around the BOP since the late-Quaternary climatic scenarios? We advocate that the answer for these questions will improve the knowledge on the biogeography and natural history of the CD, which, although recent advances, remains as one of least studied and most threatened

environments in South America (Santos, Leal, Almeida-Cortez, Fernandes, & Tabarelli, 2011; Silva, Barbosa, Leal, & Tabarelli, 2018).

Materials and Methods

Sample collection

Specimens of *J. rochae* were collected in 11 localities from both sides of the BOP (Fig. 1a), covering both historically stable and unstable areas of SDTF predicted by Werneck et al. (2011). Using tweezers and self-made ultraviolet light lanterns (wavelength = 395-400 nm), we sampled 77 specimens, with an average of seven individuals per locality (ranging from four to 10 specimens). Scorpions were collected at night (20h00min-23h00min), during random walks performed by at least two collectors, and kept alive until DNA extraction. Sample collection was authorized by the Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio), under the process number 66699-1.

Molecular procedures

Absolute ethanol was used to fix specimens under laboratory conditions. One leg from the left side of all individuals was removed and preserved in absolute ethanol at -20°C as a tissue sample for DNA extraction. The remaining of each scorpion was stored as voucher specimens and deposited at the arachnological collection of Universidade Federal de Pernambuco (Recife, Brazil). Genomic DNA extraction was performed using the Chelex-100® resin (Bio-Rad Laboratories) protocol described in Casquet, Thebaud, & Gillespie (2012). Partial fragments of mitochondrial DNA were sequenced for the protein-coding gene cytochrome c oxidase I (COI), which was successfully used in several phylogeographical studies of scorpions and other arachnids (e.g. Bartoletti et al., 2017; Graham, Jaeger, Prendini, & Riddle, 2013; Graham, Wood, Henault, Valois, & Cushing, 2017; Graham, Myers, Kaiser, & Fet, 2019; Harms, Roberts, & Harvey, 2019; Peres, Benedetti, Hiruma, Sobral-Souza, & Pinto-da-Rocha, 2019; Pfingstl et al., 2019).

The targeted gene was amplified by polymerase chain reaction, in a 25 µl volume, containing 12.5 µl of GoTaq® Colorless Master Mix (Promega), 7.5 µl of ddH₂O, 1.5 µl of each primer (forward and reverse), and 2 µl of undiluted DNA template. Forward and reverse primers are the same as described in Vink, Thomas,

Paquin, Hayashi, & Hedin (2005): C1-J-1718-spider and C1-N-2728-spider, respectively. The following thermal conditions were applied in PCR: an initial denaturation step of 5' at 95°C, 35 cycles of 95°C for 30", 52°C for 30" and 72°C for 45", and a final extension of 72°C for 10'. PCR products were checked for amplification in 1% agarose gel electrophoresis. Amplicons were purified using the Wizard® SV Gel and PCR Clean-Up System kit (Promega) and then sequenced in an automatic capillarity sequencer ABI PRISM 3500 Genetic Analyzer (Applied Biosystems).

Alignment and genetic diversity estimation

COI sequences were aligned in MAFFT 7 (Kato & Standley, 2013) using parameter configurations for closely related DNA sequences with global homology (G-INS-i strategy as iterative refinement method, and 1PAM/k=2 for parameter matrix). We use the sequence KY982232.1 from GenBank as a directional reference to the alignment of the remaining sequences. This sequence was also included in our dataset as a member of Serra Talhada (SER) population. Poorly aligned regions in the multiple sequence alignment were automatically trimmed with Gblocks (Castresana, 2000) using the stringent criterion to remove contiguous nonconserved positions. We use the 'ape' (Paradis & Schliep, 2018) and 'pegas' packages (Paradis, 2010) on the R environment (R Core Team, 2019) to compute descriptive metrics of genetic diversity, including the number of haplotypes (h), the haplotype diversity (Hd), and the nucleotide diversity (π).

Genetic clusters and population differentiation

Genetic groups were designed using the Bayesian clustering approach implemented in BAPS 6.0 (Corander, Sirén, & Arjas, 2008). For this analysis, we chose the grouping method that accounts for linkage dependences among loci (Corander & Tang, 2007), and set the number of clusters to vary between 1 and 20. The algorithm implemented in BAPS uses a genetic mixture analysis to compute the optimal number of clusters from a set of DNA sequences, which is selected by the maximization of the log marginal likelihood (Corander et al., 2008).

Spatial patterns among genetic lineages were graphically analyzed using a median-joining haplotype network produced in PopArt 1.7 (Leigh & Bryant, 2015), with a prior identification of redundant haplotypes in R software with the 'haplotype'

library (Aktas, 2019). The degree of genetic differentiation among haplogroups defined in BAPS (see Results) was measured by the Φ_{ST} metric (Excoffier, Smouse, & Quattro, 1992; Holsinger & Weir, 2009), as implemented in the 'strataG' R package (Archer, Adams, & Schneiders, 2016).

A hierarchical analysis of molecular variance (AMOVA) implemented in 'pegas' R package was applied to disentangle the genetic variation at inter and intra-population level, as well as those shared between haplogroups inferred by BAPS (Excoffier et al., 1992) – just for practical purposes, we use the term “population” to design all individuals from the same sampling locality. Isolation by distance was measured for the whole data set, and for each haplogroup defined in BAPS through Mantel tests implemented in the R package 'ade4' (Dray & Dufour, 2007), using as input, pairwise interpopulation matrices of genetic differentiation (Φ_{ST}), and geographical distance (km). Significance was achieved under 10,000 permutations.

Demographic changes

Recognizable signatures of demographic changes were inferred by neutrality tests – Tajima's D (Tajima, 1989), Fu's F_s (Fu, 1997), and R_2 statistic (Ramos-Onsins & Rozas, 2002) computed in Arlequin 3.5 (Excoffier & Lischer, 2010) and 'pegas' R library. To avoid misleading results due to technical limitations (i.e. sample size, DNA polymorphism), we calculated neutrality tests for the whole data set, and for each population with at least two haplotypes (Ramos-Onsins & Rozas, 2002; Grant, 2015). Statistical significance for each neutrality tests was achieved through 10,000 replications.

Divergence dating

Split divergence times among *J. rochae* haplotypes were accessed from a dated gene tree estimated in BEAST 1.10.4 (Suchard et al., 2018) under a Bayesian inference. We include the species *Jaguajir agamemnom* (GenBank, accession KY982209.1) as the outgroup, based on the phylogenetic hypothesis proposed by Esposito, Yamaguti, Pinto-da-Rocha, & Prendini (2018). Redundant haplotypes were removed using the 'haplotypes' R package. We then partitioned the sequences alignment in codon positions 1+2, and position 3, once asymmetric mutation rates are expected to occur at the third codon position of mitochondrial genes (Oberski et

al., 2018), which can affect the fit of substitution models (Powell, Barker, & Lanyon, 2013).

Best substitution models for each partition were selected under a Bayesian Information Criterion (BIC), as implemented in 'phangorn' R package (Schliep, 2011). Clock and substitution models were unlinked for each DNA partition, but trees were linked and adjusted to follow a Yule speciation tree prior (Gernhard, 2008). Two separate runs were performed in BEAST to generate dated phylogenies under the strict molecular clock prior, and uncorrelated lognormal clock prior. For both simulations, we ran BEAST for 50 million generations, sampling every 5000 generation. The clock rate of 0.008125 substitution/site/million years was applied to both simulations (strict and uncorrelated clock models), but for the uncorrelated lognormal clock, we set this rate as a mean for a normal distribution with a standard deviation of 0.0005. This substitution rate was specifically developed for scorpions and used to date split divergences of several South American species (Gantenbein, Fet, Gantenbein-Ritter, & Balloux, 2005; Ojanguren-Affilastro et al., 2017).

We use two mathematically distinct methods to guide the choose of the most appropriate clock model: the former is a frequentist approach based on maximum likelihood theory implemented in MEGA X (Kumar, Stecher, Knyaz, & Tamura, 2018), and the latter is the log Bayes Factor (LBF), computed from the log marginal likelihood of each clock model simulation generated in BEAST using both path sampling, and stepping stone sampling framework (Baele, Li, Drummond, Suchard, & Lemey, 2012; Baele, et al., 2013). LBF was computed in R, using the 'mtraceR' package (Pacioni et al., 2015), applying the log marginal likelihood values of each clock model as input. Convergence of MCMC chains were inspected in Tracer 1.7 (Rambaut, Drummond, Xie, Baele, & Suchard, 2018) taking the Effective Sample Size values as reference (ESS > 200). Final tree topology was assembled in TreeAnnotator 1.10.4 (Suchard et al., 2018) using a maximum clade credibility tree approach after discarding the first 25% of sampled trees as burn-in.

Migration models

We use Migrate 4.3 (Beerli & Palczewski, 2010) to test for nine explicit migration models (Fig. 2), evaluating the effect of the BOP as a physical barrier to the gene flow of *J. rochae*. Migrate 4.3 estimates migration patterns among user-defined population groups under a Bayesian framework, applying a thermodynamic

integration method to approach the marginal likelihood of each scheme (Beerli & Palczewski, 2010). The output of the Migrate-n provides specific parameters for each migration model, such as the mutation-scaled values of migration rates (M), and effective population sizes (Θ), from which, the number of migrants per generation can be calculated. We set Migrate to run a single MCMC run of 200,000 steps, sampling every 100, with a final burn-in of five million steps; swap intervals for the thermodynamic static heating were kept in default mode, and convergence of MCMC chain parameters was accessed by ESS values ($ESS > 200$). Our migration models were designed to test two hypotheses: 1) populations of *J. rochae* are geographically structured around the BOP, but migration occurs among them; and 2) there are a single panmictic population distributed along the sampling extent. We use a Python script provided by Peter Beerli (available at: <http://peterbeerli.com/downloads/scripts/>) to compute the LBF for each migration model tested in Migrate-n, prioritizing that with the greatest associated probability (Beerli & Palczewski, 2010; Graham et al., 2017). We then extract Θ and M values for the most likely migration model to obtain migration rates, in number of migrants per generation, as described in Gómez-Carballa et al. (2018).

Ecological niche modelling

We use ecological niche models (ENMs) to evaluate the niche stability of *J. rochae* through late Quaternary climatic oscillations, and also to predict the distance and direction of potential migration events from the last interglacial period (LIG, ~120-140 ka) to the Last Glacial Maximum (LGM, ~21 ka), and from the LGM to the present. ENMs were constructed with Maxent algorithm (Phillips, Anderson, & Schapire, 2006) implemented in 'dismo' R package (Hijmans, Phillips, Leathwick, & Elith, 2017). Maxent is a machine-learning tool that uses presence points and raster layers (explanatory variables) to predict the relative occupancy rate for a given species (Fithian & Hastie, 2013; Merow, Smith, & Silander, 2013; Merow, Allen, Aiello-Lammens, & Silander, 2016), outperforming others ENM methods, even working with few occurrence points (Elith et al., 2006; Merow et al., 2013). After a manual filtering process to remove records from non-natural environments (i.e. points within urban centers), we compiled 44 georeferenced presence points of *J. rochae*, obtained from several zoological collections (Tab. S1).

The spatial sampling extent for the modelling process was restricted to the latitudinal bounds of 2°20'-24°41'S, and longitudinal bounds of 60°6'-34°49'W, comprising the whole distribution of Caatinga and Cerrado formations (MMA, 2020), which is the typical habitat of scorpions from the genus *Jaguajir* (Esposito et al., 2017). As environmental predictors, we downloaded the 19 bioclimatic variables (2.5 arc-min resolution) from the WorldClim database (Hijmans, Cameron, Parra, Jones, & Jarvis, 2005), clipping it according to the spatial sampling extent cited above. To reduce model complexity and avoid multicollinearity problems, we use the 'usdm' R package (Naimi, Hamm, Groen, Skidmore, & Toxopeus, 2014) to retain only slightly autocorrelated bioclimatic variables ($r < 0.7$), with low contribution to the variance inflation factor ($VIF < 10$), following the recommendations of Guisan, Thuiller, & Zimmermann (2017), and Bond, Anderson, Henare, & Wehi (2019).

As a result, seven environmental predictors were used to generate the ENMs: maximum temperature of the warmest month (BIO5), temperature annual range (BIO7), mean temperature of warmest quarter (BIO10), mean temperature of coldest quarter (BIO11), annual precipitation (BIO12), precipitation of driest month (BIO14), and precipitation of warmest quarter (BIO18) (Hijmans et al., 2005). Presence points, and an additional of 10,000 random points generated in 'dismo' R package (background points), were partitioned in training and testing data using the 'checkerboard1' strategy, derived from the masked geographically structure approach described in Radosavljevic & Anderson (2014), and implemented on the 'ENMeval' R package (Muscarella et al., 2014). To minimize the spatial dependence of occurrence and background points, we scaled the checkerboard1 algorithm to partitioning the data set using 30×30 km grid cells. Optimal configuration for Maxent was estimated in 'ENMeval', allowing the regularization multiplier to vary between 0.5 and 4, and testing all feature classes available for Maxent (Fourcade, Besnard, & Secondi, 2017); best configuration scheme was chose by minimizing the associated AICc values, as recommended by Warren & Seifert (2011), and Radosavljevic & Anderson (2014). ENMs were constructed with current climate data, and then, projected to past climatic scenarios (LIG and LGM). For the LGM predictions, we assembled bioclimatic layers from the average values of three General Circulation Models: CCSM4 (Gent et al., 2011), MIROC (Watanabe et al., 2011), and MPI (Giorgetta et al., 2013). A complementary log-log transformation

was applied to the raw Maxent output, and the final predictions were interpreted as occurrence probability maps (Phillips, Anderson, Dudík, Schapire, & Blair 2017).

To assure reliable predictions, we use five metrics of model evaluation: the area under the receiver operating characteristic curve for the testing data (AUC_{TEST}), the difference between AUC_{TEST} and the AUC computed for the training data (AUC_{DIFF}), the true skill statistic (TSS), and omission rates based on minimum training presence threshold (OR0), and after excluding the 10% of the training localities with poorest predicted suitability (OR10). AUC_{TEST} ranges from 0 to 1, and estimates the ability of the model to differentiate predictions based on presence localities versus background points (Muscarella et al., 2014; Fourcade et al., 2017), while AUC_{DIFF} is expected to be positively correlated with model overfitting (Muscarella et al., 2014) and/or overparameterization (Warren & Seifert, 2011). The TSS is a threshold-dependent metric that take into account the balance between the true positive (i.e. sensibility) and negative (i.e. specificity) rates of the models (Allouche, Tsoar, & Kadmon, 2006; Liu, White, & Newell, 2011); accurate models are expected to have TSS near to 1 (Peres et al., 2019). OR0 and OR10 are threshold-dependent metrics used to measure model overfitting, with acceptable values near to 0 and 0.17, respectively (Wan, Wang, & Yu, 2011; Boria, Olson, Goodman, & Anderson, 2014; Muscarella et al., 2014; Wang, Wan, & Zhang, 2017).

Possible patterns consistent with local migration were mapped based on the spatio-temporal changes in niche suitability of *J. rochae* through the late Quaternary. To do that, we transform the occurrence probability maps into binary maps, using a threshold value that maximizes the sum of sensibility and specificity of each model (Liu, Berry, Dawson, & Pearson, 2005; Liu, Newell, & White, 2016). Then, we use Qgis 3.6 (QGIS Development Team, 2020) to obtain geographical centroids for every 30×30 km² grid cells in each predicted time (LIG, LGM, and current), estimating the hypothetical source of populations from one time slice to the next, by computing the length and direction between the centroid of the first time step to the nearest centroid of the following time step, that is, from the LIG to the LGM, and from the LGM to present (Gugger, Ikegami, & Sork, 2013). This analysis resulted in maps with vectors (arrows) representing possible migration distance and direction for each 30×30 km² grid cell.

Results

Genetic diversity

We obtained partial COI sequences for 77 specimens of *J. rochae*, that resulted in an alignment block of 685 bp with 73 variable sites, of which, 36 were parsimony-informative. Geographically restricted haplotypes accounts for 56.4% ($n = 44$) of the entire DNA data set, with a mean of 4.3 non-shared haplotypes per locality. CAR, and CUM were monomorphic populations in terms of haplotype composition, whereas AGU, AFO, and SER were the most diverse sampling localities, showing a specimen/haplotype ratio of 1:1, although expressive levels of haplotype diversity were also detected for other sampling localities from the western side of the BOP (Tab 1). Shared haplotypes and low levels of genetic diversity represent the populations from the eastern side of the BOP, although CAE population depicts low genetic diversity, but only exclusive haplotypes.

Population structure and differentiation

Bayesian analysis of population structure (BAPS) recovered two clusters that best represent the genetic similarities among *J. rochae* haplotypes (log-marginal likelihood = -1262.1571; posterior probability of cluster assignment = 0.9987). Accordingly, populations from the eastern side of the BOP grouped into a single cluster (eastern group), except CAE, that jointed into the second group (western group), together with all populations from the western side of the BOP (Fig. 1b). AMOVA indicated that the most expressive source of genetic variation comes from between groups comparisons (eastern vs western), which accounts for 44% of the variation (Tab. 2). We did not detected isolation by distance among populations within both western and eastern haplogroups, as well as for the whole data set ($P > 0.05$), but high and significant ($P < 0.01$) levels of genetic differentiation was reported for the whole data set ($\Phi_{ST} = 0.5361$), and western haplogroup ($\Phi_{ST} = 0.3557$). Populations from the eastern cluster, however, were genetically undifferentiated ($\Phi_{ST} = 0.2283$; $P > 0.05$). A star-shape pattern was recovered by haplotype network, depicting few substructures encompassing lineages from AFO, SER, and PAR populations (Fig. 1c). In addition, an hypothetical radiation from AGU population was suggested by the haplotype network (Fig. 1c).

Demographic changes

Neutrality tests resulted in significative values favoring the hypothesis of demographic expansion for the western haplogroup defined in BAPS, as well as for the whole data set (Tab. 1). Comparatively, non-significant values from neutrality tests were obtained for most sampling localities when it was analyzed separately, although punctual exceptions expressing genetic signatures of demographic expansion were detected within the eastern haplogroup (LIM), but most pronounced within the western haplogroup (Tab. 1).

Divergence dating

The strict clock model was rejected for the molecular data set, as evidenced by the maximum likelihood test implemented in MEGA X ($P < 0.01$). Accordingly, the uncorrelated relaxed clock was favored over the strict clock model with an assignment probability higher than 99% for both path sampling, and stepping-stone sampling strategies. The maximum clade credibility tree recovered by BEAST indicated an expressive level of genetic admixture among mitochondrial haplotypes (Fig. 3), evidenced by a low number of nodes with statistical support. However, an early split divergence including the haplotype J400 (restricted to AGU population) was estimated to be occurred around 3.05 Mya (95% HPD = 1.59-4.75), suggesting an early Plio-Pleistocene differentiation with subsequent radiation and diversification events occurring along the Pleistocene.

Migration models

Migrate-n recovered model D as the most likely hypothesis to explain the gene flow of *J. rochae* across the sampling extent (Tab. 3). Model D proposes an unidirectional pattern of gene flow, in which the western haplogroup receives immigrants from the eastern cluster (Fig. 2d). According to Θ and M parameters of model D (Tab. S2), the western haplogroup receives a mean number of 17.5 immigrants from the eastern haplogroup per generation. Although Migrate-n reported model D as the most suitable migration scheme, we cannot disregard model F as a possible alternative to explain the gene flow of *J. rochae* (Tab. 3). Model F suggests that AGU population acts like a migrant source, while gene flow occurs bidirectionally through the BOP (Fig. 2f). For model F, the group of populations from the western side of the BOP receives 67 times more immigrants

per generation from AGU than the populations from the eastern side of the BOP (Tab. S2). In addition, model F depicts an asymmetric gene flow between populations through the BOP, in which, the mean number of immigrants per generation from eastern to the western side is eight times greater than that for the reverse route.

Ecological niche modelling

Overall, the ENMs reached satisfactory values for all evaluation metrics ($AUC_{TEST} = 0.9107$; $AUC_{DIFF} = 0.0091$; $TSS = 0.70565$; $OR0 = 0.0192$; $OR10 = 0.0662$), and the levels of autocorrelation between bioclimatic variables were kept low (Fig. S1) thus, we assume their predictions as reliable proxies for niche suitability of *J. rochae*. In terms of spatial shifts, the ENM projections suggest that the climatic conditions that make up the ecological niche of *J. rochae* remained relatively stable around the BOP since the LIG (Fig. 4a-c, f). Nevertheless, a general weak displacement of the climatic niche suitability of *J. rochae* thought the late-Quaternary could be detected, moving diagonally, from northeastern to southwestern from LIG to LGM (Fig. 4d), and in the reverse course from LGM to present (Fig. 4e). At local scale, however, spatial shifts in habitat suitability of *J. rochae* were detected from the LGM to the present, moving from the eastern side of the BOP towards the continent (Fig. 4e).

Discussion

Genetic diversity

In this study, we use the scorpion *J. rochae* as a biological model to understand how climatic and geomorphological features present in Caatinga landscapes could be related to the spatial distribution of genetic diversity, and lineage diversification in low-dispersal animals at regional scale. We found an expressive number of mitochondrial haplotypes ($n = 44$), which, in turn, is a product of the additive effect of the high haplotype diversity within each sampling site (Tab. 1). The spatial distribution of such genetic diversity is not random. Instead, sampling localities distributed on the western side of the BOP depicted high levels of both nucleotide and haplotype diversity, as well as an expressive proportion of geographically restricted haplotypes (Tab. 1). Such localities may have experienced

few climatic changes over time when compared to those distributed near to the Pernambuco Atlantic Forests refugium (Carnaval & Moritz, 2008; Werneck et al., 2011). Thus, the spatial pattern of genetic diversity in populations of *J. rochae* reported by us, is in accordance with the expectation that genetic diversity can be preserved by historical habitat stability (Carnaval et al., 2009; Faye et al., 2016; Paz et al., 2019). In addition, the shifts in SDTF stability around the BOP can also be a potential drive of phylogeographic endemism (*sensu* Carnaval et al., 2014), once almost all haplotypes from the western side of the BOP are spatially restricted to their sampling localities, while shared haplotypes were reported for localities from the eastern side of the BOP (Tab. 1).

Population structure and differentiation

The geographical distribution of the two haplogroups recovered by BAPS, suggests that the BOP offers, in some degree, a resistance to the gene flow among populations of *J. rochae*, leading to a substantial genetic differentiation among these haplogroups, as evidenced by AMOVA results (Tab. 2). Such isolation makes populations in the eastern side of the BOP more susceptible to the effects of historical vegetation shifts occurred on the eastern side of this massif along the Quaternary (Werneck et al., 2011).

In fact, recent diversification events have been observed among populations from the eastern side of the BOP (Fig. 3), suggesting that Quaternary climate change may have an effect on the lineage diversification in these populations, which is not seen in the counterparts from the westerns side of the BOP. Moreover, the role of environmental features as physical barriers for structuring communities and/or populations in Caatinga environments has received increased attention (e.g. Werneck, Leite, Geurgas, & Rodrigues, 2015; Oliveira et al., 2015, 2018; Lanna et al., 2019). These studies, however, are centered on the main hypothesis that the São Francisco river is one of the most prominent barriers influencing the patterns of diversity and endemism in Caatinga environments, giving few insights about the potential role of topographical constraints in limiting the gene flow among populations.

