

UNIVERSIDADE FEDERAL DE PERNAMBUCO  
CENTRO DE CIÊNCIAS BIOLÓGICAS  
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOQUÍMICA E FISIOLOGIA

DISSERTAÇÃO DE MESTRADO

EXPOSIÇÃO À HIPERGLICEMIA MATERNA INDUZ DISFUNÇÃO ENDOTELIAL  
EM ARTÉRIAS DE RESISTÊNCIA DA PROLE ADULTA. PAPEL DAS  
PROSTAGLANDINAS DERIVADAS DA COX-2.

FERNANDA ELIZABETHE DOS RAMOS ALVES

ORIENTADOR: PROFº. DR. FABIANO ELIAS XAVIER

RECIFE, 2012.

FERNANDA ELIZABETHE DOS RAMOS