In the present case, the clusters obtained in BAPS did not perfectly match the eastern and westerns sides of the BOP, because CAE population grouped into the haplogroup that contains all populations from the western side of this geological formation, suggesting the existence of points of less resistance in the BOP, which

allow the gene flow between populations of *J. rochae*. Even so, our results indicate that BOP can represent a significant barrier to the gene flow of low-dispersal organisms, especially those adapted to the semi-arid conditions of Caatinga environments, favoring the genetic differentiation among populations of such taxa by keeping them geographically isolated.

Demographic changes

Although our initial expectation that habitat stability would be a predictor of demographic stationarity, we find an opposite pattern for *J. rochae*, in which genetic imprints of demographical expansion, and stability, were reported for the western and eastern haplogroups, respectively. In fact, populations with historical evidence of demographic expansion are likely to occur in stable environments, such as those with long-term size stability (Finn, Bogan, & Lytle, 2009). Interestingly, neutrality tests suggests that some populations of *J. rochae* from climatically stable areas (AFO, AGU, and PAR) may have experienced demographical expansion events (Tab. 1). Some of them (AFO, and PAR) represented the main geographical substructures within the western haplogroup (Fig. 1c). Eventually, signs of demographical expansions, and reduction in genetic diversity are attributed to bottlenecks suffered by populations from historically unstable environments in South America (e.g. Prado, Haddad, & Zamudio, 2012; Novaes, Ribeiro, Lemos-Filho, & Lovato, 2013; Thomé & Carstens, 2016), including arachnids (e.g. Peres et al., 2015; Bartoletti et al., 2017).

It seems not to be the case for the western haplogroup of *J. rochae*, once the evidences of demographic expansion depicted by neutrality tests are accompanied by high levels of intrapopulation genetic diversity (Tab. 1). The demographic history of this haplogroup reveals a complex pattern, reinforcing the hypothesis of an initial radiation starting from AGU population (Fig. 1c), with subsequent local adaptation and demographic expansion evidenced by genetically structured and growing populations such as AFO, and PAR. Comparatively, the weakness or the absence of molecular evidences for a demographic expansion in the eastern haplogroup (Tab. 1), suggest that their populations may be persisting under suboptimal environmental conditions or spatial limitation, because they are surrounded at west by the BOP, and at east by the Atlantic Forest. This assumption is reinforced by the high migration rate (in number of migrants per generation) reported from eastern

populations to the western ones, as evidenced by the two best migration models recovered by Migrate-n (Tab. S2). Thus, besides the importance of habitat stability on the preservation of genetic diversity, such environmental propriety can also facilitate the radiation and demographic expansion of low-dispersal taxa in Caatinga environments.

Divergence dating

The historical refugia within the Caatinga domain proposed by Werneck et al. (2011) favored the permanence of ancestral lineages, and the maintenance of genetic diversity in populations of *J. rochae* through late-Quaternary. However, the diversification processes behind the origin of such genetic diversity are older, comprising a time slice that encompasses the Plio-Pleistocene period (Fig. 3). Consequently, it implies that the constraint minimum age of about 6.4 Mya for the end of BOP uplift (Morais-Neto, Hagarty, Karner, & Alkmim, 2009; Oliveira & Medeiros, 2012; Costa, Amorim, & Mattos, 2018) is much older than the processes of lineage diversification in *J. rochae* reported by us. Allopatric diversification in Rivulidae killifishes seems to be driven by the uplift of the BOP (Costa, Amorim, & Bragança, 2013; Costa et al., 2018), but there is no information about the role of the BOP on the genetic differentiation of terrestrial animals. Here, however, the chronological incompatibility provides a counter-argument to a vicariant-based explanation for the genetic differentiation among populations of *J. rochae* mediated by the BOP uplift.

Similarly, there are at least two reasons to refuse an hypothesis of genetic differentiation favored by habitat fragmentation due to climatic cycles: 1) the spatial configuration of Caatinga vegetation though Plio-Pleistocene lead to some controversies, in which, some advocate in favor of the maintenance/expansion of Caatinga in that time (e.g. Passoni, Benozzati, & Rodrigues, 2008; Werneck et al., 2015), while others propose a scenario of expansion of humid forests occurring at the expense of dry forests, such as Caatinga (Batalha-Filho, Fjeldså, Fabre, & Miyaki, 2013); 2) even if population of *J. rochae* was segregated in geographically isolated demes, as a result of possible climatic changes in the Plio-Pleistocene, the expected absence of gene flow combined with the accumulation of genetic variation over time, would certainly cause isolation by distance among demes, which was not supported by our data. In addition, the overall low number of shared haplotypes

(Tab. 1) suggests the absence of secondary contact among demes supposedly separated by habitat fragmentation at that time. Thus, an alternative hypothesis, is that genetic divergences in populations of *J. rochae* may have occurred in the presence of gene-flow, which proved to be a common process (Nosil, 2008) observed in some lizards of Cerrado (Werneck et al., 2012), and Caatinga environments (Oliveira et al., 2015; Werneck et al., 2015). Furthermore, the expressive levels of haplotype diversity, and the high number of geographically restricted lineages within the western haplogroup (Tab. 1), agree with the Ecological Gradient Hypothesis (Endler, 1973; Machado et al., 2019), suggesting that lineages may diverged in response to local selective pressures within a continuum landscape with clinal environmental proprieties. If it is the case, the genetic admixture and the absence of reciprocally monophyly between the western and eastern haplogroups (Fig. 3), as well as the lack of isolation by distance, can be explained by the presence of gene flow among populations of *J. rochae*. A similar diversification process seems to be occurred among lineages of the lizard *Phyllopezus pollicaris* (Spix, 1825) in Caatinga environments (Werneck et al., 2012). These examples denote that diversification processes in Caatinga environments are driven by multifactorial patterns that must take into account the historical synergistic effects of geological and climatic events at regional and local scale.

Migration models and ENMs

The asymmetric gene flow of populations of *J. rochae* is evidenced by the high migration rates observed from the eastern side of the BOP to the continent (Tab S1 migrate models). Such asymmetry reinforces the hypothesis that populations from the eastern side of the BOP may be persisting under suboptimal environmental conditions (e.g. spatial limitation, recurrent habitat fragmentation, scarcity of resources). In this sense, migration and population expansion are biological processes that require a period of establishment of the new lineages into the new areas (Endler, 1973), thus, the sings of demographical stability in the eastern haplogroup of *J. rochae* (Tab. 1), as well as the increased migration toward the continent (Tab. S2) can be an indicative that migration started recently in populations from the eastern side of the BOP, suffering a potential increase since the LGM (Fig. 4e). Furthermore, the results obtained in Migrate-n, indicate that the BOP is not a stringent barrier to the gene flow of *J. rochae*, although our results

showed that this massif substantially reduces the migration rate among populations. Because the scorpion *J. rochae* are well-adapted to the xeric conditions of Caatinga environments (Lira, deSouza, & Albuquerque, 2018; Foerster et al., 2019), one can suppose that indirect factors inherent to the BOP, such as the orographic rainfalls, and the establishment of humid semideciduous forests (Rodal & Nascimento, 2006; Rodal, Barbosa, & Thomas, 2008), could be the main source of resistance to the gene flow of this scorpion. However, a purely orographic contribution of the BOP in limiting the gene flow of this scorpion cannot be disregarded, once our ENMs suggest that the BOP provides suitable climatic conditions to the occurrence of this scorpion (Fig. 4f).

In addition, most part of the BOP is within an ecotonal zone between Caatinga and Atlantic Forest (Olson et al. 2001), which, in theory, could favor the permanence of *J. rochae* in xeric-vegetation patches, or promotes its dispersion via xeric corridors through the BOP. In such a context, the geological history of the landscape is a deterministic factor influencing lineage diversification, speciation rates, and community structure in scorpions (Husemann, Schmitt, Stathi, & Habel, 2012; Foord, Gelebe, & Prendini, 2015). In South American environments for example, the new habitats and climatic regimes originated from the Andean uplift, favored the interspecific diversification of scorpion genus *Brachistosternus* Pocock, 1893 (Ceccarelli et al., 2016, 2017). In addition, Foerster et al. (2019) use similarity patterns in species composition to demonstrate that scorpion assemblages in montane forest remnants of northeastern Brazil clustered in spatially structured groups mainly separated by the BOP. Thus, even with the marked chronological association between diversification processes of neotropical species, and the Quaternary climatic changes (Turchetto-Zolet, Pinheiro, Salgueiro, & Palma-Silva, 2013), the geological history and the eventual orographic contributions cannot be neglected in future phylogeographic studies addressing such taxa. In addition, the assumption that the Caatinga landscapes offer few geological barriers to gene flow among demes or populations (Werneck et al., 2012) must be interpreted with caution, because the permeability of such barriers can differ among taxa with distinct physiological tolerances and dispersion capabilities.

Undoubtedly, the development of new analytical tools coupled with the enhancing of computational power has improved our global knowledge on the biogeographic history of several taxa. Within the tropics, however, there is a marked

asymmetry in the number of studies that expose the scarcity of phylogeographic information regarding taxa from arid or semi-arid environments, compared to those from rainforest (Turchetto-Zolet et al., 2013; Pabijan et al., 2015). In conclusion, our results refuse a simplistic interpretation of intraspecific diversification processes of taxa distributed under the CD. In this sense, we provide empirical evidences that habitat stability within the CD can predict high levels of genetic diversity, at least for low-dispersal taxa, as suggested by Werneck et al. (2011). Such historical refuges can also facilitate the demographic and spatial expansion of populations over time, especially if it is preceded by complex diversification processes, that can occur even in the presence of gene flow. Finally, habitat stability seems to have been more important for the diversification of *J. rochae* at regional scale, than the potential changes in Caatinga environments mediated by climatic cycles of the Quaternary, or geological events such as the uplift of the BOP, although these factors are fundamental to preserve the genetic diversity of this scorpion. Finally, we advocate that the biogeographical history of the Caatinga and the evolution of its biota can only be understood if the urgent need for new phylogeographic approaches are met, which would mitigate the negative effects of meaningless generalizations.

References

- Aktas, C. (2019). haplotypes: manipulating DNA sequences and estimating unambiguous haplotype network with statistical parsimony. R Package Version 1.1. <https://CRAN.R-project.org/package=haplotypes>
- Allouche, O., Tsoar, A., & Kadmon, R. (2006). Assessing the accuracy of species distribution models: prevalence, kappa and the true skill statistic (TSS): Assessing the accuracy of distribution models. *Journal of Applied Ecology*, 43(6), 1223–1232. <https://doi.org/10.1111/j.1365-2664.2006.01214.x>
- Álvarez-presas, M., Carbayo, F., Rozas, J., & Riutort, M. (2011). Land planarians (Platyhelminthes) as a model organism for fine-scale phylogeographic studies: understanding patterns of biodiversity in the Brazilian Atlantic Forest hotspot. *Journal of Evolutionary Biology*, 24, 887–896. <https://doi.org/10.1111/j.1420-9101.2010.02220.x>
- Archer, F. I., Adams, P. E., & Schneiders, B. B. (2016). strataG: An R package for manipulating, summarizing and analysing population genetic data. *Molecular Ecology Resources*, 17(1), 5-11. <https://doi.org/10.1111/1755-0998.12559>

- Arruda, D. M., Schaefer, C. E. G. R., Fonseca, R. S., Solar, R. R. C., & Fernandes-Filho, E. I. (2018). Vegetation cover of Brazil in the last 21 ka: New insights into the Amazonian refugia and Pleistocenic arc hypotheses. *Global Ecology and Biogeography*, 27(1), 47–56. <https://doi.org/10.1111/geb.12646>
- Avice, J. C. (2009). Phylogeography: retrospect and prospect. *Journal of Biogeography*, 36(1), 3–15. <https://doi.org/10.1111/j.1365-2699.2008.02032.x>
- Baele, G., Lemey, P., Bedford, T., Rambaut, A., Suchard, M. A., & Alekseyenko, A. V. (2012). Improving the accuracy of demographic and molecular clock model comparison while accommodating phylogenetic uncertainty. *Molecular Biology and Evolution*, 29(9), 2157–2167. <https://doi.org/10.1093/molbev/mss084>
- Baele, G., Li, W. L. S., Drummond, A. J., Suchard, M. A., & Lemey, P. (2012). Accurate model selection of relaxed molecular clocks in Bayesian phylogenetics. *Molecular Biology and Evolution*, 30(2), 239–243. <https://doi.org/10.1093/molbev/mss243>
- Bartoletti, L. F. M., Peres, E. A., Fontes, F. von H. M., da Silva, M. J., & Solferini, V. N. (2018). Phylogeography of the widespread spider *Nephila clavipes* (Araneae: Araneidae) in South America indicates geologically and climatically driven lineage diversification. *Journal of Biogeography*, 45(6), 1246–1260. <https://doi.org/10.1111/jbi.13217>
- Bartoletti, L. F. M., Peres, E. A., Sobral-Souza, T., Fontes, F. von H. M., Silva, M. J. da, & Solferini, V. N. (2017). Phylogeography of the dry vegetation endemic species *Nephila sexpunctata* (Araneae: Araneidae) suggests recent expansion of the Neotropical Dry Diagonal. *Journal of Biogeography*, 44(9), 2007–2020. <https://doi.org/10.1111/jbi.12998>
- Batalha-Filho, H., Fjeldså, J., Fabre, P. H., & Miyaki, C. Y. (2013). Connections between the Atlantic and the Amazonian forest avifaunas represent distinct historical events. *Journal of Ornithology*, 154(1), 41–50. <https://doi.org/10.1007/s10336-012-0866-7>
- Beerli, P., & Palczewski, M. (2010). Unified framework to evaluate panmixia and migration direction among multiple sampling locations. *Genetics*, 185(1), 313–326. <https://doi.org/10.1534/genetics.109.112532>
- Binks, R. M., Gibson, N., Ottewell, K. M., Macdonald, B., & Byrne, M. (2019). Predicting contemporary range-wide genomic variation using climatic, phylogeographic and morphological knowledge in an ancient, unglaciated

- landscape. *Journal of Biogeography*, 46(3), 503–514.
<https://doi.org/10.1111/jbi.13522>
- Bond, M. O., Anderson, B. J., Henare, T. H. A., & Wehi, P. M. (2019). Effects of climatically shifting species distributions on biocultural relationships. *People and Nature*, 1, 87–102.
- Boria, R. A., Olson, L. E., Goodman, S. M., & Anderson, R. P. (2014). Spatial filtering to reduce sampling bias can improve the performance of ecological niche models. *Ecological Modelling*, 275, 73–77.
<https://doi.org/10.1016/j.ecolmodel.2013.12.012>
- Bryson, R. W., Savary, W. E., & Prendini, L. (2013). Biogeography of scorpions in the *Pseudouroctonus minimus* complex (Vaejovidae) from south-western North America: implications of ecological specialization for pre-Quaternary diversification. *Journal of Biogeography*, 40, 1850–1860.
<https://doi.org/10.1111/jbi.12134>
- Bryson, R. W., Savary, W. E., Zellmer, A. J., Bury, R. B., & McCormack, J. E. (2016). Genomic data reveal ancient microendemism in forest scorpions across the California Floristic Province. *Molecular Ecology*, 25(15), 3731–3751.
<https://doi.org/10.1111/mec.13707>
- Camps, G. A., Martínez-Meyer, E., Verga, A. R., Sérsic, A. N., & Cosacov, A. (2018). Genetic and climatic approaches reveal effects of Pleistocene refugia and climatic stability in an old giant of the Neotropical Dry Forest. *Biological Journal of the Linnean Society*, 125(2), 401–420.
<https://doi.org/10.1093/biolinnean/bly115>
- Carnaval, A. C., Hickerson, M. J., Haddad, C. F. B., Rodrigues, M. T., & Moritz, C. (2009). Stability predicts genetic diversity in the Brazilian Atlantic Forest Hotspot. *Science*, 323(5915), 785–789.
<https://doi.org/10.1126/science.1166955>
- Carnaval, A. C., & Moritz, C. (2008). Historical climate modelling predicts patterns of current biodiversity in the Brazilian Atlantic Forest. *Journal of Biogeography*, 35(7), 1187–1201.
- Carnaval, A. C., Waltari, E., Rodrigues, M. T., Rosauer, D., VanDerWal, J., Damasceno, R., Prates, I., Strangas, M., Spanos, Z., Rivera, D., Pie, M. R., Firkowski, C. R., Bornschein, M. R., Ribeiro, L. F., & Moritz, C. (2014). Prediction of phylogeographic endemism in an environmentally complex

- biome. *Proceedings of the Royal Society B: Biological Sciences*, 281(1792), 20141461. <https://doi.org/10.1098/rspb.2014.1461>
- Casquet, J., Thebaud, C., & Gillespie, R. G. (2012). Chelex without boiling, a rapid and easy technique to obtain stable amplifiable DNA from small amounts of ethanol-stored spiders. *Molecular Ecology Resources*, 12(1), 136–141. <https://doi.org/10.1111/j.1755-0998.2011.03073.x>
- Castresana, J. (2000). Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution*, 17(4), 540–552. <https://doi.org/10.1093/oxfordjournals.molbev.a026334>
- Cavalcanti, L. M. P., Paiva, L. V. de, & França, L. F. (2016). Effects of rainfall on bird reproduction in a semi-arid Neotropical region. *Zoologia (Curitiba)*, 33(6), e20160018. <https://doi.org/10.1590/s1984-4689zool-20160018>
- Ceccarelli, F. S., Ojanguren-Affilastro, A. A., Ramírez, M. J., Ochoa, J. A., Mattoni, C. I., & Prendini, L. (2016). Andean uplift drives diversification of the bothriurid scorpion genus *Brachistosternus*. *Journal of Biogeography*, 43(10), 1942–1954. <https://doi.org/10.1111/jbi.12760>
- Ceccarelli, F. S., Pizarro-Araya, J., & Ojanguren-Affilastro, A. A. (2017). Phylogeography and population structure of two *Brachistosternus* species (Scorpiones: Bothriuridae) from the Chilean coastal desert - the perils of coastal living. *Biological Journal of the Linnean Society*, 120, 75–89. <https://doi.org/10.1111/bij.12877>
- Corander, J., Sirén, J., & Arjas, E. (2008). Bayesian spatial modeling of genetic population structure. *Computational Statistics*, 23(1), 111–129. <https://doi.org/10.1007/s00180-007-0072-x>
- Corander, J., & Tang, J. (2007). Bayesian analysis of population structure based on linked molecular information. *Mathematical Biosciences*, 205(1), 19–31. <https://doi.org/10.1016/j.mbs.2006.09.015>
- Costa, G. C., Hampe, A., Ledru, M.-P., Martinez, P. A., Mazzochini, G. G., Shepard, D. B., Werneck, F. P., Moritz, C., & Carnaval, A. C. (2017). Biome stability in South America over the last 30 kyr: Inferences from long-term vegetation dynamics and habitat modelling. *Global Ecology and Biogeography*, 27(3), 285–297. <https://doi.org/10.1111/geb.12694>
- Costa, W. J. E. M., Amorim, P. F., & Bragança, P. H. N. (2013). Species limits and phylogenetic relationships of red-finned cryptic species of the seasonal killifish

- genus *Hypsolebias* from the Brazilian semi-arid Caatinga (Teleostei: Cyprinodontiformes: Rivulidae). *Journal of Zoological Systematics and Evolutionary Research*, 52(1), 52–58. <https://doi.org/10.1111/jzs.12041>
- Costa, W. J. E. M., Amorim, P. F., & Mattos, J. L. O. (2018). Synchronic historical patterns of species diversification in seasonal aplocheiloid killifishes of the semi-arid Brazilian Caatinga. *Plos One*, 13(2), e0193021. <https://doi.org/10.1371/journal.pone.0193021>
- Dray, S., & Dufour, A. (2007). The ade4 Package: implementing the Duality Diagram for ecologists. *Journal of Statistical Software*, 22(4), 1–20. <https://doi.org/10.18637/jss.v022.i04>
- Elith, J., H. Graham, C., P. Anderson, R., Dudík, M., Ferrier, S., Guisan, A., J. Hijmans, R., Huettmann, F., R. Leathwick, J., Lehmann, A., Li, J., G. Lohmann, L., A. Loiselle, B., Manion, G., Moritz, C., Nakamura, M., Nakazawa, Y., McC. M. Overton, J., Townsend Peterson, A., ... E. Zimmermann, N. (2006). Novel methods improve prediction of species' distributions from occurrence data. *Ecography*, 29(2), 129–151. <https://doi.org/10.1111/j.2006.0906-7590.04596.x>
- Endler, J. A. (1973). Gene Flow and Population Differentiation: Studies of clines suggest that differentiation along environmental gradients may be independent of gene flow. *Science*, 179(4070), 243–250. <https://doi.org/10.1126/science.179.4070.243>
- Esposito, L. A., Yamaguti, H. Y., Pinto-Da-Rocha, R., & Prendini, L. (2018). Plucking with the plectrum: phylogeny of the New World buthid scorpion subfamily Centruroidinae Kraus, 1955 (Scorpiones: Buthidae) reveals evolution of three pecten-sternite stridulation organs. *Arthropod Systematics & Phylogeny*, 76, 87–122.
- Esposito, L. A., & Prendini, L. (2019). Island ancestors and New World biogeography: a case study from the scorpions (Buthidae: Centruroidinae). *Scientific Reports*, 9(1), 3500. <https://doi.org/10.1038/s41598-018-33754-8>
- Esposito, L. A., Yamaguti, H. Y., Souza, C. A., Pinto-Da-Rocha, R., & Prendini, L. (2017). Systematic revision of the Neotropical club-tailed scorpions, *Physoctonus*, *Rhopalurus*, and *Troglorhopalurus*, revalidation of *Heteroctenus*, and descriptions of two new genera and three new species

- (Buthidae: Rhopalurusinae). *Bulletin of the American Museum of Natural History*, 415, 1–136. <https://doi.org/10.1206/0003-0090-415.1.1>
- Excoffier, L., & Lischer, H. E. L. (2010). Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*, 10(3), 564–567. <https://doi.org/10.1111/j.1755-0998.2010.02847.x>
- Excoffier, L., Smouse, P. E., & Quattro, J. M. (1992). Analysis of Molecular Variance Inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics*, 131, 479–491.
- Faye, A., Deblauwe, V., Mariac, C., Richard, D., Sonké, B., Vigouroux, Y., & Couvreur, T. L. P. (2016). Phylogeography of the genus *Podococcus* (Palmae/Arecaceae) in Central African rain forests: Climate stability predicts unique genetic diversity. *Molecular Phylogenetics and Evolution*, 105, 126–138. <https://doi.org/10.1016/j.ympev.2016.08.005>
- Finn, D. S., Bogan, M. T., & Lytle, D. A. (2009). Demographic stability metrics for conservation prioritization of isolated populations. *Conservation Biology*, 23(5), 1185–1194. <https://doi.org/10.1111/j.1523-1739.2009.01226.x>
- Fithian, W., & Hastie, T. (2013). Finite-sample equivalence in statistical models for presence-only data. *The Annals of Applied Statistics*, 7(4), 1917–1939. <https://doi.org/10.1214/13-AOAS667>
- Foerster, S. I. A., DeSouza, A. M., & Lira, A. F. A. (2019). Macroecological approach for scorpions (Arachnida, Scorpiones): β -diversity in Brazilian montane forests. *Canadian Journal of Zoology*, 97(10), 914–921. <https://doi.org/10.1139/cjz-2019-0008>
- Foord, S. H., Gelebe, V., & Prendini, L. (2015). Effects of aspect and altitude on scorpion diversity along an environmental gradient in the Soutpansberg, South Africa. *Journal of Arid Environments*, 113, 114–120. <https://doi.org/10.1016/j.jaridenv.2014.10.006>
- Fourcade, Y., Besnard, A. G., & Secondi, J. (2017). Paintings predict the distribution of species, or the challenge of selecting environmental predictors and evaluation statistics. *Global Ecology and Biogeography*, 1–12. <https://doi.org/10.1111/geb.12684>
- Gantenbein, B., Fet, V., Gantenbein-Ritter, I. A., & Balloux, F. (2005). Evidence for recombination in scorpion mitochondrial DNA (Scorpiones: Buthidae).

- Proceedings of the Royal Society B: Biological Sciences*, 272(1564), 697–704.
<https://doi.org/10.1098/rspb.2004.3017>
- Gent, P. R., Danabasoglu, G., Donner, L. J., Holland, M. M., Hunke, E. C., Jayne, S. R., Lawrence, D. M., Neale, R. B., Rasch, P. J., Vertenstein, M., Worley, P. H., Yang, Z. L., & Zhang, M. (2011). The Community Climate System Model Version 4. *Journal of Climate*, 24(19), 4973–4991.
<https://doi.org/10.1175/2011JCLI4083.1>
- Gernhard, T. (2008). The conditioned reconstructed process. *Journal of Theoretical Biology*, 253(4), 769–778. <https://doi.org/10.1016/j.jtbi.2008.04.005>
- Giorgetta, M. A., Jungclaus, J., Reick, C. H., Legutke, S., Bader, J., Böttinger, M., Brovkin, V., Crueger, T., Esch, M., Fieg, K., Glushak, K., Gayler, V., Haak, H., Hollweg, H. D., Ilyina, T., Kinne, S., Kornblueh, L., Matei, D., Mauritsen, T., ... Stevens, B. (2013). Climate and carbon cycle changes from 1850 to 2100 in MPI-ESM simulations for the Coupled Model Intercomparison Project phase 5: Climate Changes in MPI-ESM. *Journal of Advances in Modeling Earth Systems*, 5(3), 572–597. <https://doi.org/10.1002/jame.20038>
- Gómez-Carballa, A., Pardo-Seco, J., Brandini, S., Achilli, A., Perego, U. A., Coble, M. D., Diegoli, T. M., Álvarez-Iglesias, V., Martínón-Torres, F., Olivieri, A., Torroni, A., & Salas, A. (2018). The peopling of South America and the trans-Andean gene flow of the first settlers. *Genome Research*, 28(6), 767–779.
<https://doi.org/10.1101/gr.234674.118>
- Gonçalves, G. S. R., Cerqueira, P. V., Brasil, L. S., & Santos, M. P. D. (2017). The role of climate and environmental variables in structuring bird assemblages in the Seasonally Dry Tropical Forests (SDTFs). *Plos One*, 12(4), e0176066.
<https://doi.org/10.1371/journal.pone.0176066>
- Graham, M. R., Jaeger, J. R., Prendini, L., & Riddle, B. R. (2013). Phylogeography of the Arizona hairy scorpion (*Hadrurus arizonensis*) supports a model of biotic assembly in the Mojave Desert and adds a new Pleistocene refugium. *Journal of Biogeography*, 40(7), 1298–1312. <https://doi.org/10.1111/jbi.12079>
- Graham, M. R., Myers, E. A., Kaiser, R. C., & Fet, V. (2019). Cryptic species and co-diversification in sand scorpions from the Karakum and Kyzylkum deserts of Central Asia. *Zoologica Scripta*, 48(6), 801–812.
<https://doi.org/10.1111/zsc.12381>

- Graham, M. R., Wood, D. A., Henault, J. A., Valois, Z. J., & Cushing, P. E. (2017). Ancient lakes, Pleistocene climates and river avulsions structure the phylogeography of a large but little-known rock scorpion from the Mojave and Sonoran deserts. *Biological Journal of the Linnean Society*, 122(1), 133–146. <https://doi.org/10.1093/biolinnean/blx058>
- Grant, W. S. (2015). Problems and cautions with sequence mismatch analysis and Bayesian Skyline Plots to Infer historical demography. *Journal of Heredity*, 106(4), 333–346. <https://doi.org/10.1093/jhered/esv020>
- Gugger, P. F., Ikegami, M., & Sork, V. L. (2013). Influence of late Quaternary climate change on present patterns of genetic variation in valley oak, *Quercus lobata* Née. *Molecular Ecology*, 22(13), 3598–3612. <https://doi.org/10.1111/mec.12317>
- Guisan, A., Thuiller, W., & Zimmermann, N. E. (2017). *Habitat Suitability and Distribution Models with Applications in R*. Cambridge University Press.
- Gutiérrez-Rodríguez, J., Barbosa, A. M., & Martínez-Solano, Í. (2017). Present and past climatic effects on the current distribution and genetic diversity of the Iberian spadefoot toad (*Pelobates cultripes*): an integrative approach. *Journal of Biogeography*, 44(2), 245–258. <https://doi.org/10.1111/jbi.12791>
- Harms, D., Roberts, J. D., & Harvey, M. S. (2019). Climate variability impacts on diversification processes in a biodiversity hotspot: a phylogeography of ancient pseudoscorpions in south-western Australia. *Zoological Journal of the Linnean Society*, 186(4), 934–949. <https://doi.org/10.1093/zoolinnean/zlz010>
- Hijmans, R. J., Cameron, S. E., Parra, J. L., Jones, P. G., & Jarvis, A. (2005). Very high-resolution interpolated climate surfaces for global land areas. *International Journal of Climatology*, 25(15), 1965–1978. <https://doi.org/10.1002/joc.1276>
- Hijmans, R. J., Phillips, S., Leathwick, J., & Elith, J. (2017). dismo: Species Distribution Modeling. R Package Version 1.1-4. <https://CRAN.R-project.org/package=dismo>
- Holsinger, K. E., & Weir, B. S. (2009). Genetics in geographically structured populations: defining, estimating and interpreting F_{ST} . *Nature Reviews Genetics*, 10(9), 639–650. <https://doi.org/10.1038/nrg2611>
- Husemann, M., Schmitt, T., Stathi, I., & Habel, J. C. (2012). Evolution and Radiation in the Scorpion *Buthus elmoutaouakili* Lourenço and Qi 2006 (Scorpiones:

- Buthidae) at the Foothills of the Atlas Mountains (North Africa). *Journal of Heredity*, 103(2), 221–229. <https://doi.org/10.1093/jhered/esr130>
- Katoh, K., & Standley, D. M. (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution*, 30(4), 772–780. <https://doi.org/10.1093/molbev/mst010>
- Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution*, 35(6), 1547–1549. <https://doi.org/10.1093/molbev/msy096>
- Lanna, F. M., Gehara, M., Werneck, F. P., Fonseca, E. M., Colli, G. R., Sites, J. W., Rodrigues, M. T., & Garda, A. A. (2019). Dwarf geckos and giant rivers: the role of the São Francisco River in the evolution of *Lygodactylus klugei* (Squamata: Gekkonidae) in the semi-arid Caatinga of north-eastern Brazil. *Biological Journal of the Linnean Society*, 129(1), 88–98. <https://doi.org/10.1093/biolinnean/blz170>
- Leigh, J. W., & Bryant, D. (2015). POPART: full-feature software for haplotype network construction. *Methods in Ecology and Evolution*, 6(9), 1110–1116. <https://doi.org/10.1111/2041-210X.12410>
- Lima-Rezende, C. A., Rocha, A. V., Júnior, A. F. C., Martins, É. de S., Vasconcelos, V., & Caparroz, R. (2019). Late Pleistocene climatic changes promoted demographic expansion and population reconnection of a Neotropical savanna-adapted bird, *Neothraupis fasciata* (Aves: Thraupidae). *Plos One*, 14(3), e0212876. <https://doi.org/10.1371/journal.pone.0212876>
- Lira, A. F. A., DeSouza, A. M., & Albuquerque, C. M. R. (2018). Environmental variation and seasonal changes as determinants of the spatial distribution of scorpions (Arachnida: Scorpiones) in Neotropical forests. *Canadian Journal of Zoology*, 96(9), 963–972. <https://doi.org/10.1139/cjz-2017-0251>
- Lira, A. F. A., Pordeus, L. M., & Albuquerque, C. M. R. (2017). A new species of *Ananteris* (Scorpiones: Buthidae) from Caatinga biome, Brazil. *Acta Arachnologica*, 66(1), 9–15.
- Liu, C., Berry, P. M., Dawson, T. P., & Pearson, R. G. (2005). Selecting thresholds of occurrence in the prediction of species distributions. *Ecography*, 28(3), 385–393. <https://doi.org/10.1111/j.0906-7590.2005.03957.x>

- Liu, C., Newell, G., & White, M. (2016). On the selection of thresholds for predicting species occurrence with presence-only data. *Ecology and Evolution*, 6(1), 337–348. <https://doi.org/10.1002/ece3.1878>
- Liu, C., White, M., & Newell, G. (2011). Measuring and comparing the accuracy of species distribution models with presence-absence data. *Ecography*, 34(2), 232–243. <https://doi.org/10.1111/j.1600-0587.2010.06354.x>
- Lourenço, W. R. (2008). The geographic pattern of distribution of the genus *Rhopalurus* Thorell, 1876 in the Guayana-Amazon region (Scorpiones: Buthidae). *Euscorpius*, 2008(73), 1–14. <https://doi.org/10.18590/euscorpius.2008.vol2008.iss73.1>
- Machado, A. F., Nunes, M. S., Silva, C. R., Santos Jr, M. A., Farias, I. P., da Silva, M. N. F., & Anciães, M. (2019). Integrating phylogeography and ecological niche modelling to test diversification hypotheses using a Neotropical rodent. *Evolutionary Ecology*, 33(1), 111–148. <https://doi.org/10.1007/s10682-019-09968-1>
- Menezes, R., Sampaio, E., Giongo, V., & Pérez-Marin, A. (2012). Biogeochemical cycling in terrestrial ecosystems of the Caatinga Biome. *Brazilian Journal of Biology*, 72, 643–653. <https://doi.org/10.1590/S1519-69842012000400004>
- Merow, C., Allen, J. M., Aiello-Lammens, M., & Silander, J. A. (2016). Improving niche and range estimates with Maxent and point process models by integrating spatially explicit information: Minxent. *Global Ecology and Biogeography*, 25(8), 1022–1036. <https://doi.org/10.1111/geb.12453>
- Merow, C., Smith, M. J., & Silander, J. A. (2013). A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. *Ecography*, 36(10), 1058–1069. <https://doi.org/10.1111/j.1600-0587.2013.07872.x>
- MMA Ministério do Meio Ambiente. (2020). Mapas de cobertura vegetal dos biomas Brasileiros. Ministério Do Meio Ambiente Do Brasil (MMA). <http://mapas.mma.gov.br/mapas/aplic/probio/datadownload.htm?/caatinga/documentos/index.html>
- Morais-Neto, J. M., Hegarty, K. A., Karner, G. D., & Alkmim, F. F. (2009). Timing and mechanisms for the generation and modification of the anomalous topography of the Borborema Province, northeastern Brazil. *Marine and*

- Petroleum Geology*, 26(7), 1070–1086.
<https://doi.org/10.1016/j.marpetgeo.2008.07.002>
- Moura, A. C. A., & Lee, P. C. (2004). Capuchin Stone Tool Use in Caatinga Dry Forest. *Science*, 306(5703), 1909–1909.
<https://doi.org/10.1126/science.1102558>
- Muscarella, R., Galante, P. J., Soley-Guardia, M., Boria, R. A., Kass, J. M., Uriarte, M., & Anderson, R. P. (2014). ENMeval: An R package for conducting spatially independent evaluations and estimating optimal model complexity for MAXENT ecological niche models. *Methods in Ecology and Evolution*, 5, 1198–1205.
- Naimi, B., Hamm, N. A. S., Groen, T. A., Skidmore, A. K., & Toxopeus, A. G. (2014). Where is positional uncertainty a problem for species distribution modelling? *Ecography*, 37(2), 191–203. <https://doi.org/10.1111/j.1600-0587.2013.00205.x>
- Noguerales, V., Cordero, P. J., & Ortego, J. (2018). Inferring the demographic history of an oligophagous grasshopper: Effects of climatic niche stability and host-plant distribution. *Molecular Phylogenetics and Evolution*, 118, 343–356.
<https://doi.org/10.1016/j.ympev.2017.10.012>
- Nosil, P. (2008). Speciation with gene flow could be common: news and views. *Molecular Ecology*, 17(9), 2103–2106. <https://doi.org/10.1111/j.1365-294X.2008.03715.x>
- Novaes, R. M. L., Ribeiro, R. A., Lemos-Filho, J. P., & Lovato, M. B. (2013). Concordance between Phylogeographical and Biogeographical Patterns in the Brazilian Cerrado: Diversification of the Endemic Tree *Dalbergia miscolobium* (Fabaceae). *Plos One*, 8(12), e82198.
<https://doi.org/10.1371/journal.pone.0082198>
- Oberski, J. T., Sharma, P. P., Jay, K. R., Coblens, M. J., Lemon, K. A., Johnson, J. E., & Boyer, S. L. (2018). A dated molecular phylogeny of mite harvestmen (Arachnida: Opiliones: Cyphophthalmi) elucidates ancient diversification dynamics in the Australian Wet Tropics. *Molecular Phylogenetics and Evolution*, 127, 813–822. <https://doi.org/10.1016/j.ympev.2018.06.029>
- Ojanguren-Affilastro, A. A., Adilardi, R. S., Mattoni, C. I., Ramírez, M. J., & Ceccarelli, F. S. (2017). Dated phylogenetic studies of the southernmost

- American buthids (Scorpiones; Buthidae). *Molecular Phylogenetics and Evolution*, 110, 39–49. <https://doi.org/10.1016/j.ympev.2017.02.018>
- Ojanguren-Affilastro, A. A., Mattoni, C. I., Ochoa, J. A., Ramírez, M. J., Ceccarelli, F. S., & Prendini, L. (2016). Phylogeny, species delimitation and convergence in the South American bothriurid scorpion genus *Brachistosternus* Pocock 1893: Integrating morphology, nuclear and mitochondrial DNA. *Molecular Phylogenetics and Evolution*, 94, 159–170. <https://doi.org/10.1016/j.ympev.2015.08.007>
- Oliveira, E. F., Gehara, M., São-Pedro, V. A., Chen, X., Myers, E. A., Burbrink, F. T., Mesquita, D. O., Garda, A. A., Colli, G. R., Rodrigues, M. T., Arias, F. J., Zaher, H., Santos, R. M. L., & Costa, G. C. (2015). Speciation with gene flow in whiptail lizards from a Neotropical xeric biome. *Molecular Ecology*, 24(23), 5957–5975. <https://doi.org/10.1111/mec.13433>
- Oliveira, E. F., Martinez, P. A., São-Pedro, V. A., Gehara, M., Burbrink, F. T., Mesquita, D. O., Garda, A. A., Colli, G. R., & Costa, G. C. (2018). Climatic suitability, isolation by distance and river resistance explain genetic variation in a Brazilian whiptail lizard. *Heredity*, 120(3), 251–265. <https://doi.org/10.1038/s41437-017-0017-2>
- Oliveira, R. G., & de Medeiros, W. E. (2012). Evidences of buried loads in the base of the crust of Borborema Plateau (NE Brazil) from Bouguer admittance estimates. *Journal of South American Earth Sciences*, 37, 60–76.
- Olson, D. M., Dinerstein, E., Wikramanayake, E. D., Burgess, N. D., Powel, G. V., Underwood, E. C., D'amico, J. A., Itoua, I., Strand, H. E., Morrison, J. C., Loucks, C. J., Allunt, T. F., Ricketts, T. H., Kura, Y., Lamoreux, J. F., Wettengel, E. E., Hedao, P., & Kesseem, K. R. (2001). Terrestrial Ecoregions of the World: A New Map of Life on EarthA new global map of terrestrial ecoregions provides an innovative tool for conserving biodiversity. *BioScience*, 51, 933–938.
- Pabijan, M., Brown, J. L., Chan, L. M., Rakotondravony, H. A., Raselimanana, A. P., Yoder, A. D., Glaw, F., & Vences, M. (2015). Phylogeography of the arid-adapted Malagasy bullfrog, *Laliostoma labrosum*, influenced by past connectivity and habitat stability. *Molecular Phylogenetics and Evolution*, 92, 11–24. <https://doi.org/10.1016/j.ympev.2015.05.018>

- Pacioni, C., Hunt, H., Allentoft, M. E., Vaughan, T. G., Wayne, A. F., Baynes, A., Haouchar, D., Dortch, J., & Bunce, M. (2015). Genetic diversity loss in a biodiversity hotspot: ancient DNA quantifies genetic decline and former connectivity in a critically endangered marsupial. *Molecular Ecology*, 24(23), 5813–5828. <https://doi.org/10.1111/mec.13430>
- Paradis, E. (2010). pegas: an R package for population genetics with an integrated-modular approach. *Bioinformatics*, 26, 419–420.
- Paradis, E., & Schliep, K. (2018). ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics*, 35, 526–528.
- Passoni, J. C., Benozzati, M. L., & Rodrigues, M. T. (2008). Phylogeny, species limits, and biogeography of the Brazilian lizards of the genus *Eurolophosaurus* (Squamata: Tropiduridae) as inferred from mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution*, 46(2), 403–414. <https://doi.org/10.1016/j.ympev.2007.10.022>
- Paz, A., Spanos, Z., Brown, J. L., Lyra, M., Haddad, C., Rodrigues, M., & Carnaval, A. (2019). Phylogeography of Atlantic Forest glassfrogs (*Vitreorana*): when geography, climate dynamics and rivers matter. *Heredity*, 122(5), 545–557. <https://doi.org/10.1038/s41437-018-0155-1>
- Pennington, R. T., Lavin, M., & Oliveira-Filho, A. (2009). Woody Plant Diversity, Evolution, and Ecology in the Tropics: Perspectives from Seasonally Dry Tropical Forests. *Annual Review of Ecology, Evolution, and Systematics*, 40(1), 437–457. <https://doi.org/10.1146/annurev.ecolsys.110308.120327>
- Pennington, R. T., Prado, D. E., & Pendry, C. A. (2000). Neotropical seasonally dry forests and Quaternary vegetation changes. *Journal of Biogeography*, 27(2), 261–273. <https://doi.org/10.1046/j.1365-2699.2000.00397.x>
- Peres, E. A., Benedetti, A. R., Hiruma, S. T., Sobral-Souza, T., & Pinto-da-Rocha, R. (2019). Phylogeography of Sodreaninae harvestmen (Arachnida: Opiliones: Gonyleptidae): Insights into the biogeography of the southern Brazilian Atlantic Forest. *Molecular Phylogenetics and Evolution*, 138, 1–16. <https://doi.org/10.1016/j.ympev.2019.05.028>
- Peres, E. A., Sobral-Souza, T., Perez, M. F., Bonatelli, I. A. S., Silva, D. P., Silva, M. J., & Solferini, V. N. (2015). Pleistocene Niche Stability and Lineage Diversification in the Subtropical Spider *Araneus omnicolor* (Araneidae). *Plos One*, 10(4), e0121543. <https://doi.org/10.1371/journal.pone.0121543>

- Pfingstl, T., Wagner, M., Hiruta, S. F., Koblmüller, S., Hagino, W., & Shimano, S. (2019). Phylogeographic patterns of intertidal arthropods (Acari, Oribatida) from southern Japanese islands reflect paleoclimatic events. *Scientific Reports*, 9(1), 19042. <https://doi.org/10.1038/s41598-019-55270-z>
- Phillips, S. J., Anderson, R. P., Dudík, M., Schapire, R. E., & Blair, M. E. (2017). Opening the black box: an open-source release of Maxent. *Ecography*, 40(7), 887–893. <https://doi.org/10.1111/ecog.03049>
- Phillips, S. J., Anderson, R. P., & Schapire, R. E. (2006). Maximum entropy modeling of species geographic distributions. *Ecological Modelling*, 190(3–4), 231–259. <https://doi.org/10.1016/j.ecolmodel.2005.03.026>
- Polis, G. A., McReynolds, C. N., & Ford, R. G. (1985). Home range geometry of the desert scorpion *Paruroctonus mesaensis*. *Oecologia*, 67(2), 273–277.
- Porto T. J., Carvalho, L. S., de Souza, C. A. R., Oliveira, U., & Brescovit, A. D. (2011). Escorpiões da Caatinga: conhecimentos atuais e desafios. In F. Bravo & A. Calor (Eds.), *Artrópodes do Semiárido Biodiversidade e Conservação* (pp. 33–46). Printmídia.
- Powell, A. F. L. A., Barker, F. K., & Lanyon, S. M. (2013). Empirical evaluation of partitioning schemes for phylogenetic analyses of mitogenomic data: an avian case study. *Molecular Phylogenetics and Evolution*, 66(1), 69–79. <https://doi.org/10.1016/j.ympev.2012.09.006>
- Prado, C. P. A., Haddad, C. F. B., & Zamudio, K. R. (2012). Cryptic lineages and Pleistocene population expansion in a Brazilian Cerrado frog: Brazilian Cerrado frog phylogeography. *Molecular Ecology*, 21(4), 921–941. <https://doi.org/10.1111/j.1365-294X.2011.05409.x>
- QGIS Development Team. (2020). QGIS Geographic Information System. Open Source Geospatial Foundation Project. <http://qgis.osgeo.org>
- Queiroz, L. P., Cardoso, D., Fernandes, M. F., & Moro, M. F. (2018). Diversity and evolution of flowering plants of the Caatinga Domain. In J. M. C. da Silva, I. R. Leal, & M. Tabarelli (Eds.), *Caatinga The Largest Tropical Dry Forest in South America* (pp. 23–65). Springer Nature.
- Radosavljevic, A., & Anderson, R. P. (2014). Making better MAXENT models of species distributions: complexity, overfitting and evaluation. *Journal of Biogeography*, 41, 629–643.

- R Core Team. (2019). R: A language and environment for statistical computing. R Foundation for Statistical Computing. <https://www.R-project.org/>
- Rambaut, A., Drummond, A. J., Xie, D., Baele, G., & Suchard, M. A. (2018). Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. *Systematic Biology*, 67(5), 901–904. <https://doi.org/10.1093/sysbio/syy032>
- Ramos-Onsins, S. E., & Rozas, J. (2002). Statistical Properties of New Neutrality Tests Against Population Growth. *Molecular Biology and Evolution*, 19, 2092–2100.
- Rodal, M. J. N., & Nascimento, L. M. (2006). The arboreal component of a dry forest in Northeastern Brazil. *Brazilian Journal of Biology*, 66(2a), 479–491. <https://doi.org/10.1590/S1519-69842006000300014>
- Rodal, M. J. N., Barbosa, M. R. V., & Thomas, W. W. (2008). Do the seasonal forests in northeastern Brazil represent a single floristic unit? *Brazilian Journal of Biology*, 68(3), 467–475. <https://doi.org/10.1590/S1519-69842008000300003>
- Rodrigues, J. F. M., & Silva, J. R. F. (2014). How *Phrynosops tuberosus* (Testudines: Chelidae) reproduce in the Brazilian Caatinga? *North-Western Journal of Zoology*, 10, 143–148.
- Salimon, C., & Anderson, L. (2018). How strong is the relationship between rainfall variability and caatinga productivity? A case study under a changing climate. *Anais Da Academia Brasileira de Ciências*, 90, 2121–2127. <https://doi.org/10.1590/0001-3765201720170143>
- Santos, J. C., Leal, I. R., Almeida-Cortez, J. S., Fernandes, G. W., & Tabarelli, M. (2011). Caatinga: the scientific negligence experienced by a Dry Tropical Forest. *Tropical Conservation Science*, 4(3), 276–286. <https://doi.org/10.1177/194008291100400306>
- Santos-Da-Silva, A. D. P., Carvalho, L. S., & Brescovit, A. D. (2017). Two new species of *Bothriurus* Peters, 1861 (Scorpiones, Bothriuridae) from Northeastern Brazil. *Zootaxa*, 4258(3), 238. <https://doi.org/10.11646/zootaxa.4258.3.2>
- Schliep, K. P. (2011). phangorn: phylogenetic analysis in R. *Bioinformatics*, 27(4), 592–593. <https://doi.org/10.1093/bioinformatics/btq706>
- Silva, A. C. A., Bragg, J. G., Potter, S., Fernandes, C., Coelho, M. M., & Moritz, C. (2017). Tropical specialist vs. climate generalist: diversification and

- demographic history of sister species of *Carlia* skinks from northwestern Australia. *Molecular Ecology*, 26, 4045–4058.
- Silva, J. M. C., Barbosa, L. C. F., Leal, I. R., & Tabarelli, M. (2018). The Caatinga: Understanding the Challenges. In J. M. C. da Silva, I. R. Leal, & M. Tabarelli (Eds.), *Caatinga The Largest Tropical Dry Forest in South America* (pp. 3–23). Springer Nature.
- Sobral-Souza, T., Lima-Ribeiro, M. S., & Solferini, V. N. (2015). Biogeography of Neotropical Rainforests: past connections between Amazon and Atlantic Forest detected by ecological niche modeling. *Evolutionary Ecology*, 29(5), 643–655. <https://doi.org/10.1007/s10682-015-9780-9>
- Štundlová, J., Šmíd, J., Nguyen, P., & Šťáhlavský, F. (2019). Cryptic diversity and dynamic chromosome evolution in Alpine scorpions (Euscorpiidae: *Euscorpius*). *Molecular Phylogenetics and Evolution*, 134, 152–163. <https://doi.org/10.1016/j.ympev.2019.02.002>
- Suchard, M. A., Lemey, P., Baele, G., Ayres, D. L., Drummond, A. J., & Rambaut, A. (2018). Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. *Virus Evolution*, 4(1). <https://doi.org/10.1093/ve/vey016>
- Tajima, F. (1989). Statistical Method for Testing the Neutral Mutation Hypothesis by DNA Polymorphism. *Genetics*, 123, 585–595.
- Teruel, R., & Tietz, A. K. (2008). The true identity of *Rhopalurus pinto* Mello-Leitão, 1932, with notes on the status and distribution of *Rhopalurus crassicauda* Caporiacco, 1947 (Scorpiones: Buthidae). *Euscorpius*, 2008(70), 1–14. <https://doi.org/10.18590/euscorpius.2008.vol2008.iss70.1>
- Thomé, M. T. C., & Carstens, B. C. (2016). Phylogeographic model selection leads to insight into the evolutionary history of four-eyed frogs. *Proceedings of the National Academy of Sciences*, 113(29), 8010–8017. <https://doi.org/10.1073/pnas.1601064113>
- Turchetto-Zolet, A. C., Pinheiro, F., Salgueiro, F., & Palma-Silva, C. (2013). Phylogeographical patterns shed light on evolutionary process in South America. *Molecular Ecology*, 22(5), 1193–1213. <https://doi.org/10.1111/mec.12164>
- Velloso, A. L., Sampaio, E. V. S. B., & Pareyn, F. G. C. (2002). *Ecorregiões propostas para o Bioma Caatinga*. Recife: The Nature Conservancy do Brasil.

- Vieira, W. L. S., Santana, G. G., & Arzabe, C. (2009). Diversity of reproductive modes in anurans communities in the Caatinga (dryland) of northeastern Brazil. *Biodiversity and Conservation*, 18(1), 55–66. <https://doi.org/10.1007/s10531-008-9434-0>
- Vink, C. J., Thomas, S. M., Paquin, P., Hayashi, C. Y., & Hedin, M. (2005). The effects of preservatives and temperatures on arachnid DNA. *Invertebrate Systematics*, 19(2), 99. <https://doi.org/10.1071/IS04039>
- Wan, J. Z., Wang, C. J., & Yu, F. H. (2017). Spatial conservation prioritization for dominant tree species of Chinese forest communities under climate change. *Climatic Change*, 144(2), 303–316. <https://doi.org/10.1007/s10584-017-2044-7>
- Wang, C. J., Wan, J. Z., & Zhang, Z. X. (2017). Expansion potential of invasive tree plants in ecoregions under climate change scenarios: an assessment of 54 species at a global scale. *Scandinavian Journal of Forest Research*, 32(8), 663–670. <https://doi.org/10.1080/02827581.2017.1283049>
- Warren, D. L., & Seifert, S. N. (2011). Ecological niche modeling in Maxent: the importance of model complexity and the performance of model selection criteria. *Ecological Applications*, 21, 335–342.
- Watanabe, S., Hajima, T., Sudo, K., Nagashima, T., Takemura, T., Okajima, H., Nozawa, T., Kawase, H., Abe, M., Yokohata, T., Ise, T., Sato, H., Kato, E., Takata, K., Emori, S., & Kawamiya, M. (2011). MIROC-ESM 2010: model description and basic results of CMIP5-20c3m experiments. *Geoscientific Model Development*, 4(4), 845–872. <https://doi.org/10.5194/gmd-4-845-2011>
- Werneck, F. P. (2011). The diversification of eastern South American open vegetation biomes: Historical biogeography and perspectives. *Quaternary Science Reviews*, 30(13–14), 1630–1648. <https://doi.org/10.1016/j.quascirev.2011.03.009>
- Werneck, F. P., Costa, G. C., Colli, G. R., Prado, D. E., & Sites Jr, J. W. (2011). Revisiting the historical distribution of Seasonally Dry Tropical Forests: new insights based on palaeodistribution modelling and palynological evidence. *Global Ecology and Biogeography*, 20(2), 272–288. <https://doi.org/10.1111/j.1466-8238.2010.00596.x>
- Werneck, F. P., Gamble, T., Colli, G. R., Rodrigues, M. T., & Sites Jr, J. W. (2012). Deep diversification and long-term persistence in the South American ‘Dry

Diagonal': integrating continent-wide phylogeography and distribution modeling of geckos. *Evolution*, 66(10), 3014–3034.

<https://doi.org/10.1111/j.1558-5646.2012.01682.x>

Werneck, F. P., Leite, R. N., Geurgas, S. R., & Rodrigues, M. T. (2015). Biogeographic history and cryptic diversity of saxicolous Tropiduridae lizards endemic to the semiarid Caatinga. *BMC Evolutionary Biology*, 15(1), 94.

<https://doi.org/10.1186/s12862-015-0368-3>

Supplementary information

Additional Supporting Information may be found in the online version of this article:

Appendix S1 Supplementary tables.

Appendix S2 Supplementary figures.

Biosketch

Stênio Ítalo A. Foerster is interested in phylogeography and macroecology of neotropical arachnids adapted to arid environments, especially those with narrower ecological plasticity and low-dispersion capabilities. All authors are interested in population genetics and evolutionary biology.

Author contributions:

SIAF, AFAL and VQB conceived the study; SIAF and AFAL performed the fieldwork; BNSP and SIAF conducted the molecular procedures; SIAF analyzed the data and wrote the initial text, that was evaluated and discussed by AFAL, VQB, BNSP, JFL and RP-R. All authors are in accordance with the final version of this manuscript.

Tables

Table 1. Genetic diversity estimations and neutrality tests output for sampling locations of *Jaguajir rochae*, in northeastern Brazil: Afogados da Ingazeira (AFO), Água Branca (AGU), Buíque (BUI), Caetés (CAE), Caruaru (CAR), Cumaru (CUM), Juazeiro do Norte (JUA), Limoeiro (LIM), Parnamirim (PAR), Serra Talhada (SER), Triunfo (TRI). Results for the eastern (EAS) and western (WES) haplogroups recovered by BAPS are also shown. N = Number of specimens; h = number of haplotypes; grh = number of geographically restricted haplotypes; Hd = haplotype diversity; π = nucleotide diversity; F_s = Fu's F_s ; D = Tajima's D; R_2 = R_2 statistic; SD = standard deviation. * $P < 0.05$; ** $P < 0.02$.

Sampling site	N	h	grh	Hd (SD)	π (SD)	F_s	D	R_2
AFO	7	7	7	1.0000 (0.00291)	0.0096 (0.0000350)	-2.3853	-0.5895	0.1134*
AGU	7	7	6	1.0000 (0.00291)	0.0053 (0.0000122)	-3.8565**	-1.4217	0.1183*
BUI	8	6	6	0.8928 (0.01037)	0.0059 (0.0000141)	-0.9398	-0.9878	0.1392
CAE	7	2	2	0.2857 (0.04010)	0.0004 (0.0000003)	-0.0947	-1.0062	0.3499
JUA	4	3	2	0.8333 (0.03516)	0.0029 (0.0000060)	0.1335	-0.7801	0.3062
PAR	9	8	8	0.9722 (0.00256)	0.0079 (0.0000228)	-4.6211**	-1.3164	0.0808*
SER	6	6	6	1.0000 (0.00463)	0.0068 (0.0000201)	-2.2989	-0.6816	0.1430
TRI	10	4	4	0.7778 (0.00633)	0.0064 (0.0000153)	2.5516	1.0012	0.2088
CAR	7	1	0	0.0000 (0.00000)	0.0000 (0.0000000)	-	-	-
CUM	7	1	0	0.0000 (0.00000)	0.0000 (0.0000000)	-	-	-
LIM	6	2	1	0.5333 (0.02469)	0.0008 (0.0000007)	-3.1497**	0.8506	0.2667
WES	58	42	42	0.9806 (0.00006)	0.0087 (0.0000220)	-25.3085**	-2.1152*	0.0353*
EAS	20	2	2	0.1895 (0.01191)	0.0003 (0.0000002)	-3.8855**	-0.5915	0.0947
Total	78	44	-	0.9381 (0.00041)	0.0090 (0.0000229)	-25.2403**	-1.9887*	0.0365*

Table 2. Summary results of the Analysis of Molecular Variance (AMOVA) computed from the mitochondrial data set of *Jaguajir rochae*, showing the proportion of genetic variation explained between haplogroups (eastern and western), as well as for comparisons among populations within haplogroups, and at intrapopulation level. d.f. = degrees of freedom.

Comparative level	d.f.	Variation (%)	P-value
Among haplogroups	1	44.05	< 0.01
Among populations within haplogroups	9	19.40	< 0.01
Within populations	67	36.55	0.01

Table 3. Migrate-n model comparisons for different migration schemes among populations of *Jaguajir rochae*, based on the partial sequences of the mitochondrial gene cytochrome c oxidase I. Model choice was ranked by the lowest values of log Bayes Factor (LBF) computed for each migration model.

Model	Bézier (lnL)	LBF	Model probability	Model choice
A	-1774.68	-19.90	0.00	9
B	-1756.85	-2.07	0.04	7
C	-1760.46	-5.68	0.00	8
D	-1754.78	0.00	0.29	1
E	-1756.72	-1.94	0.04	6
F	-1754.88	-0.01	0.27	2
G	-1755.88	-1.10	0.10	4
H	-1755.32	-0.54	0.17	3
I	-1755.98	-1.20	0.09	5

Figure Legends

Figure 1. (a) Spatial distribution of sampling locations of *Jaguajir rochae* in northeastern Brazil: Afogados da Ingazeira (AFO), Água Branca (AGU), Buíque (BUI), Caetés (CAE), Caruaru (CAR), Cumaru (CUM), Juazeiro do Norte (JUA), Limoeiro (LIM), Parnamirim (PAR), Serra Talhada (SER), and Triunfo (TRI). The point matrix represents the spatial cover of the Caatinga domain, while the transition zone between Caatinga and Coastal Atlantic Forest (crossline matrix) is represented by the horizontal-line matrix; BOP = Borborema Plateau. (b) BAPS chart depicting the western (green) and eastern (red) recovered haplogroups; the colors in the vertical bars are in accordance to the map, identifying the population of which each specimen (horizontal bar) belongs. (c) Median-joining haplotype network illustrating the spatial pattern of sampled haplotypes; the size of each circle is proportional to haplotype frequencies, while colors are population identifiers, according to the map.

Figure 2. Migration models implemented in Migrate-n to test the hypothesis of a single panmitic population of *Jaguajir rochae* distributed around the Borborema Plateau (BOP), and three migration schemes between the western (W) and eastern (E) predefined groups of populations. Five additional migration models were tested (e-i), considering the population of Água Branca (I) as a migration source for the group comprising the remaining populations from the western side of the BOP (II), as well as for a second group containing populations from the eastern side of the BOP (III).

Figure 3. Bayesian dated phylogeny inferred for non-redundant haplotypes of *Jaguajir rochae*, estimated from a partial sequence of the mitochondrial gene cytochrome c oxidase I. Black circles represent nodes with posterior probability ≥ 0.95 .

Figure 4. Binary maps reconstructed from ecological niche models, illustrating the potential distribution of niche suitability of *Jaguajir rochae* at three hypothetical climatic scenarios: (a) Last Interglacial (LIG), (b) Last Glacial Maxima (LGM), and current climate (c). Spatial shifts on niche suitability of *J. rochae* from LIG to LGM, and from LGM to the present are displayed by arrows in map (d), and (e), respectively (see Materials and Methods for details). (f) Niche suitability map

showing historically stable (black), and unstable (shades of gray) areas for the occurrence of *J. rochae* through late-Quaternary. The spatial delimitation of the Borborema Plateau ecoregion (dashed polygon) follows Velloso et al. (2002).

Figure 1

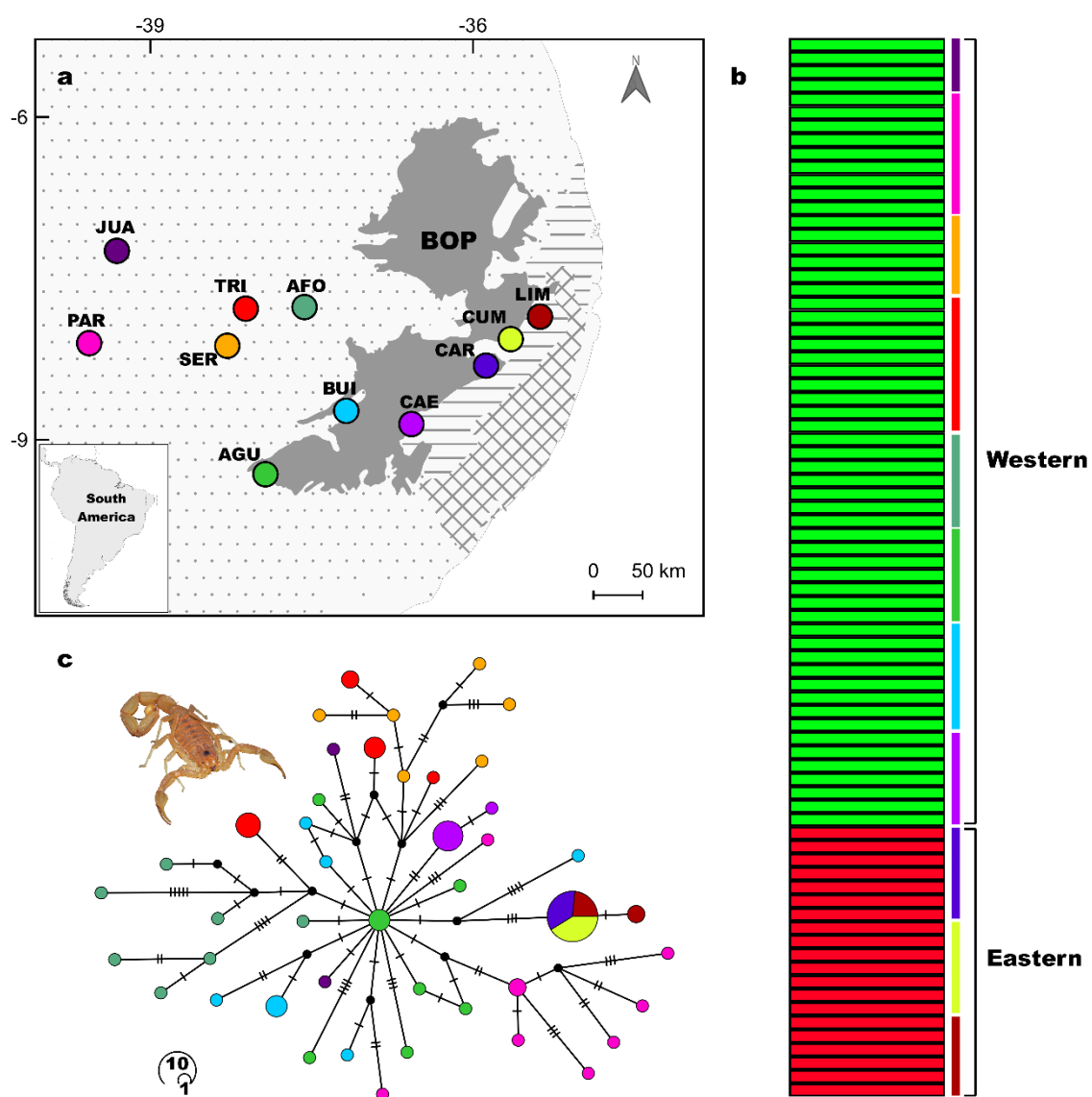


Figure 2

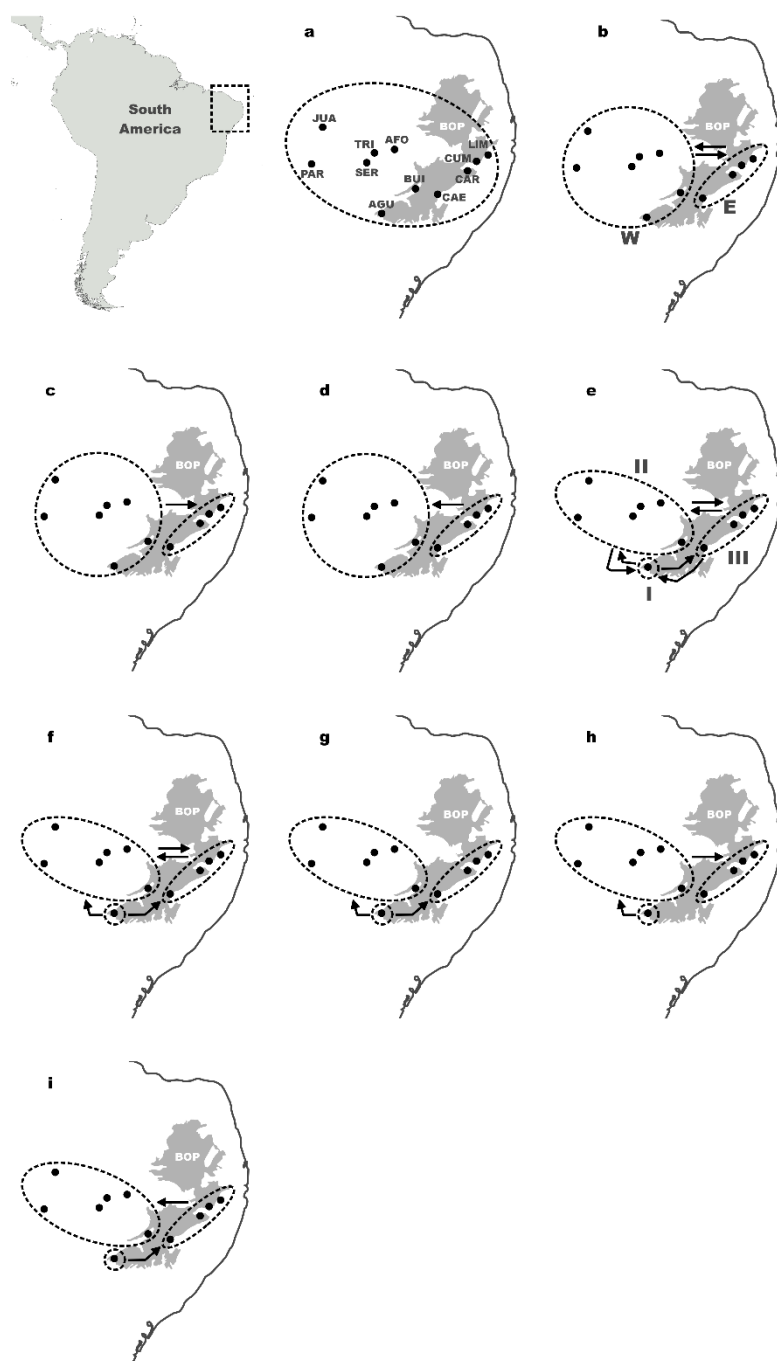


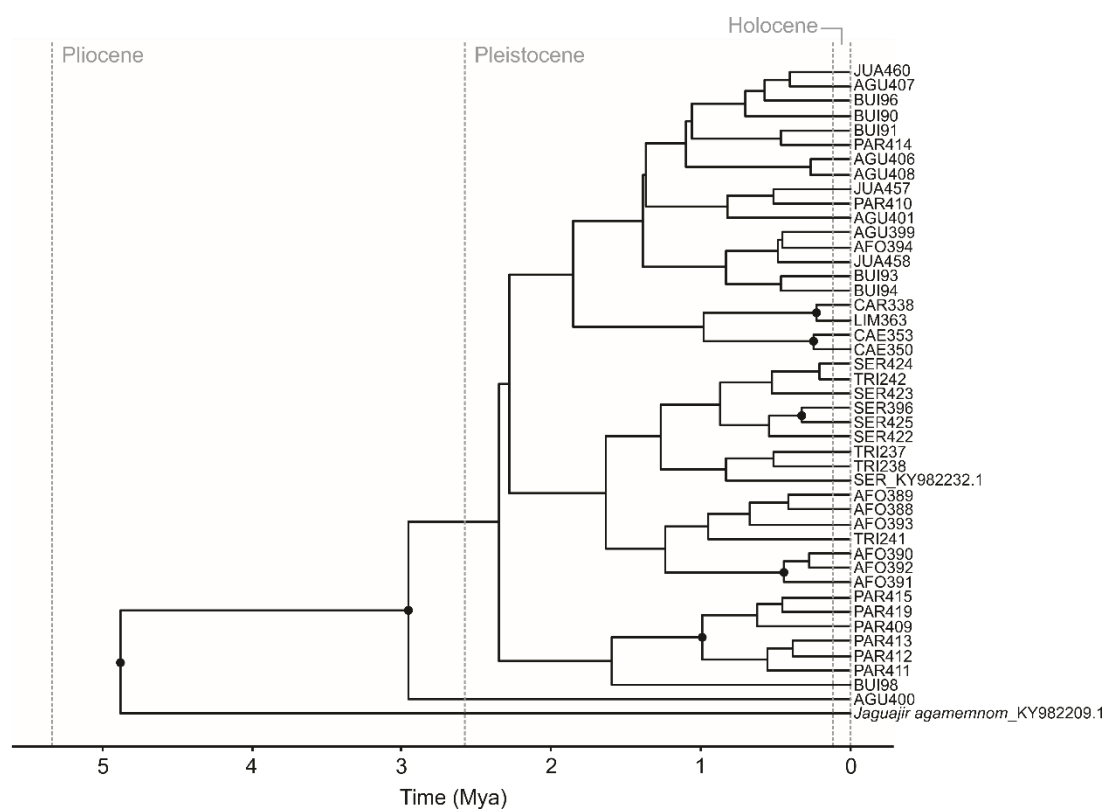
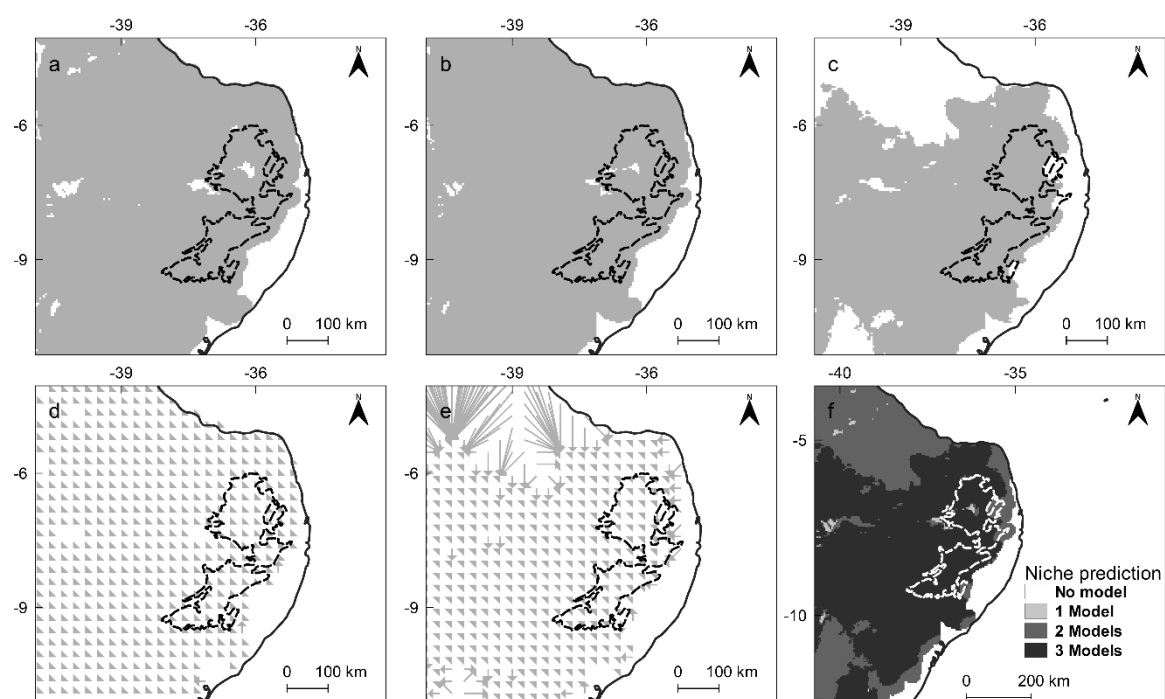
Figure 3

Figure 4

SUPPORTING INFORMATION

Spatial patterns of genetic diversity in the scorpion *Jaguajir rochae* (Scorpiones, Buthidae) as proxies to understand the role of historical habitat stability and geological barriers on lineage diversification processes in Caatinga environments

Stênio Ítalo A. Foerster, Bárbara Natieli S. Pereira, André Felipe de A. Lira, Juliana F. Lima, Ricardo Pinto-da-Rocha and Valdir de Queiroz Balbino

Appendix S1. Supplementary tables

Table S1. List of zoological collections (institutions) accessed to retrieve occurrence points of *Jaguajir rochae* used to construct ecological niche models.

Institution	Location	Curator
Instituto Butantan	São Paulo, Brazil	A.D. Brescovit
Museu de História Natural da Bahia	Salvador, Brazil	T.K. Brazil
Museu Nacional/Universidade Federal do Rio de Janeiro	Rio de Janeiro, Brazil	A.B. Kury
Museu Paraense Emílio Goeldi	Belém, Brazil	A.B. Bonaldo
Universidade Federal de Minas Gerais	Belo Horizonte, Brazil	A.J. Santos
Universidade Federal da Paraíba	João Pessoa, Brazil	M.B. da Silva
Universidade Federal de Pernambuco	Recife, Brazil	C.M.R. Albuquerque
Universidade Federal do Piauí	Floriano, Brazil	L.S. Carvalho

Table S2. Numerical results of model (D) and (F) estimated in Migrate-n using the mitochondrial data set of *Jaguajir rochae*. The mean values of mutation-scaled immigration rates (M) are presented, followed by the mean number of immigrants per generation given in parenthesis. The mean scaled values of effective population size (Θ) of each predefined group are also shown.

Model	Θ_W	Θ_E	M_{E-W}	Θ_I	Θ_{II}	Θ_{III}	M_{I-II}	M_{I-III}	M_{II-III}	M_{III-II}
D	0.088	0.003	198.5 (17.5)	-	-	-	-	-	-	-
F	-	-	-	0.023	0.082	0.003	490.1 (40.2)	201.8 (0.60)	232.8 (0.70)	68.7 (5.63)

SUPPORTING INFORMATION

Spatial patterns of genetic diversity in the scorpion *Jaguajir rochae* (Scorpiones, Buthidae) as proxies to understand the role of historical habitat stability and geological barriers on lineage diversification processes in Caatinga environments

Stênio Ítalo A. Foerster, Bárbara Natieli S. Pereira, André Felipe de A. Lira, Juliana F. Lima, Ricardo Pinto-da-Rocha and Valdir de Queiroz Balbino

Appendix S2. Supplementary figures

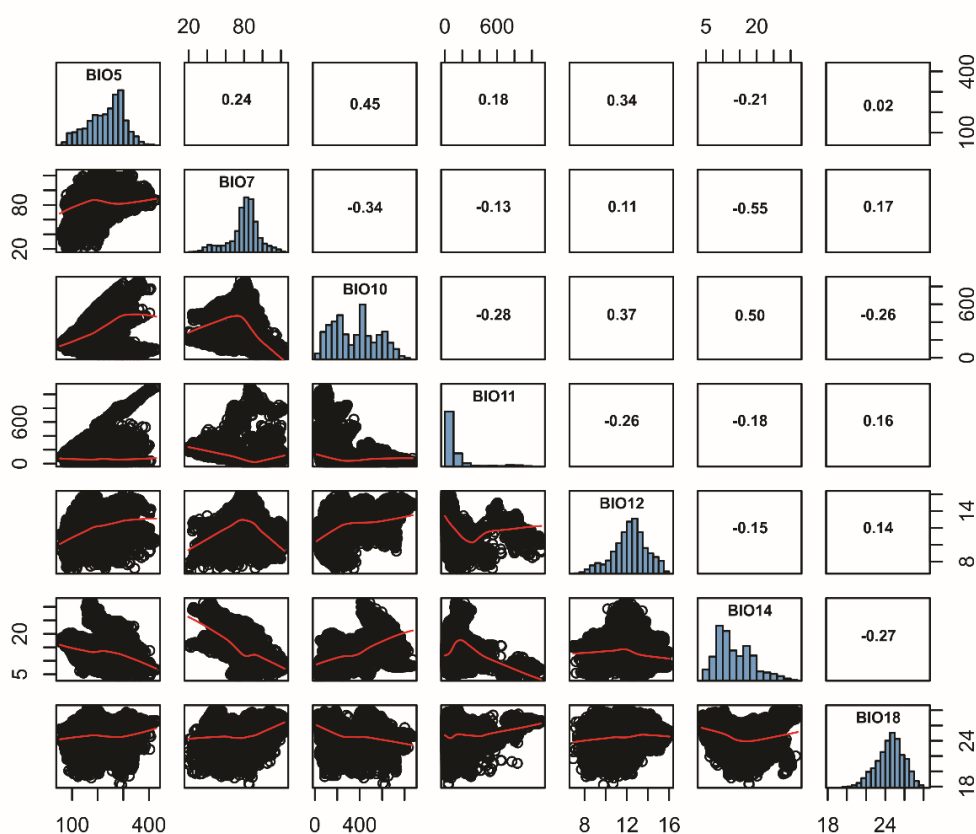


Figure S1. Pair plot depicting patterns of autocorrelation among bioclimatic variables used to construct ecological niche models for *Jaguajir rochae* (lower diagonal). Histograms at diagonal display the shape distribution of bioclimatic predictors, and Pearson correlation values among them are given at upper diagonal.

4 DISCUSSÃO GERAL

O alto poder informativo oriundo de análises filogeográficas, e a notória escassez de estudos desta natureza com invertebrados da Caatinga (Turchetto-Zolet *et al.*, 2013; Magalhães *et al.*, 2014), nos incentivaram a avaliar os padrões espaciais de diversidade genética do escorpião *Jaguajir rochae*. Nossos resultados indicam que áreas de Caatinga historicamente estáveis abrigam populações de *J. rochae* com elevados níveis de diversidade genética, mostrando que tais áreas certamente atuam como refúgios históricos para espécies com baixa capacidade de dispersão (Werneck *et al.*, 2011). Em adição, nós demonstramos empiricamente que o Planalto da Borborema restringe expressivamente o fluxo gênico entre populações de *J. rochae*, mas que sua impermeabilidade não é absoluta. Embora nossos dados não nos tenham permitido determinar os pontos de menor resistência ao fluxo gênico ao longo do Planalto da Borborema, isto pode servir como iniciativa para um novo estudo, com o objetivo de testar diferentes padrões de conectividade entre populações de *J. rochae* a partir de dados genéticos, climáticos e topográficos, os quais podem por exemplo, ser implementados em modelos de conectividade baseados na teoria de circuitos (McRae *et al.*, 2008).

As análises de diferenciação genética e datação molecular indicam um padrão complexo de diversificação e divergência entre as linhagens mitocondriais de *J. rochae*, que não podem ser interpretadas unicamente por possíveis alterações de paisagem ocasionadas pelas mudanças climáticas do Quaternário. Similarmente, nossos resultados indicam que os processos de diversificação intraespecífica em *J. rochae* ocorreram após o fim do soerguimento do Planalto da Borborema, sugerindo que uma hipótese de diversificação alopátrica mediada por esta formação geológica é pouco provável. Considerando que os nossos resultados

também indicaram que o nicho climático de *J. rochae* permaneceu praticamente estável desde o último período interglacial (~ 120-140 mil anos atrás), é possível que os haplótipos tenham se diferenciado dentro de um habitat historicamente estável, mas com variações clinais, e que tais processos ocorreram na presença de fluxo gênico (Nosil, 2008). É importante destacar, no entanto, que os nossos resultados devem ser interpretados como indicativos, e que abordagens de caráter multiloci são necessárias para validar os modelos propostos por nós. Similarmente, a inclusão de informações sobre plasticidade ecológica/fisiológica e história natural (ainda indisponíveis para *J. rochae*) podem potencializar o poder preditivo e a acurácia dos modelos de distribuição de espécie (Dormann *et al.*, 2011; Mellert *et al.*, 2011), e por isso, nós incentivamos a elaboração de estudos dessa natureza não apenas com escorpiões, mas com quaisquer táxons distribuídos em ambientes de Caatinga.

Recentemente, a comunidade acadêmica tem empenhado um esforço progressivo aos estudos sobre a biodiversidade da Caatinga, favorecendo o surgimento de novas perspectivas sobre os padrões e processos responsáveis por gerar e preservar a fauna e a flora destes ambientes. Certamente, o maior legado destes avanços foi a mudança de percepção da própria comunidade acadêmica em relação à biodiversidade da Caatinga, antes vista como irrelevante perante alguns pesquisadores (Leal *et al.*, 2005). Os benéficos desta mudança de percepção podem ser traduzidos no âmbito acadêmico, pelo crescente interesse por parte dos pesquisadores, e social, uma vez que o acúmulo progressivo de informações sobre a biodiversidade da Caatinga assegura o planejamento de políticas ambientais que visam preservar os diversos serviços ecossistêmicos. Nesse sentido, nós acreditamos que o presente estudo servirá, primariamente, como fonte de estudos

para fins acadêmicos, mas com potencial para fundamentar práticas de conservação da Caatinga, visto que a estabilidade histórica do habitat deve ser incluída como critério de avaliação de áreas de conservação da biodiversidade.

5 CONCLUSÕES

Nós concluimos que, em termos históricos, a estabilidade climática da Caatinga pode prever elevados índices de diversidade genética em *J. rochae*, e que o Planalto da Borborema constitui uma importante barreira física ao fluxo gênico entre populações desse escorpião. Em adição, há um padrão complexo de diversificação de linhagens mitocondriais de *J. rochae*, que certamente é influenciado pela estabilidade da Caatinga ao longo do tempo, ao passo em que pouco efeito é atribuído à história geomorfológica do Planalto da Borborema. Por fim, nós demonstramos que o nicho climático de *J. rochae* certamente sofreu poucas mudanças espaciais desde o último período interglacial, corroborando a hipótese de que os processos de diversificação intraespecífica ocorreram em um ambiente estável e provavelmente na presença de fluxo gênico.

REFERÊNCIAS BIBLIOGRÁFICAS

- Ab'Sáber AN (1974) O Domínio Morfoclimático Semi-árido das Caatingas Brasileiras. *Geomorfologia* 1:1-38.
- Ab'Sáber AN (1977) Espaços ocupados pela expansão dos climas secos na América do Sul por ocasião dos períodos glaciais Quaternários. *Paleoclimas* 3:1-19.
- Aleixo A and de Fátima-Rossetti D (2007) Avian gene trees, landscape evolution, and geology: towards a modern synthesis of Amazonian historical biogeography?. *J Ornithol* 148:443-453.
- Alfaro LJW, Boubli JP, Olson LE, Di Fiore A, Wilson B, Espeleta G, Chiou KL, Schulte M, Neitzel S, Ross V, Schowochow D, Nguyen MTT, Farias I, Janson CH and Alfaro ME (2012). Explosive Pleistocene range expansion leads to widespread Amazonian sympatry between robust and gracile capuchin monkeys. *J Biogeogr* 39:272-288.
- Álvarez-presas M, Carbayo F, Rozas J and Riutort M (2011) Land planarians (Platyhelminthes) as a model organism for fine-scale phylogeographic studies: understanding patterns of biodiversity in the Brazilian Atlantic Forest hotspot. *J Evol Biol* 24:887-896.
- Angelieri CCS, Adams-Hosking C, de Barros KMPM, de Souza MP and McAlpine CA (2016) Using species distribution models to predict potential landscape restoration effects on puma conservation. *PLoS One* 11:e0145232.
- Apgaua DMG, dos Santos RM, Pereira DGS, Menino GCO, Pires GG, Fontes MAL and Tng DYP (2014) Beta-diversity in seasonally dry tropical forests (SDTF) in the Caatinga Biogeographic Domain, Brazil, and its implications for conservation. *Biodiversity and Conserv* 23:217-232.
- Arruda DM, Schaefer CE, Fonseca RS, Solar RR and Fernandes-Filho EI (2018) Vegetation cover of Brazil in the last 21 ka: New insights into the Amazonian refugia and Pleistocenic arc hypotheses. *Global Ecol Biogeogr* 27:47-56.
- Avice JC (2000) *Phylogeography: the history and formation of species*. Harvard University Press, Cambridge, 447 p.
- Avice JC (2009) *Phylogeography: retrospect and prospect*. *J Biogeogr* 36:3-15.
- Bartoletti LFDM, Peres EA, Sobral-Souza T, Fontes FVHM, Silva MJD and Solferini VN (2017) Phylogeography of the dry vegetation endemic species *Nephila*

- sexpunctata* (Araneae: Araneidae) suggests recent expansion of the Neotropical Dry Diagonal. *J Biogeogr* 44:2007-2020.
- Behling H (1998) Late Quaternary vegetational and climatic changes in Brazil. *Rev Palaeobot Palynol* 99:143-156.
- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsche G, Pütz J, Middendorf M and Stadler P (2013). MITOS: improved de novo metazoan mitochondrial genome annotation. *Mol Phylogenetics Evol* 69:313-319.
- Bond N, Thomson J, Reich P and Stein J (2011) Using species distribution models to infer potential climate change-induced range shifts of freshwater fish in south-eastern Australia. *Mar Freshw Res* 62:1043-1061.
- Brown KS, Sheppard PM, and Turner JRG (1974) Quaternary refugia in tropical America: evidence from race formation in *Heliconius* butterflies. *Proc R Soc Lond B Biol Sci* 187: 369-378.
- Bucher EH (1982) Chaco and Caatinga South American arid savannas, woodlands and thickets. In: Huntley BJ and Walker BH (eds) *Ecology of Tropical Savannas*. Springer-Verlag Berlin Heidelberg, Germany, pp 48-79.
- Bueno ML, Pennington RT, Dexter KG, Kamino LHY, Pontara V, Neves DM, Ratter JA and de Oliveira-Filho AT (2016) Effects of Quaternary climatic fluctuations on the distribution of Neotropical savanna tree species. *Ecography* 40:403-414.
- Bush MB and Oliveira PED (2006) The rise and fall of the Refugial Hypothesis of Amazonian speciation: a paleoecological perspective. *Biot Neotrop* 6:1-17.
- Bryson Jr RW, Savary WE and Prendini L (2013a) Biogeography of scorpions in the *Pseudouroctonus minimus* complex (Vaejovidae) from south-western North America: implications of ecological specialization for pre-Quaternary diversification. *J Biogeogr* 40:1850-1860.
- Bryson Jr RW, Riddle BR, Graham MR, Smith BT and Prendini L (2013b) As old as the hills: montane scorpions in southwestern North America reveal ancient associations between biotic diversification and landscape history. *PLoS One* 8:e52822.
- Bryson RW, Prendini L, Savary WE and Pearman PB (2014) Caves as microrefugia: Pleistocene phylogeography of the troglomorphic North American scorpion *Pseudouroctonus reddelli*. *BMC Evol Biol* 14:1-16.

- Bryson Jr RW, Savary WE, Zellmer AJ, Bury RB and McCormack JE (2016) Genomic data reveal ancient microendemism in forest scorpions across the California Floristic Province. *Mol Ecol* 25:3731-3751.
- Cabanne GS, d'Horta FM, Sari EH, Santos FR and Miyaki CY (2008) Nuclear and mitochondrial phylogeography of the Atlantic forest endemic *Xiphorhynchus fuscus* (Aves: Dendrocolaptidae): Biogeography and systematics implications. *Mol Phylogenetics Evol* 49:760-773.
- Caetano S, Prado D, Pennington RT, Beck S, Oliveira-Filho A, Spichiger R and Naciri Y (2008) The history of seasonally dry tropical forests in eastern South America: inferences from the genetic structure of the tree *Astronium urundeuva* (Anacardiaceae). *Mol Ecol* 17:3147-3159.
- Camargo AJA and Becker VO (1999) Saturniidae (Lepidoptera) from the Brazilian Cerrado: Composition and Biogeographic Relationships. *Biotropica* 3:696-705.
- Cardoso DBOS and Queiroz LP (2007) Diversidade de Leguminosae nas Caatingas de Tucano, Bahia: implicações para a fitogeografia do semi-árido do Nordeste do Brasil. *Rodriguésia* 58:379-391.
- Carmo RFR, Amorim HP and Vasconcelos SD (2013) Scorpion diversity in two types of seasonally dry tropical forest in the semi-arid region of Northeastern Brazil. *Biota Neotrop* 13:340-344.
- Carnaval AC and Bates JM (2007) Amphibian DNA shows marked genetic structure and tracks Pleistocene climate change in northeastern Brazil. *Evolution: Int J Org Evol* 61:2942-2957.
- Carnaval AC and Moritz C (2008) Historical climate modelling predicts patterns of current biodiversity in the Brazilian Atlantic forest. *J Biogeogr* 35:1187-1201.
- Carnaval AC, Hickerson MJ, Haddad CF, Rodrigues MT and Moritz C (2009) Stability predicts genetic diversity in the Brazilian Atlantic forest hotspot. *Science* 323:785-789.
- Castro DP, Rodrigues JFM, Borges-Leite MJ, Lima DC and Borges-Nojosa DM (2019) Anuran diversity indicates that Caatinga relictual Neotropical forests are more related to the Atlantic Forest than to the Amazon. *PeerJ* 6:e6208.
- Ceccarelli FS, Ojanguren-Affilastro AA, Ramírez MJ, Ochoa JA, Mattoni CI and Prendini L (2016a) Andean uplift drives diversification of the bothriurid scorpion genus *Brachistosternus*. *J Biogeogr* 43:1942-1954.

- Ceccarelli FS, Pizarro-Araya J and Ojanguren-Affilastro AA (2017) Phylogeography and population structure of two *Brachistosternus* species (Scorpiones: Bothriuridae) from the Chilean coastal desert—the perils of coastal living. *Biol J Linn Soc* 120:75-89.
- Chen WJ, Bu Y, Carapelli A, Dallai R, Li S, Yin WY and Luan YX (2011) The mitochondrial genome of *Sinentomon erythranum* (Arthropoda: Hexapoda: Protura): an example of highly divergent evolution. *BMC Evol Biol* 11:246.
- Cheng H, Sinha A, Cruz FW, Wang X, Edwards RL, d'Horta FM, Ribas CC, Vuille M, Stott LD and Auler AS (2013) Climate change patterns in Amazonia and biodiversity. *Nature Comm* 4:n1411.
- Choi EH, Park SJ, Jang KH and Hwang W (2007) Complete mitochondrial genome of a Chinese scorpion *Mesobuthus martensii* (Chelicerata, Scorpiones, Buthidae). *DNA Sequence* 18:461-473.
- Chunco AJ, Phimmachak S, Sivongxay N and Stuart BL (2013) Predicting environmental suitability for a rare and threatened species (Lao Newt, *Laotriton laoensis*) using validated species distribution models. *PLoS One* 8:e59853.
- Colinvaux PA, Irion G, Räsänen ME, Bush MB and Nunes de Mello JAS (2001) A paradigm to be discarded: Geological and paleoecological data falsify the Haffer & Prance refuge hypothesis of Amazonian speciation. *Amazoniana* 16:609-646.
- Connor EF (1986) The role of Pleistocene forest refugia in the evolution and biogeography of tropical biotas. *Trends Ecol Evol* 1:165-168.
- Côrtes ALA, Rapini A and Daniel TF (2015) The *Tetramerium* lineage (Acanthaceae: Justicieae) does not support the Pleistocene Arc hypothesis for South American seasonally dry forests. *Am J Bot* 102:992-1007.
- Costa LP (2003) The historical bridge between the Amazon and the Atlantic Forest of Brazil: a study of molecular phylogeography with small mammals. *J Biogeogr* 30:71-86.
- Costa LP, Leite YL, da Fonseca GA and da Fonseca MT (2000) Biogeography of South American forest mammals: endemism and diversity in the Atlantic Forest. *Biotropica* 32:872-881.
- Costa GM, Cardoso D, Queiroz LP and Conceição AA (2015) Variações locais na riqueza florística em duas ecorregiões de caatinga. *Rodriguésia* 66:685-709.

- Costa WJ, Amorim PF and Mattos JLO (2018) Synchronic historical patterns of species diversification in seasonal aplocheiloid killifishes of the semi-arid Brazilian Caatinga. *PloS one* 13:e0193021.
- Couvreux TL, Forest F and Baker WJ (2011) Origin and global diversification patterns of tropical rain forests: inferences from a complete genus-level phylogeny of palms. *BMC Biology* 9:1-12.
- Crawford PH and Hoagland BW (2010) Using species distribution models to guide conservation at the state level: the endangered American burying beetle (*Nicrophorus americanus*) in Oklahoma. *J Insec Conserv* 14:511-521.
- Dagosta FC and Pinna MD (2017) Biogeography of Amazonian fishes: deconstructing river basins as biogeographic units. *Neotrop Ichthyol* 15:e170034.
- Damuth JE and Fairbridge RW (1970) Equatorial Atlantic deep-sea arkosic sands and ice-age aridity in tropical South America. *Geo Soc Am Bull* 81:189-206.
- De-Nova JA, Medina R, Montero JC, Weeks A, Rosell JA, Olson ME, Eguiarte LE and Megallón S (2012) Insights into the historical construction of species-rich Mesoamerican seasonally dry tropical forests: the diversification of *Bursera* (Burseraceae, Sapindales). *New Phytol* 193:276-287.
- Dormann CF (2007) Promising the future? Global change projections of species distributions. *Basic Appl Ecol* 8:387-397.
- Dormann CF, Schymanski SJ, Cabral J, Chuine I, Graham C, Hartig F, Kearney M, Morin X, Römermann C, Schröder B and Singer A (2011) Correlation and process in species distribution models: bridging a dichotomy. *J Biogeogr* 39:2119-2131.
- Elith J and Leathwick JR (2009) Species distribution models: ecological explanation and prediction across space and time. *Annu Rev Ecol Evol Syst* 40:677-697.
- Esposito LA and Prendini L (2019) Island Ancestors and New World Biogeography: A Case Study from the Scorpions (Buthidae: Centruroidinae). *Scientific Rep* 9:3500.
- Esposito LA, Yamaguti HY, Souza CA, Pinto-Da-Rocha R and Prendini L (2017) Systematic Revision of the Neotropical Club-Tailed Scorpions, *Physoctonus*, *Rhopalurus*, and *Troglorhopalurus*, Revalidation of *Heteroctenus*, and Descriptions of Two New Genera and Three New Species (Buthidae: Rhopalurusinae). *Bull Am Mus Nat Hist* 2017:1-137.

- Faye A, Deblauwe V, Mariac C, Richard D, Sonké B, Vigouroux Y and Couvreur TLP (2016) Phylogeography of the genus *Podococcus* (Palmae/Arecaceae) in Central African rain forests: Climate stability predicts unique genetic diversity. *Mol Phylogenetics Evol* 105:126-138.
- Fernandes AM, Wink M and Aleixo A (2012) Phylogeography of the chestnut-tailed antbird (*Myrmeciza hemimelaena*) clarifies the role of rivers in Amazonian biogeography. *J Biogeogr* 39:1524-1535.
- Fernandes AM, Gonzalez J, Wink M and Aleixo A (2013) Multilocus phylogeography of the Wedge-billed Woodcreeper *Glyphorhynchus spirurus* (Aves, Furnariidae) in lowland Amazonia: Widespread cryptic diversity and paraphyly reveal a complex diversification pattern. *Mol Phylogenetics Evol* 66:270-282.
- Ficetola GF, Thuiller W and Miaud C (2007) Prediction and validation of the potential global distribution of a problematic alien invasive species – the American bullfrog. *Divers Distrib* 13:476-485.
- Fischer C, Koblmüller S, Güllý C, Schlötterer C, Sturmbauer C and Thallinger GG (2013) Complete mitochondrial DNA sequences of the threadfin cichlid (*Petrochromis trewavasae*) and the blunthead cichlid (*Tropheus moorii*) and patterns of mitochondrial genome evolution in cichlid fishes. *PLoS One* 8: e67048.
- Fitzpatrick MC and Hargrove WW (2009) The projection of species distribution models and the problem of non-analog climate. *Biodivers Conserv* 18:e2255.
- Fois M, Bacchetta G, Cuenca-Lombrana A, Cogoni D, Pinna MS, Sulis E and Fenu G (2018) Using extinctions in species distribution models to evaluate and predict threats: a contribution to plant conservation planning on the island of Sardinia. *Environm Conserv* 45:11-19.
- Franklin J (2013) Species distribution models in conservation biogeography: developments and challenges. *Divers Distrib* 19:1217-1223.
- Fritz U, Stuckas H, Vargas-Ramírez M, Hundsdörfer AK, Maran J, and Päckert M (2011) Molecular phylogeny of Central and South American slider turtles: implications for biogeography and systematics (Testudines: Emydidae: *Trachemys*). *J Zool Syst Evol Res* 50:125-136.
- Gantenbein B and Largiadèr CR (2003) The phylogeographic importance of the Strait of Gibraltar as a gene flow barrier in terrestrial arthropods: a case study

- with the scorpion *Buthus occitanus* as model organism. *Mol Phylogenetics Evol* 28:119-130.
- Gehara M, Barth A, de Oliveira EF, Costa MA, Haddad CFB and Vences M (2017) Model-based analyses reveal insular population diversification and cryptic frog species in the *Ischnocnema parva* complex in the Atlantic forest of Brazil. *Mol Phylogenetics Evol* 112:68-78.
- Gillespie RG and Roderick GK (2014) Evolution: geology and climate drive diversification. *Nature* 509:297.
- Gomes APS, Rodal MJN and Melo AL (2006) Florística e fitogeografia da vegetação arbustiva subcaducifólia da Chapada de São José, Buíque, PE, Brasil. *Acta Bot Bras* 20:37-48.
- Gory JJ, Herbei R and Kubatko LS (2018) Bayesian inference of selection in the Wright-Fisher diffusion model. *Stat Appl Genet Mol Biol* 17:1-33.
- Graham MR, Oláh-Hemmings V and Fet V (2012) Phylogeography of co-distributed dune scorpions identifies the Amu Darya River as a long-standing component of Central Asian biogeography: (Scorpiones: Buthidae). *Zool Middle East* 55:95-110.
- Graham MR, Jaeger JR, Prendini L and Riddle BR (2013a) Phylogeography of Beck's desert scorpion, *Paruroctonus becki*, reveals Pliocene diversification in the Eastern California Shear Zone and postglacial expansion in the Great Basin Desert. *Mol Phylogenetics Evol* 69:502-513.
- Graham MR, Jaeger JR, Prendini L and Riddle BR (2013b) Phylogeography of the Arizona hairy scorpion (*Hadrurus arizonensis*) supports a model of biotic assembly in the Mojave Desert and adds a new Pleistocene refugium. *J Biogeogr* 40:1298-1312.
- Graham MR, Bryson Jr RW and Riddle BR (2014) Late Pleistocene to Holocene distributional stasis in scorpions along the Baja California peninsula. *Biol J Linn Soc* 111:450-461.
- Guisan A and Thuiller W (2005) Predicting species distribution: offering more than simple habitat models. *Ecol Lett* 8:993-1009.
- Guisan A and Zimmermann NE (2000) Predictive habitat distribution models in ecology. *Ecol Modell* 135:147-186.
- Habel JC, Husemann M, Schmitt T, Zachos FE, Honnen AC, Petersen B, Parmakelis A and Stathi I (2012) Microallopatry caused strong diversification in

- Buthus scorpions* (Scorpiones: Buthidae) in the Atlas Mountains (NW Africa). PLoS one 7:e29403.
- Haffer J (1969) Speciation in Amazonian Forest Birds. Science 165:131-136.
- Hamann A and Aitken SN (2013) Conservation planning under climate change: accounting for adaptive potential and migration capacity in species distribution models. Divers Distrib 19:268-280.
- Hein J, Schierup MH and Wiuf C (2005) Gene Genealogies, Variaton and Evolution. Oxford University Press, NY, 291 p.
- Hellberg ME (2006) No variation and low synonymous substitution rates in coral mtDNA despite high nuclear variation. BMC Evol Biol 6:24.
- Heuner M, Weber A, Schröder U, Kleinschmit B and Schröder B (2016) Facilitating political decisions using species distribution models to assess restoration measures in heavily modified estuaries. Mar Poll Bull 110:250-260.
- Hewitt GM (2004) Genetic consequences of climatic oscillations in the Quaternary. Philos Trans R Soc Lond B Biol Sci 359:183-195.
- Hillis DM and Dixon MT (1991) Ribossomal DNA: Molecular Evolution and Phylogenetic Inference. Q Rev Biol. 66:411-453.
- Hoorn C, Wesselingh FP, ter Steege H, Bermudez MA, Mora A, Sevink J, Sanmartín I, Sanchez-Meseguer A, Anderson CL, Figueiredo JP, Jaramillo C, Riff D, Negri FR, Hooghiemstra H, Lundberg J, Stadler T, Särkinen T and Antonelli A (2010) Amazonia Through Time: Andean Uplift, Climate Change, Landscape Evolution, and Biodiversity. Science 330:927-931.
- Hudson RR (2002) Generating samples under a Wright–Fisher neutral model of genetic variation. Bioinformatics 18:337-338.
- Hugall A, Moritz C, Moussalli A and Stanisic J (2002) Reconciling paleodistribution models and comparative phylogeography in the Wet Tropics rainforest land snail *Gnarosophia bellendenkerensis* (Brazier 1875). Proc Nat Acad Sc 99:6112-6117.
- Husemann M, Schmitt T, Stathi I and Habel JC (2012) Evolution and radiation in the scorpion *Buthus elmoutaouakili* Lourenco and Qi 2006 (Scorpiones: Buthidae) at the foothills of the Atlas Mountains (North Africa). J Heredity 103:221-229.
- Hwang UW, Kim W, Tautz D and Friedrich M (1998) Molecular phylogenetics at the Felsenstein zone: approaching the Strepsiptera problem using 5.8 S and 28S rDNA sequences. Mol Phylogenetics Evol 9:470-480.

- IBGE (2004): Instituto Brasileiro de Geografia e Estatística, <https://www.ibge.gov.br/geociencias-novoportal/informacoes-ambientais/estudos-ambientais/15842-biomas.html?=&t=downloads> (January 15, 2019).
- Ji YJ, Zhang DX and He LJ (2003) Evolutionary conservation and versatility of a new set of primers for amplifying the ribosomal internal transcribed spacer regions in insects and other invertebrates. *Mol Ecol* 3:581-585.
- Kaky E and Gilbert F (2016) Using species distribution models to assess the importance of Egypt's protected areas for the conservation of medicinal plants. *J Arid Environm* 135:140-146.
- Knapp S and Mallet J (2003) Refuting refugia?. *Science* 300:71-72.
- Keppel G, Van Niel KP, Wardell-Johnson GW, Yates CJ, Byrne M, Mucina L, Schut AG, Hopper SD and Franklin SE (2012) Refugia: identifying and understanding safe havens for biodiversity under climate change. *Global Ecol and Biogeogr* 21:393-404.
- Kingman JFC (1982a) On the Genealogy of Large Populations. *J Appl Prob* 19:27-43.
- Kingman JFC (1982b) The Coalescent. *Stoch Process Their Appl* 13:253-248.
- Knowles LL (2009) Statistical phylogeography. *Ann Rev Ecol Evol Syst* 40:593-612.
- Leal BS, Medeiros LR, Peres EA, Sobral-Souza T, Palma-Silva C, Romero GQ and Carareto CM (2018) Insights into the evolutionary dynamics of Neotropical biomes from the phylogeography and paleodistribution modeling of *Bromelia balansae*. *Am J Bot* 105:1725-1734.
- Leal IR, Silva JD, Tabarelli M and Lacher Jr TE (2005) Mudando o curso da conservação da biodiversidade na Caatinga do Nordeste do Brasil. *Megadiversidade* 1:139-146.
- Leite RN and Rogers DS (2013) Revisiting Amazonian phylogeography: insights into diversification hypotheses and novel perspectives. *Org Divers Evol* 13:639-664.
- Leite YL, Costa LP, Loss AC, Rocha RG, Batalha-Filho H, Bastos AC, Quaresma VS, Fagundes V, Paresque R, Passamani M and Pardini R (2016). Neotropical forest expansion during the last glacial period challenges refuge hypothesis. *Proc Nat Acad Sci* 113:1008-1013.

- Lima MG, Buckner JC, Silva-Júnior Jd, Aleixo A, Martins AB, Boubli JP, Link A, Farias IP, Silva MN, Röhe F, Queiroz H, Chiou KL, Di Fiore A, Alfaro ME and Lynch-Alfaro JW (2017) Capuchin monkey biogeography: understanding *Sapajus* Pleistocene range expansion and the current sympatry between *Cebus* and *Sapajus*. *J Biogeogr* 44:810-820.
- Lenarducci ÂRI, Pinto-da-Rocha R and Lucas SM (2005) Descrição de uma nova espécie de *Rhopalurus* Thorell, 1876 (Scorpiones: Buthidae) do Nordeste brasileiro. *Biota Neotrop* 5:173-180.
- Lira AFA, Pordeus LM and de Albuquerque CMR (2017) A new species of *Ananteris* (Scorpiones: Buthidae) from Caatinga biome, Brazil. *Acta Arachnol* 66:9-15.
- Lira AFA, DeSouza AM and Albuquerque CMR (2018). Environmental variation and seasonal changes as determinants of the spatial distribution of scorpions (Arachnida: Scorpiones) in Neotropical forests. *Canad J Zool* 96:963-972.
- Lourenço WR (1986) Biogéographie et phylogénie des Scorpions du genre *Rhopalurus* Thorell, 1876 (Scorpiones, Buthidae). *Mém Soc r belge Ent* 33:129-137.
- Lourenço WR (1990) Caractéristiques biogéographiques de la Caatinga brésilienne. Associations avec le Chaco et d'autres formations végétales ouvertes de l'Amérique du Sud. L'exemple des Scorpions. *Compt Rendus Somm Séanc Soc Biogéograp* 66:149-169.
- Lourenço WR (1994) Biogeographic patterns of tropical South American scorpions. *Stud Neotrop Fauna Environ* 29:219-231.
- Lourenço WR, Wilmé L and Waeber PO (2016) More about the geographical pattern of distribution of the genus *Pseudouroplectes* Lourenço, 1995 (Scorpiones: Buthidae) from Madagascar. *Compt Rendus Biol* 339:37-43.
- Magalhães IL, Oliveira U, Santos FR, Vidigal TH, Brescovit AD and Santos AJ (2014) Strong spatial structure P liocene diversification and cryptic diversity in the Neotropical dry forest spider *Sicarius cariri*. *Mol Ecol* 23:5323-5336.
- Mainali KP, Warren DL, Dhileepan K, McConnachie A, Strathie L, Hassan G, Karki D, Shrestha BB and Parmesan C (2015) Projecting future expansion of invasive species: comparing and improving methodologies for species distribution modeling. *Glob Chang Biol* 21:4464-4480.

- Mares MA, Willig MR, Lacher Jr TE (1985) The Brazilian Caatinga in South American Zoogeography: Tropical Mammals in a Dry Region. *J Biogeogr* 12:57-69.
- Marini MÂ, Barbet-Massin M, Lopes LE and Jiguet F (2010) Predicting the occurrence of rare Brazilian birds with species distribution models. *J Ornithol* 151:857-866.
- Martins FM (2011) Historical biogeography of the Brazilian Atlantic forest and the Carnaval–Moritz model of Pleistocene refugia: what do phylogeographical studies tell us?. *Biol J Linn Soc* 104:499-509.
- McRae BH, Dickson BG, Keitt TH, and Shah VB (2008) Using circuit theory to model connectivity in ecology and conservation. *Ecology* 10:2712-2724.
- Mellert KH, Fensterer V, Küchenhoff H, Reger B, Kölling C, Klemmt HJ and Ewald J (2011) Hypothesis-driven species distribution models for tree species in the Bavarian Alps. *J Veget Sci* 22:635-646.
- Menezes MO, Zappi DC, Moraes EM, Franco FF, Taylor NP, Costa IR and Loiola MI (2016) Pleistocene radiation of coastal species of *Pilosocereus* (Cactaceae) in eastern Brazil. *J Arid Environ* 135:22-32.
- Miller AL, Makowsky RA, Formanowicz DR, Prendini L and Cox CL (2014) Cryptic genetic diversity and complex phylogeography of the boreal North American scorpion, *Paruroctonus boreus* (Vaejovidae). *Mol Phylogenetics Evol* 71:298-307.
- Miraldo A, Hewitt GM, Dear PH, Paulo OS and Emerson BC (2012) Numts help to reconstruct the demographic history of the ocellated lizard (*Lacerta lepida*) in a secondary contact zone. *Mol Ecol* 21:1005-1018.
- Miranda EA, Ferreira KM, Carvalho AT, Martins CF, Fernandes CR and Del Lama MA (2017) Pleistocene climate changes shaped the population structure of *Partamona seridoensis* (Apidae Meliponini) an endemic stingless bee from the Neotropical dry forest. *PloS one* 12: e0175725.
- Monod L, Harvey MS and Prendini L (2013) Stenotopic *Hormurus* Thorell, 1876 scorpions from the monsoon ecosystems of northern Australia, with a discussion on the evolution of burrowing behaviour in Hormuridae Laurie, 1896. *Rev Suisse Zool* 120:281-346.
- Moraes MR, Ríos-Uzeda B, Moreno LR, Huanca-Huarachi G and Larrea-Alcázar D (2014) Using potential distribution models for patterns of species richness,

- endemism, and phytogeography of palm species in Bolivia. *Trop Conserv Sci* 7:45-60.
- Moritz C, Patton JL, Schneider CJ and Smith TB (2000) Diversification of rainforest faunas: an integrated molecular approach. *Ann Rev Ecol Syst* 31:533-563.
- Moro MF, Silva IA, de Araujo FS, Lughadha EN, Meagher TR and Martins FR (2015) The role of edaphic environment and climate in structuring phylogenetic pattern in seasonally dry tropical plant communities. *PLoS One*, 10:e0119166.
- Moro MF, Lughadha EN, de Araújo FS and Martins FR (2016) A phytogeographical metaanalysis of the semiarid Caatinga domain in Brazil. *Bot Rev* 82:91-148.
- Morrone JJ (2000) A new regional biogeography of the Amazonian subregion, mainly based on animal taxa. *An Inst Biol Ser Zool* 71:99-123.
- Motta PE, Curi N and Franzmeier DP (2002) Relation of soils and geomorphic surfaces in the Brazilian Cerrado. In: Oliveira PS and Marquis RJ (eds) *The cerrados of Brazil: ecology and natural history of a neotropical savanna*. Columbia University Press, New York, pp 13-32.
- Nelson BW, Ferreira CA, da Silva MF and Kawasaki ML (1990) Endemism centres, refugia and botanical collection density in Brazilian Amazonia. *Nature* 345:714-716.
- Nordborg M (2007) Coalescent Theory. In Baldin DJ, Bishop M and Cannings C (eds) *Handbook of Statistical Genetics*. Wiley, England, pp 843-877.
- Nosil P (2008) Speciation with gene flow could be common: news and views. *Mol Ecol* 17:2103-2106.
- Ojanguren-Affilastro AA, Mattoni CI, Ochoa JA, Ramírez MJ, Ceccarelli FS, and Prendini L (2016) Phylogeny, species delimitation and convergence in the South American bothriurid scorpion genus *Brachistosternus* Pocock 1893: Integrating morphology, nuclear and mitochondrial DNA. *Mol Phylogenetics Evol* 94:159-170.
- Ojanguren-Affilastro AA, Adilardi RS, Mattoni CI, Ramírez MJ and Ceccarelli FS (2017) Dated phylogenetic studies of the southernmost American buthids (Scorpiones; Buthidae). *Mol Phylogenetics Evol* 110:39-49.
- Oke OA, Heard SB and Lundholm JT (2014) Integrating phylogenetic community structure with species distribution models: an example with plants of rock barrens. *Ecography* 37:614-625.

- Oliveira PE, Barreto AMF and Suguio K (1999) Late Pleistocene/Holocene climatic and vegetational history of the Brazilian caatinga: the fossil dunes of the middle São Francisco River. *Palaeogeogr Palaeoclimatol Palaeoecol* 152:319-337.
- Oliveira EF, Gehara M, São-Pedro VA, Chen X, Myers EA, Burbrink FT, Mesquita DO, Garda AA, Colli GR, Rodrigues MT, Arias FJ, Zaher H, Santos RML and Costa GC (2015). Speciation with gene flow in whiptail lizards from a Neotropical xeric biome. *Mol Ecol* 24:5957-5975.
- Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GV, Underwood EC, D'amico JA, Itoua I, Strand HE, Morrison JC, *et al.* (2001) Terrestrial Ecoregions of the World: A New Map of Life on Earth A new global map of terrestrial ecoregions provides an innovative tool for conserving biodiversity *BioScience* 51:933-938.
- Padonou EA, Tekla O, Bachmann Y, Schmidt M, Lykke AM and Sinsin B (2015) Using species distribution models to select species resistant to climate change for ecological restoration of bowé in West Africa. *Afr J Ecol* 53:83-92.
- Palumbi SR and Baker CS (1994) Contrasting population structure from nuclear intron sequences and mtDNA of humpback whales. *Mol Biol Evol* 11:426-435.
- Parida L (2010) Ancestral recombinations graph: a reconstructability perspective using random-graphs framework. *J Comp Biol* 17:1345-1370.
- Parida L (2012) Nonredundant Representation of Ancestral Recombinations Graphs. In Anisimova M (ed) *Evolutionary Genomics: Statistical and Computational Methods*. Springer, pp 315-332.
- Patel S, Weckstein JD, Patané JS, Bates JM and Aleixo A (2011) Temporal and spatial diversification of *Pteroglossus aracaris* (Aves: Ramphastidae) in the Neotropics: constant rate of diversification does not support an increase in radiation during the Pleistocene. *Mol Phylogenetics Evol* 58:105-115.
- Pennington RT, Prado DE and Pendry CA (2000) Neotropical seasonally dry forests and Quaternary vegetation changes. *J Biogeogr* 27:261-273.
- Pennington RT, Lavin M and Oliveira-Filho A (2009) Woody plant diversity, evolution, and ecology in the tropics: perspectives from seasonally dry tropical forests. *Annual Rev Ecol Evol Syst* 40:437-457.
- Peretolchina T, Pavan MG, Corrêa-Antônio J, Gurgel-Gonçalves R, Lima MM and Monteiro FA (2018) Phylogeography and demographic history of the Chagas

- disease vector *Rhodnius nasutus* (Hemiptera: Reduviidae) in the Brazilian Caatinga biome. PLoS Negl Trop Dis 12:e0006731.
- Pérez N and Font X (2012) Predicting vascular plant richness patterns in Catalonia (NE Spain) using species distribution models. Appl Veget Sci 15:390-400.
- Pessenda LCR, Gouveia SEM, de Souza Ribeiro A, De Oliveira PE and Aravena R (2010) Late Pleistocene and Holocene vegetation changes in northeastern Brazil determined from carbon isotopes and charcoal records in soils. Palaeogeogr Palaeoclimatol Palaeoecol 297:597-608.
- Piganeau G, Gardner M and Eyre-Walker A (2004) A broad survey of recombination in animal mitochondria. Mol Biol Evol 21:2319-2325.
- Pineda E and Lobo JM (2009) Assessing the accuracy of species distribution models to predict amphibian species richness patterns. J Anim Ecol 78:182-190.
- Pirie MD, Klitgaard BB and Pennington RT (2009) Revision and Biogeography of *Centrolobium* (Leguminosae-Papilionoideae). Syst Bot 34:345-359.
- Polis GA, McReynolds CN and Ford RG (1985) Home range geometry of the desert scorpion *Paruroctonus mesaensis*. Oecologia 67:273-277.
- Porfírio LL, Harris RM, Lefroy EC, Hugh S, Gould SF, Lee G, Bindoff NL and Mackey B (2014). Improving the use of species distribution models in conservation planning and management under climate change. PLoS One 9:e113749.
- Porto TJ, Carnaval AC and da Rocha PLB (2012) Evaluating forest refugial models using species distribution models, model filling and inclusion: a case study with 14 Brazilian species. Divers Distrib 19:330-340.
- Porto TJ, Carvalho LS, de Souza CAR, Oliveira U and Brescovit AD (2014) Escorpiões da Caatinga: conhecimentos atuais e desafios. In: Bravo F and Calor A (eds) Artrópodes do Semiárido Biodiversidade e Conservação. Printmídia, Feira de Santana, pp 33-46.
- Prado DE (2000) Seasonally dry forests of tropical South America: from forgotten ecosystems to a new phytogeographic unit. Edinb J Bot 57:437-461.
- Prado DE and Gibbs PE (1993) Patterns of species distributions in the dry seasonal forests of South America. Ann Mo Bot Gard 80:902-927.
- Prado CP, Haddad CF and Zamudio KR (2012) Cryptic lineages and Pleistocene population expansion in a Brazilian Cerrado frog. Mol Ecol 21:921-941.

- Prendini L, Esposito LA, Huff JC and Volschenk ES (2009) Redescription of *Rhopalurus abudi* (Scorpiones, Buthidae), with first description of the male and first record from mainland Hispaniola. *J Arachnol* 37:206-225.
- Queiroz LP, Cardoso D, Fernandes MF and Moro MF (2018) Diversity and Evolution of Flowering Plants of the Caatinga Domain. In da Silva JMC, Leal IR and Tabarelli M (eds) *Caatinga The Largest Tropical Dry Forest In South America*. Springer Nature, Switzerland, pp 23-65.
- Raes N, Roos MC, Slik JF, Van Loon EE and Steege HT (2009) Botanical richness and endemism patterns of Borneo derived from species distribution models. *Ecography* 32:180-192.
- Rahman MA, Gawin DF and Moritz C (2010) Patterns of genetic variation in the little spiderhunter (*Arachnothera longirostra*) in Southeast Asia. *Raffl Bull Zool* 58:381-390.
- Randin CF, Dirnböck T, Dullinger S, Zimmermann NE, Zappa M and Guisan A (2006) Are niche-based species distribution models transferable in space?. *J Biogeogr* 33:1689-1703.
- Richards CL, Carstens BC and Lacey-Knowles L (2007) Distribution modelling and statistical phylogeography: an integrative framework for generating and testing alternative biogeographical hypotheses. *J Biogeogr* 34:1833-1845.
- Rosenberg NA and Nordborg M (2002) Genealogical trees, coalescent theory and the analysis of genetic polymorphisms. *Nat Rev Gen* 3:e380.
- Rull V (2006) Quaternary speciation in the Neotropics. *Mol Ecol* 15:4257-4259.
- Rutschmann F (2006) Molecular dating of phylogenetic trees: a brief review of current methods that estimate divergence times. *Divers Distrib* 12:35-48.
- Santos AMM, Cavalcanti DR, Silva JMC, Tabarelli M (2007) Biogeographical Relationships among Tropical Forests in North-Eastern Brazil. *J Biogeogr* 34:437-446.
- Santos RM, Oliveira-Filho AT, Eisenlohr PV, Queiroz LP, Cardoso DB and Rodal MJ (2012) Identity and relationships of the Arboreal Caatinga among other floristic units of seasonally dry tropical forests (SDTFs) of north-eastern and Central Brazil. *Ecol Evol* 2:409-428.
- Santos-Da-Silva ADP, Carvalho LS and Brescovit AD (2017) Two new species of *Bothriurus* Peters, 1861 (Scorpiones, Bothriuridae) from Northeastern Brazil. *Zootaxa* 4258:238-256.

- Saraiva NEV and DaSilva MB (2016) Event-based biogeography of *Eusarcus dandara* sp. nov. (Opiliones: Gonyleptidae), an endemic species of the Northern Atlantic Rainforest of Brazil, and its closely related species. *Zootaxa* 4205:532-548.
- Short LL (1975) A zoogeographic analysis of the South American Chaco avifauna. *Bull Am Mus Nat Hist* 154: 235-252.
- Silva ACA, Bragg JG, Potter S, Fernandes C, Coelho MM and Moritz C (2017) Tropical specialist vs. climate generalist: diversification and demographic history of sister species of *Carlia* skinks from northwestern Australia. *Mol Ecol* 26:4045-4058.
- Silva JMC, Barbosa LCF, Leal IR and Tabarelli M (2018) The Caatinga: Understanding the Challenges. In da Silva JMC, Leal IR and Tabarelli M (eds) *Caatinga The Largest Tropical Dry Forest In South America*. Springer Nature, Switzerland, pp 3-23.
- Sinclair S, White M and Newell G (2010) How useful are species distribution models for managing biodiversity under future climates?. *Ecol Soc* 15:1-13.
- Sobral-Souza T, Lima-Ribeiro MS and Solferini VN (2015) Biogeography of Neotropical Rainforests: past connections between Amazon and Atlantic Forest detected by ecological niche modeling. *Evol Ecol* 29:643-655.
- Sousa P, Harris DJ, Froufe E and van der Meijden A. (2012) Phylogeographic patterns of *Buthus scorpions* (Scorpiones: Buthidae) in the Maghreb and South-Western Europe based on CO1 mtDNA sequences. *J Zool* 288:66-75.
- Sousa-Silva R, Alves P, Honrado J and Lomba A (2014) Improving the assessment and reporting on rare and endangered species through species distribution models. *Global Ecol Conserv* 2:226-237.
- Souza MCP, Moura F, Silva JV and Almeida C (2018) Phylogeography of the palm *Syagrus coronata* (Martius) Beccari (Arecaceae): distribution in the “Caatinga” and Atlantic forest domains. *Braz J Bot* 41:849-857.
- Spichiger R, Calenge C and Bise B. (2004) Geographical zonation in the Neotropics of tree species characteristic of the Paraguay-Paraná Basin. *J Biogeogr* 31:1489-1501.
- Štundlová J, Šmíd J, Nguyen P and Šťáhlavský F (2019) Cryptic diversity and dynamic chromosome evolution in Alpine scorpions (Euscorpiidae: *Euscorpius*). *Mol Phylogenetics Evol* 134:152-163.

- Sunday JM, Bates AE and Dulvy NK (2012) Thermal tolerance and the global redistribution of animals. *Nat Clim Chang* 2:e686.
- Syfert MM, Joppa L, Smith MJ, Coomes DA, Bachman SP and Brummitt NA (2014) Using species distribution models to inform IUCN Red List assessments. *Biol Conserv* 177:174-184.
- Tabarelli M, Vicente, A and Barbosa DCA (2003) Variation of seed dispersal spectrum of woody plants across a rainfall gradient in north-eastern Brazil. *J Arid Environm* 53:197-210.
- Templeton AR (2004) Statistical phylogeography: methods of evaluating and minimizing inference errors. *Mol Ecol* 13:789-809.
- Thomé MTC, Sequeira F, Brusquetti F, Carstens B, Haddad CF, Rodrigues MT and Alexandrino J (2016) Recurrent connections between Amazon and Atlantic forests shaped diversity in Caatinga four-eyed frogs. *J Biogeogr* 43:1045-1056.
- Thuiller W, Richardson DM, Pyšek P, Midgley GF, Hughes GO and Rouget M (2005) Niche-based modelling as a tool for predicting the risk of alien plant invasions at a global scale. *Glob Chang Biol* 11:2234-2250.
- Tricart J (1958) Division morphoclimatique du Brésil atlantique central. *Rev Géomorph Dynamique* 9:1-22.
- Tronstad LM, Brown KM and Andersen MD (2018) Using Species Distribution Models to Guide Field Surveys for an Apparently Rare Aquatic Beetle. *J Wild Manag* 9:330-339.
- Tsaousis AD, Martin DP, Ladoukakis ED, Posada D and Zouros E (2005) Widespread recombination in published animal mtDNA sequences. *Mol Biol Evol* 22:925-933.
- Tu Z, Liu M, Wang Y, Xu S, Song N, Gao T and Han Z (2016) The low mitochondrial diversities in lizardfish *Saurida elongate*: Recent population expansion and selection. *Biochem Syst Ecol* 68:44-50.
- Turchetto-Zolet AC, Pinheiro F, Salgueiro F and Palma-Silva C (2013) Phylogeographical patterns shed light on evolutionary process in South America. *Mol Ecol* 22:1193-1213.
- Vanzolini PE (1974) Ecological and geographical distribution of lizards in Pernambuco, northeastern Brazil (Sauria). *Pap Avulsos Zool* 28:61-90.

- Velloso AL, Sampaio EVSB and Pareyn FGC (2002) Ecorregiões propostas para o Bioma Caatinga. Associação Plantas do Nordeste; Instituto de Conservação Ambiental. The Nature Conservancy do Brasil, Recife, 80 pp.
- Vitt LJ (1991) An introduction to the ecology of Cerrado lizards. *J Herpetol* 25:79-90.
- Webb SD (1978) A history of savanna vertebrates in the New World. Part II: South America and the Great Interchange. *Ann Rev Ecol Syst* 9:393-426.
- Werneck FP (2011) The diversification of eastern South American open vegetation biomes: historical biogeography and perspectives. *Quat Sci Rev* 30:1630-1648.
- Werneck FP and Colli GR (2006) The lizard assemblage from Seasonally Dry Tropical Forest enclaves in the Cerrado biome, Brazil, and its association with the Pleistocenic Arc. *J Biogeogr* 33:1983-1992.
- Werneck FP, Costa GC, Colli GR, Prado DE and Sites Jr JW (2011) Revisiting the historical distribution of Seasonally Dry Tropical Forests: new insights based on palaeodistribution modelling and palynological evidence. *Global Ecol Biogeogr* 20:272-288.
- Werneck FP, Nogueira C, Colli GR, Sites JW and Costa GC (2012) Climatic stability in the Brazilian Cerrado: implications for biogeographical connections of South American savannas, species richness and conservation in a biodiversity hotspot. *J Biogeogr* 39:1695-1706.
- Werneck FP, Leite RN, Geurgas SR and Rodrigues MT (2015) Biogeographic history and cryptic diversity of saxicolous Tropiduridae lizards endemic to the semiarid Caatinga. *BMC Evol Biol* 2015:15-94.
- Williams JN, Seo C, Thorne J, Nelson JK, Erwin S, O'Brien JM and Schwartz MW (2009) Using species distribution models to predict new occurrences for rare plants. *Divers Distrib* 15:565-576.
- Wüster W, Ferguson JE, Quijada-Mascareñas JA, Pook CE, Graca-Salomao M and Thorpe RS (2005) Tracing an invasion: landbridges, refugia, and the phylogeography of the Neotropical rattlesnake (Serpentes: Viperidae: *Crotalus durissus*). *Mol Ecol* 14:1095-1108.
- Yannic G, Pellissier L, Ortego J, Lecomte N, Couturier S, Cuyler C, Dussalt C, Hundertmark KJ, Irvine J, Jenkins DA, Kolpashikov L, Mager K, Musiani M, Parker KL, Røed KH, Sipko T, Pórisson SG, Weckworth BV, Guisan A,

- Bernatchez L and Côté SD (2014). Genetic diversity in caribou linked to past and future climate change. *Nat Clim Chang* 4:132.
- Young KE, Abbott LB, Caldwell CA and Schrader TS (2013) Estimating suitable environments for invasive plant species across large landscapes: A remote sensing strategy using Landsat 7 ETM+. *Int J Biodivers Conserv* 5:122-134.
- Zink RM and Barrowclough GF (2008) Mitochondrial DNA under siege in avian phylogeography. *Mol Ecol* 17:2107-2121.

ANEXO I. NORMAS DE FORMATAÇÃO EXIGIDAS PELO PERIÓDICO

JOURNAL OF BIOGEOGRAPHY

1. SUBMISSION

Authors should kindly note that submission implies that the content has not been published or submitted for publication elsewhere except as a brief abstract in the proceedings of a scientific meeting or symposium. All submissions must be concisely and clearly written in grammatically correct English.

Once the submission materials have been prepared in accordance with the Author Guidelines, manuscripts should be submitted online at <https://mc.manuscriptcentral.com/jbi>

The submission system will prompt authors to use an ORCID iD (a unique author identifier) to help distinguish their work from that of other researchers. [Click here](#) to find out more.

[Click here](#) for more details on how to use [ScholarOne](#)

For help with submissions, please contact the Editorial Office at jbiooffice@wiley.com.

2. AIMS AND SCOPE

SCOPE:

The *Journal of Biogeography* publishes research at the intersection of biology and geography that is scientifically important and of broad general interest. We seek papers describing patterns and revealing mechanisms that shape biodiversity, through time, throughout the planet, from the deep past into the future, and from local to global scales. Diverse approaches are encouraged—including ecological, evolutionary, genomic, geographic, empirical, theoretical—considering any aspect of biogeography, from molecules to ecosystems and from microbes to plants and megafauna. Through this broad and inclusive scope, we aim for papers in *Journal of Biogeography* to address understudied, vexing, and urgent questions and to advance basic understanding of the origins, distributions, and fates of life on Earth.

Manuscripts submitted to *Journal of Biogeography* should be original and innovative, concise, well written, rigorously analyzed and argued, and consequential. While many such studies will be multifaceted, comparative, and draw generalities, we also welcome exceptional case studies that illustrate particularly interesting deviations that, in their aggregate, shift preconceptions.

The *Journal of Biogeography* is edited and reviewed for the community by a team of practising biogeographers. We support open data, accessibility to publish and read, and a constructive peer-review process.

VISION:

The *Journal of Biogeography* is the discipline's first and foremost journal. Its established history publishing influential papers in biogeography, its topical breadth, and its strong reputation in the community, provide the foundation for the journal to continue to grow as the most respected journal in the field. Nonetheless, as disciplinary and publishing trends change, to remain at the forefront of biogeography, the journal must innovate such that it represents not only core biogeography but also novel advances in emerging areas. Biogeography is an integrative discipline and the journal aims to increasingly complement its strong foundations with the most exciting multidisciplinary research.

3. MANUSCRIPT CATEGORIES AND REQUIREMENTS

The Journal publishes articles under the following main headers: 1) **Research Paper**, 2) **Methods and Tools**, 3) **Data**, 4) **Synthesis**, 5) **Perspective**, 6) **Commentary** and 7) **Correspondence**. All submissions are subject to peer review.

1) Research Paper. Research papers present new biogeographic research resulting from the analysis of a question in biogeography. For a typical Research paper, in which illustrative material (Tables and Figures) occupies about 3 pages of the journal when printed at final journal sizing, the text, inclusive of abstract and reference list, should not exceed 7000 words. Manuscripts should include a biosketch (see below); tables with their legends above; list of figure legends; and embedded figures, and the main headers in the main text of Research Papers should normally be Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References. Methods need to be described in a manner that allows a competent practitioner in the field to repeat the study. Authors must allow repeatability by either providing a thorough description of the methods or by providing relevant computer code.

Structured abstracts. Abstracts should be of no more than 300 words, presented as a series of factual statements under the following headings: Aim, Location, Taxon, Methods, Results and Main conclusions. The Aim should give a clear statement of the principal research question(s) or hypotheses, the Taxon indicate the main group (eg angiosperms), the Methods should give details of materials/sampling/methods of analysis, and the Main conclusions should give the main take-home message.

Biosketch/Biosketches. A short Biosketch/Biosketches entry (30-100 words for one author/150 words total for the first three authors, respectively) describing the research interests of the author(s) should be provided. For papers with four or more authors, biosketch details should be supplied for the first author only and/or a general statement of the focus of the research team (which may include a link to a group web page) plus, in all cases, a statement of author contributions, e.g. Author contributions: A.S. and K.J. conceived the ideas; K.J. and R.L.M. collected the data; R.L.M. and P.A.K. analysed the data; and A.S. and K.J. led the writing.

For an example click [here](#).

2) Methods and Tools. These are structured as in Research papers, but the main focus is to present or investigate a new method, rather than to explore a biogeographical problem. Papers in this section are expected to apply new methods to the analysis of biogeographic data and discuss the potential of those methods for advancing the study of the field. For an example [click here](#).

3) Data. Datasets that are likely to be of interest to the broader biogeography community are published under this category. Data papers allow scientists to publish and receive credit for work in which the nature of the collected, mobilized, or integrated data more than a specific analysis may be most impactful. The structure of a Data paper should be similar to that of a Research paper. Data papers must include a characterization of overall scope (e.g. organismal, spatial), a description of how the data were collected (protocols), a detailed characterization of all data fields and metadata, information on data records (e.g. SI or in a suitable repository), a section on technical validation, and usage notes addressing potential caveats for analysis and interpretation. Additional analyses that exemplify potential uses of the data are encouraged but not essential. Note that the Data papers category is intended for novel datasets - data used in a published or submitted research paper should be fully addressed and made available there.

4) Synthesis. Papers that have the character of a theoretical synthesis or review, even if incorporating an element of original analysis within them, should use the article type Synthesis. Guidelines are as for Research papers but submissions to the Synthesis section may be of up to 10,000 words, or exceptionally more, if the additional length is fully justified. Authors of synthesis papers are encouraged to discuss their planned paper with one of the Chief Editors, especially if the length will exceed 10,000 words. For an example click [here](#).

5) Perspective. Perspectives papers are should be stimulating and reflective essays providing personal perspectives on key research fields and issues within biogeography. When published, Perspectives should be of no more than eight printed pages (main text maximum 5000 words; word count including abstract, main text and references 7000 words maximum but note that shorter articles are encouraged), and they should include a short, single-paragraph abstract. A biosketch (see below) may be included after the references providing the overall paper length limit is not exceeded. For an example click [here](#).

6) Commentary. Commentary submissions should provide readily intelligible comment on the latest original research in biogeography. The prose style should be light, and the article should be written with the minimum of technical language and jargon, so as to be understandable to a general audience or an undergraduate taking an introductory course in biogeography. Contributions will be subject to rapid peer review. Commentaries should occupy a maximum of two pages of the journal, and should have a maximum of 10 references: thus the overall word count should not exceed 1600. No biosketch is included for commentaries. Should you wish to include a small figure or other illustration, this can be accommodated by a reduction in the number of words on a pro rata basis. For an example click [here](#).

7) Correspondence. The Journal welcomes short items of correspondence prompted by papers previously published in this or occasionally in other journals. The text should not normally exceed 2500 words, inclusive of a short one-paragraph abstract (up to 150 words), and a list of 6–10 keywords. No biosketch is necessary for Correspondence papers. For an example click [here](#).

4. PREPARING THE SUBMISSION

Article Preparation Support

[Wiley Editing Services](#) offers expert help with English Language Editing, as well as translation, manuscript formatting, figure illustration, figure formatting, and graphical abstract design – so you can submit your manuscript with confidence. Also, check out our resources for [Preparing Your Article](#) for general guidance about writing and preparing your manuscript.

Cover Letters

A cover letter to the editor, indicating in less than 100 words why this paper is of interest to the readers of the Journal, must be uploaded separately.

Parts of the Manuscript

The manuscript should be submitted in separate files: main text file with embedded figures; supporting information.

LaTeX users do not have to translate their manuscripts into MSWord, but may upload them as PDF files. Any explanatory notes, companion papers etc. for the attention of reviewers should be uploaded under 'Comments to reviewers'.

Main Text File

The text file should be single spaced, or 1.3 spaced, and presented in the following order:

- i. Title
- ii. A short running title of less than 40 characters
- iii. The full names of the authors, only 1 corresponding author may be included
- iv. The author's institutional affiliations where the work was carried out, with a footnote for the author's present address if different from where the work was carried out
- v. Acknowledgements
- vi. Abstract and keywords
- vii. Main text
- viii. Tables embedded in the text (each table complete with title and footnotes)
- ix. Figures embedded in the text, each with a figure legend
- x. Data availability statement
- xi. References
- xii. Biosketch
- xiii. Appendices (if relevant)
- xiv. Supporting information should be supplied as separate files.

Title. The title should be short and informative, containing major keywords related to the content. The title should not contain abbreviations (see [Wiley's best practice SEO tips](#)).

Authorship. For details on eligibility for author listing, please refer to the journal's Authorship policy outlined in the [Editorial Policies and Ethical Considerations](#) section. Only 1 corresponding author may be included.

Acknowledgements. Contributions from individuals who do not meet the criteria for authorship should be listed, with permission from the contributor, in an Acknowledgements section. Financial and material support should also be mentioned. Thanks to anonymous reviewers are not appropriate.

Conflict of Interest Statement. Authors will be asked to provide a conflict of interest statement during the submission process. See 'Conflict of Interest' section in [Editorial Policies and Ethical Considerations](#) for details on what to include in this section. Authors should ensure they liaise with all co-authors to confirm agreement with the final statement.

Abstract and Keywords

Abstracts and keywords are required for some manuscript types. For details on manuscript types that require abstracts and/or keywords, as well as how to prepare them, please refer to the 'Manuscript Categories and Requirements' section. Please provide 6-10 keywords, arranged alphabetically, separated by commas. Note that optimally the most important keywords are repeated in the title and the keywords.

Main Text

The journal uses British spelling; however, authors may submit using either option, as spelling of accepted papers is converted during the production process.

References

References are styled according to the sixth edition of the Publication Manual of the American Psychological Association. List all sources in the reference alphabetically by name.

In text citations should follow the author-date method. This means that the author's last name and the year of publication for the source should appear in the text, for example, (Jones, 1998), and a complete reference should appear in the reference list at the end of the paper.

When a work has two authors, cite both names every time the reference occurs in text. When a work has three, four, or five authors, cite all authors the first time the reference occurs; subsequent citations include only the surname of the first author followed by et al., (not italicized and with a period after "al.") and the year if it is the first citation of the reference within a paragraph.

If there are two or more citations that shorten to the same lead author and date, give as many additional names as needed to identify them, e.g., (Smith, Jones, et al., 1991) and (Smith, Burke, et al., 1991).

Unpublished data, works in preparation and papers submitted but not yet accepted may be cited in the text as personal communication, giving the author's initials and surname, but should not be included in the reference list. It is the author's responsibility to obtain permission from colleagues to include their work as a personal communication. Please add the person's initials, surname and if applicable institute for personal communications.

The basic reference form for a journal paper is: Author (date). Paper title. Journal, Volume, page; and for a book citation: Author (date). Book title. Place of publication, publisher.

Please note that for journal articles, issue numbers are not included unless each issue in the volume begins with page one. Journals names are written out in full.

Please ensure that in the paper titles only proper names are capitalized, and that all scientific binomials are in italics.

Please include up to seven authors in the list (use "&" before last author name). For eight or more authors please list the first six and then use ellipses followed by last author (do not use "&" before last author name)

Journal article:

Light, M. A., & Light, I. H. (2008). The geographic expansion of Mexican immigration in the United States and its implications for local law enforcement. *Law Enforcement Executive Forum Journal*, 8(1), 73–82.

Book:

Goldstein, H. (1990). *Problem-oriented policing*. New York, NY: McGraw-Hill. Miles, M. B., & Huberman, A. M. (1994). *Qualitative data analysis* (2nd ed.). Thousand Oaks, CA: Sage.

Edited Book:

Gilbert, D. G., McClernon, J. F., Rabinovich, N. E., Sugai, C., Plath, L. C., Asgaard, G., ... Botros, N. (1983). Situational crime prevention: Its theoretical basis and practical scope. In M. Tonry & N. Morris (Eds.), *Crime and justice: An annual review of research* (Vol. 4, pp. 225–256). Chicago, IL: University of Chicago Press.

Citations to data sources

Some studies (e.g., meta-analyses) use data drawn from multiple published sources. If these sources are not otherwise cited in the main text, they should be listed in one or more appendices with titles similar to the following: "Appendix 1 – Data sources". These data appendices will be printed in the main paper (so that citation indexing services will capture them), but in a reduced font. These appendices should be cited in the main text (e.g. "A list of the data sources is found in Appendix 1."), and be placed after the biosketch in the manuscript.

Tables

Tables should be self-contained and complement, not duplicate, information contained in the text. They should be supplied as editable files, not pasted as images. Legends should be concise but comprehensive – the table, legend, and footnotes must be understandable without reference to the text, giving the study organism and study location and 'n' values where applicable. Column headings should be brief, with units of measurement in parentheses. All abbreviations must be defined in footnotes.

Figure Legends

Legends should be concise but comprehensive – the figure and its legend must be understandable without reference to the text, to this end both the geographical region and the taxon should be mentioned in each caption. Include definitions of any symbols used and define/explain all abbreviations and units of measurement.

Figures

For review purposes, figures should be embedded in the text file. All illustrations (including photographs and maps) are classified as figures and they should be numbered consecutively as first cited in the text. Panels should be labelled (a), (b), (c), etc. rather than (A), (B), (C) etc. and referred to in the text as, for example, Fig. 1a. Figure legends should be listed at the end of the paper before the embedded figures. Legends should be explicit and informative and should 'stand alone' from the main text, giving the study organism and study location where applicable. All abbreviations should be defined.

[Click here](#) for the basic figure requirements for figures submitted with manuscripts for initial peer review, as well as the more detailed post-acceptance figure requirements.

If and when your paper is accepted for publication, the editorial office will request you to upload your figures as separate files in the format(s) specified below. When supplying these files, use the following naming convention: manuscript number, figure number and then the appropriate file extension e.g. 'JBI-08-0500_Fig1.tif'.

Photographic figures should be saved in .tif format at 300 d.p.i. (or failing that in .jpg format with low compression). Line figures should be saved as vector graphics (i.e. composed of lines, curves, points and fonts) in .eps or .pdf format, as this enhances their display when published online. Combination figures (those composed of vector and pixel/raster elements) should also be saved in .eps or .pdf format where possible. If line figures and combination figures cannot be saved in vector graphics format, they should be saved in .tif format at high resolution (i.e. 600–800 d.p.i.) (do not save them in .jpg format). If you are unsure about the resolution of your .tif files, please zoom in and check that fonts, curves and diagonal lines are smooth-edged and do not appear blocky. Note that .tif files are downsampled for online publication and so authors should preferentially opt for vector graphic formats for line and combination figures (full resolution .tif files are used for print publication). Colour figures should be saved in CYMK rather than RGB.

Prepare figures such that, after reduction to print size, all lettering and symbols will be clear and easily read, and such that each figure makes effective use of space. Font size in figures should be 8 pt. To check this, fix the image size in Illustrator to the required column width, and check the font size. Possible figure sizes: single column = 79mm, 2/3rd column = 110mm, double column = 168mm, maximum height of figure = 230mm.

Bar scales for maps and photographs are preferred to numerical scales and must be given on all such items. Maps that display area data and organism distribution at a continental, hemispheric, or world scale must always use an equal-area map projection (e.g. Mollweide or Aitoff's). Note especially that Mercator's projection is not acceptable for such data. Please indicate the precise projection employed in the caption. On these maps, the equatorial scale should be indicated, while scale information should be provided, preferably as a scale bar within the figure, for all maps of whatever size and area; use 'km' or 'kilometres', not 'kilometers'. Maps should include adequate geo-referencing information (preferably the latitude and longitude).

Additional Files

Supporting Information

Supporting information is information that is not essential to the article, but provides greater depth and background. It is hosted online and appears without editing or typesetting. It may include tables, figures, videos, datasets, etc. [Click here](#) for Wiley's FAQs on supporting information.

Note: if data, scripts, or other artefacts used to generate the analyses presented in the paper are available via a publicly available data repository, authors should include a reference to the location of the material within their paper.

Such supporting information should be referred to in the text as, for example, 'see Appendix S1 in Supporting Information'; subsequent mention should be in the form 'see Appendix S2'. Figures and tables in the Supporting Information must be numbered consecutively by Appendix number and figure number: e.g. the first figure in Appendix 1 as Fig. S1.1, the first in Appendix 2 as Fig. S2.2 (if there is only one figure in Appendix 1). All appendices, figures and tables must be cited in the text.

Supporting Information files are hosted by the Publisher in the format supplied by the author and are not copy-edited by the Publisher. **It is the responsibility of the author to supply Supporting Information in an appropriate file format and to ensure that it is accurate and correct.** Authors should therefore prepare Supporting Information with the same rigour as their main paper, including adherence to journal style (e.g. formatting of references, figure captions, headings). Sources cited only in the Supporting Information should be listed in a reference section within the supplementary files and not with the main paper. Supporting Information can be provided as separate editable files or, preferably, as one combined file. Authors are discouraged from supplying very large files or files in non-standard file formats, both of which may reduce their use to the readership. At the point a paper is accepted, these files should be prepared without line numbers or wide line spacing, and with all track-change edits accepted.

General Style Points

The following points provide general advice on formatting and style.

- **Abbreviations:** In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Initially, use the word in full, followed by the abbreviation in parentheses. Thereafter use the abbreviation only.
- **Units of measurement:** Measurements should be given in SI or SI-derived units. Visit the Bureau International des Poids et Mesures (BIPM) website at www.bipm.fr for more information about SI units.
- **Numbers:** numbers under 10 are spelt out, except for: measurements with a unit (8mmol/l); age (6 weeks old), or lists with other numbers (11 dogs, 9 cats, 4 gerbils).
- **Computer programs:** All software programs should be written in small caps, first written in roman (e.g. MrBayes or BEAST) and then saved as small caps, followed at first mention by the version number and reference. Packages in R should in roman and quotations (e.g. 'vegan') and the relevant reference provided.

Wiley Author Resources

Manuscript Preparation Tips: Wiley has a range of resources for authors preparing manuscripts for submission available [here](#). In particular, authors may benefit from referring to Wiley's best practice tips on [Writing for Search Engine Optimization](#).

Editing, Translation, and Formatting Support: [Wiley Editing Services](#) can greatly improve the chances of a manuscript being accepted. Offering expert help in English language editing, translation, manuscript formatting, and figure preparation, Wiley Editing Services ensures that the manuscript is ready for submission.

Guidelines for Cover Image Submissions: If you would like to send suggestions for artwork related to your manuscript to be considered to appear on the cover of the journal, [please follow these general guidelines](#).

5. EDITORIAL POLICIES AND ETHICAL CONSIDERATIONS

Editorial Review and Acceptance

The acceptance criteria for all papers are the quality and originality of the research and its significance to journal readership. Papers will only be sent to review if the Editor-in-Chief determines that the paper meets the appropriate quality and relevance requirements.

Wiley's policy on confidentiality of the review process is [available here](#).

Referrals to the Open Access Journal "Ecology and Evolution" and "Geo: Geography and Environment"

This Journal works together with Wiley's Open Access journals, [Ecology and Evolution](#) and [Geo: Geography and Environment](#), to enable rapid publication of good quality research that we are unable to accept for publication. Authors may be offered the option of having their paper, along with any related reviews, automatically transferred for consideration by the Editors of *Ecology and Evolution* or *Geo: Geography and Environment*. Authors will not need to reformat or rewrite their manuscript at this stage, and publication decisions will be made a short time after the transfer takes place. The Editors of *Ecology and Evolution* and *Geo: Geography and Environment* will accept submissions that report well-conducted research and which reach the standard acceptable for publication. Accepted papers can be published rapidly, typically within 15 days of acceptance. *Ecology and Evolution* and *Geo: Geography and Environment* are Wiley Open Access journals and article publication fees apply. More information can be found [here](#). Occasionally we refer papers to our sister journals DDI or GEB.

Data Storage and Documentation

Journal of Biogeography supports open research, therefore as a condition for publication, requires that the data supporting the results in the paper will be archived in an appropriate public repository, such as Dryad, TreeBASE, NERC data centre, GenBank, figshare or another archive of the author's choice that provides comparable access and guarantee of preservation. Authors are required to provide a data availability statement, including a link to the repository they have used, and to cite the data they have shared. Whenever possible the scripts and other artefacts used to generate the analyses presented in the paper should also be publicly archived in a repository. Exceptions may be granted at the discretion of the editor. If authors are unable to share data (for example, if sharing data compromises ethical standards or legal requirements) then authors are not required to share it and must describe restrictions in their data availability statement.

Data Sharing with Dryad

Journal of Biogeography have partnered with Dryad to enable authors to store and share their data without charge. The cost of depositing data of up to 20GB will be covered, should authors choose Dryad as their preferred public repository, upon acceptance of an article in *Journal of Biogeography*. For more details, please see the Dryad webpage [here](#).

Preprints

The *Journal of Biogeography* will consider for review articles previously available as preprints on non-commercial servers. Authors may also post the submitted version of a manuscript to non-commercial servers at any time. Authors are requested to update any pre-publication versions with a link to the final published article.

Sequence Data

Sequence data have to be submitted in electronic form to any one of the three major collaborative databases: DDBJ, EMBL, or GenBank. The suggested wording for referring to accession-number information is: 'These sequence data have been submitted to the DDBJ/EMBL/GenBank databases under accession number U12345'. Addresses are as follows:

- DNA Data Bank of Japan (DDBJ) www.ddbj.nig.ac.jp
- EMBL Nucleotide Archive: ebi.ac.uk/ena
- GenBank www.ncbi.nlm.nih.gov/genbank

Collecting permission and the Nagoya Protocol

Authors must ensure that any data utilised in the submitted manuscript have been lawfully acquired in accordance with The Nagoya Protocol on Access to Genetic Resources and the Fair and Equitable Sharing of Benefits Arising from Their Utilization to the Convention on Biological Diversity. It is recommended that it is explicitly stated that the relevant fieldwork permission was obtained, and to list the permit numbers, in Materials and Methods or the Acknowledgements.

Species Names

Upon its first use in the title, abstract, and text, the common name of a species should be followed by the scientific name (genus, species) in parentheses. For well-known species, however, scientific names may be omitted from article titles. If no common name exists in English, only the scientific name should be used. For the focal species in the study, the authority(ies) should be provided at the first mention in the main text, in the format specified by the relevant code.

Conflict of Interest

The journal requires that all authors disclose any potential sources of conflict of interest. Any interest or relationship, financial or otherwise that might be perceived as influencing an author's objectivity is considered a potential source of conflict of interest. These must be disclosed when directly relevant or directly related to the work that the authors describe in their manuscript. Potential sources of conflict of interest include, but are not limited to: patent or stock ownership, membership of a company board of directors, membership of an advisory board or committee for a company, and consultancy for or receipt of speaker's fees from a company. The existence of a conflict of interest does not preclude publication. If the authors have no conflict of interest to declare, they must also state this at submission. It is the responsibility of the corresponding author to review this policy with all authors and collectively to disclose with the submission ALL pertinent commercial and other relationships.

Funding

Authors should list all funding sources in the Acknowledgements section. Authors are responsible for the accuracy of their funder designation. If in doubt, please check the Open Funder Registry for the correct nomenclature: <https://www.crossref.org/services/funder-registry/>

Authorship

The list of authors should accurately illustrate who contributed to the work and how. All those listed as authors should qualify for authorship according to all of the following criteria:

1. Have made substantial contributions to conception and design, or acquisition of data, or analysis and interpretation of data;
2. Been involved in drafting the manuscript or revising it critically for important intellectual content;
3. Given final approval of the version to be published. Each author should have participated sufficiently in the work to take public responsibility for appropriate portions of the content; and

4. Agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Contributions from anyone who does not meet the criteria for authorship should be listed, with permission from the contributor, in an Acknowledgements section (for example, to recognize contributions from people who provided technical help, collation of data, writing assistance, acquisition of funding, or a department chairperson who provided general support). Prior to submitting the article all authors should agree on the order in which their names will be listed in the manuscript.

Additional Authorship Options: Joint first or senior authorship: In the case of joint first authorship, a footnote should be added to the author listing, e.g. 'X and Y should be considered joint first author' or 'X and Y should be considered joint senior author.'

ORCID

As part of the journal's commitment to supporting authors at every step of the publishing process, the journal requires the submitting author (only) to provide an ORCID iD when submitting a manuscript. This takes around 2 minutes to complete. [Find more information here](#).

Publication Ethics

This journal is a member of the [Committee on Publication Ethics \(COPE\)](#). Note this journal uses iThenticate's CrossCheck software to detect instances of overlapping and similar text in submitted manuscripts. Read the Top 10 Publishing Ethics Tips for Authors [here](#). Wiley's Publication Ethics Guidelines can be found at authorservices.wiley.com/ethics-guidelines/index.html.

6. AUTHOR LICENSING

If a paper is accepted for publication, the author identified as the formal corresponding author will receive an email prompting them to log in to Author Services, where via the Wiley Author Licensing Service (WALS) they will be required to complete a copyright license agreement on behalf of all authors of the paper.

Authors may choose to publish under the terms of the journal's standard copyright agreement, or [OnlineOpen](#) under the terms of a Creative Commons License.

General information regarding licensing and copyright is available [here](#). To review the Creative Commons License options offered under OnlineOpen, please [click here](#). (Note that certain funders mandate a particular type of CC license be used; to check this please click [here](#).)

Self-Archiving Definitions and Policies: Note that the journal's standard copyright agreement allows for self-archiving of different versions of the article under specific conditions. Please click [here](#) for more detailed information about self-archiving definitions and policies.

Open Access fees: Authors who choose to publish using OnlineOpen will be charged a fee. A list of Article Publication Charges for Wiley journals is available [here](#).

Funder Open Access: Please click [here](#) for more information on Wiley's compliance with specific Funder Open Access Policies.

7. PUBLICATION PROCESS AFTER ACCEPTANCE

Accepted Article Received in Production

When an accepted article is received by Wiley's production team, the corresponding author will receive an email asking them to login or register with [Wiley Author Services](#). The author will be asked to sign a publication license at this point.

Proofs

Once the paper is typeset, the author will receive an email notification with the URL to download a PDF typeset page proof, as well as associated forms and full instructions on how to correct and return the file.

Please note that the author is responsible for all statements made in their work, including changes made during the editorial process – authors should check proofs carefully. Note that proofs should be returned within 48 hours from receipt of first proof.

At proof correction stage authors will be given access to their Supporting Information (via the web) and should check it for accuracy and updates. If changes are required, corrected versions of the files that were received with the proof must be emailed to the Production Editor, with a brief description of the changes made. Supporting Information **must be checked alongside the main proof** and corrections for both returned to the Production Editor at the same time.

Social Media Promotion

To share your new publication on social media, information and an image for tweets, facebook, and instagram will be invited immediately prior to publication.

Publication Charges

Colour figures may be published online free of charge; however, the journal charges for publishing figures in colour in print. If the author supplies colour figures, they will be sent a Colour Work Agreement once the accepted paper moves to the production process. If the Colour Work Agreement is not returned by the specified date, figures will be converted to black and white for print publication.

Please note that the vast majority of readers access the digital versions of the journal; printed copies are increasingly rare. For the convenience of readers, we ask that you design your colour artwork so that it can be understood as best as possible in greyscale. Note that the same figure file must be used for both the print and online versions (we do not accept differing colour and black-and-white versions of the same figure).

Early View

The journal offers rapid publication via Wiley's Early View service. [Early View](#) (Online Version of Record) articles are published on Wiley Online Library before inclusion in an issue. Note there may be a delay after corrections are received before the article appears online, as Editors also need to review proofs. Once the article is published on Early View, no further changes to the article are possible. The Early View article is fully citable and carries an online publication date and DOI for citations.

8. DATA PROTECTION

By submitting a manuscript to or reviewing for this publication, your name, email address, and affiliation, and other contact details the publication might require, will be used for the regular operations of the publication, including, when necessary, sharing with the publisher (Wiley) and partners for production and publication. The publication and the publisher recognize the importance of protecting the personal information collected from users in the operation of these services, and have practices in place to ensure that steps are taken to maintain the security, integrity, and privacy of the personal data collected and processed. You can learn more at <https://authorservices.wiley.com/statements/data-protection-policy.html>.

9. POST PUBLICATION

Article Promotion Support

[Wiley Editing Services](#) offers professional video, design, and writing services to create shareable video abstracts, infographics, conference posters, lay summaries, and research news stories for your research – so you can help your research get the attention it deserves.

Access and Sharing

Please review Wiley's guidelines on sharing your research [here](#).

When the article is published online:

- The author receives an email alert (if requested).
- The link to the published article can be shared through social media.
- The author will have free access to the paper (after accepting the Terms & Conditions of use, they can view the article).
- The corresponding author and co-authors can nominate up to ten colleagues to receive a publication alert and free online access to the article.

Print copies of the article can now be ordered (instructions are sent at proofing stage or use the below contact details). Email www.sheridan.com/wiley/eoc

To find out how to best promote an article, click [here](#).

Measuring the Impact of an Article

Wiley also helps authors measure the impact of their research through specialist partnerships with [Kudos](#) and [Altmetric](#).

10. EDITORIAL OFFICE CONTACT DETAILS

jbiooffice@wiley.com

Author Guidelines updated December 2019

CURRICULUM VITAE (LATTES)

Stênio Ítalo Araújo Foerster

Endereço para acessar este CV: <http://lattes.cnpq.br/0589282540409945>

Última atualização do currículo em 18/03/2020

Resumo informado pelo autor

Bacharel em Ciências Biológicas pela Universidade Federal Rural de Pernambuco (2015), e discente do Programa de Pós-Graduação em Genética da Universidade Federal de Pernambuco (2018). Atua nas seguintes temáticas: Filogeografia, Ecologia e Biogeografia, utilizando escorpiões neotropicais como organismos modelo para elucidar padrões filogeográficos e macroecológicos que podem auxiliar na identificação de refúgios e/ou centros de diversificação, bem como na compreensão dos mecanismos de formação e distribuição de comunidades biológicas em ambientes de Caatinga, Floresta Atlântica e enclaves de floresta úmida situados dentro do bioma Caatinga (Brejos de Altitude). **(Texto informado pelo autor).**

Formação acadêmica/titulação

2018 Mestrado em Programa de Pós-Graduação em Genética.
Universidade Federal de Pernambuco, UFPE, Recife, Brasil
Título: Diversidade genética e padrões filogeográficos do escorpião *Jaguajir rochae* (Borelli, 1910) em um gradiente de estabilidade climática da Caatinga

Orientador: Valdir de Queiroz Balbino



Bolsista do(a): Conselho Nacional de Desenvolvimento Científico e Tecnológico

2010 - 2016 Graduação em Bacharelado em Ciências Biológicas.
Universidade Federal Rural de Pernambuco, Unidade Acadêmica de Serra Talhada, UFRPE/UAST, Brasil
Título: Escorpiões do Parque Estadual Mata da Pimenteira e o escorpionismo no município de Serra Talhada, Pernambuco
Orientador: Cauê Guion de Almeida

Formação complementar

2016 - 2016 Formação Inicial e Continuada para Agente de Combate às Endemias. . (Carga horária: 40h).
Prefeitura Municipal de Afogados da Ingazeira, IPMAI, Afogados Da Ingazeira, Brasil

2016 - 2016 Curso de curta duração em Ecologia dos Crocodilianos. (Carga horária: 5h).
Universidade de Pernambuco, UPE, Recife, Brasil

2013 - 2015 Monitor Bolsista das Disciplinas Bioquímica I e II. (Carga horária: 1170h).
Universidade Federal Rural de Pernambuco, Unidade Acadêmica de Serra Talhada, UFRPE/UAST, Brasil
Bolsista do(a): Pró-Reitoria de Gestão Estudantil (UFRPE)

2013 - 2013 Curso de curta duração em Biologia, Diversidade e Taxonomia de Aranhas. (Carga horária: 8h).
Universidade Federal de Alagoas, UFAL, Maceio, Brasil

2013 - 2013 Extensão universitária em Animais Peçonhentos. (Carga horária: 15h).
Universidade do Estado do Rio de Janeiro, UERJ, Rio De Janeiro, Brasil

2013 - 2013 Curso de curta duração em Filogenia de Aranhas e Escorpiões no Brasil. (Carga horária: 8h).
Universidade Federal de Pernambuco, UFPE, Recife, Brasil

- 2012 - 2012** Curso de curta duração em Sequenciamento e Análise de Fragmento. (Carga horária: 4h). Universidade Federal do Vale do São Francisco, UNIVASF, Petrolina, Brasil
- 2012 - 2012** Curso de curta duração em Análise da Expressão Gênica pela Técnica de PCR. (Carga horária: 4h). Universidade Federal do Vale do São Francisco, UNIVASF, Petrolina, Brasil
- 2011 - 2011** Curso de curta duração em Técnicas para o Estudo em Etologia. (Carga horária: 9h). Universidade Federal Rural de Pernambuco, Unidade Acadêmica de Serra Talhada, UFRPE/UAST, Brasil
- 2011 - 2011** Extensão universitária em IV Curso de Atualização em Animais Peçonhentos. (Carga horária: 20h). Universidade Federal de Pernambuco, UFPE, Recife, Brasil
- 2010 - 2010** Curso de curta duração em Técnicas de Mostragem de Répteis e Mamíferos. (Carga horária: 10h). Universidade Federal Rural de Pernambuco, Unidade Acadêmica de Serra Talhada, UFRPE/UAST, Brasil

Revisor de periódico

1. Studies on Neotropical Fauna and Environment -

Vínculo

2019 - Atual Regime: Parcial

2. Journal of Mountain Science -

Vínculo

2018 - Atual Regime: Parcial

Áreas de atuação

1. Filogeografia
2. Macroecologia
3. Biogeografia
4. Genética de Populações

Idiomas

Inglês	Compreende Razoavelmente, Fala Razoavelmente, Escreve Razoavelmente, Lê Razoavelmente
Espanhol	Compreende Razoavelmente, Fala Razoavelmente, Escreve Razoavelmente, Lê Razoavelmente
Francês	Compreende Razoavelmente, Fala Razoavelmente, Escreve Pouco, Lê Razoavelmente
Português	Compreende Bem, Fala Bem, Escreve Bem, Lê Bem

Produção

Produção bibliográfica

Artigos completos publicados em periódicos

1.  **FOERSTER, S.I.A.**; DESOUZA, A.M.; LIRA, A.F.A.
Macroecological approach for scorpions (Arachnida, Scorpiones): β -diversity in Brazilian montane forests. CANADIAN JOURNAL OF ZOOLOGY (ONLINE). , v.97, p.914 - 921, 2019.

Trabalhos publicados em anais de eventos (resumo)

1. **FOERSTER, S.I.A.**; PEREIRA, B. N. S.; LIRA, A. F. A.; da SILVA, W. D.; BALBINO, V. Q.
Validação de método de extração de DNA com Chelex 100 resin e amplificação da região COI (mtDNA) em duas espécies de escorpiões In: VIII Jornada de Pós-Graduação de Genética UFPE, 2018, Recife.
Anais da VIII Jornada de Pós-Graduação de Genética UFPE. , 2018.

Trabalhos publicados em anais de eventos (resumo expandido)

1. **FOERSTER, S.I.A.**; PEREIRA, B. N. S.; LIRA, A. F. A.; BALBINO, V. Q.
A estabilidade climática da Caatinga pode prever padrões de identidade genética? Um estudo com o escorpião *Jaguajir rochae* (Borelli, 1910) In: XX Encontro de Zoologia do Nordeste, 2019, Maceió.
Anais do XX Encontro de Zoologia do Nordeste. Maceió (AL): , 2019.
2. **FOERSTER, S.I.A.**; LIRA, A. F. A.; SILVA, V. G.; ALMEIDA, C. G.
Complexidade ambiental como fator determinante sobre a composição de comunidades de escorpiões em áreas de Caatinga In: XX Encontro de Zoologia do Nordeste, 2019, Maceió.
Anais do XX Encontro de Zoologia do Nordeste. Maceió (AL), 2019.

Página gerada pelo sistema Currículo Lattes em 18/03/2020 às 00:19:48.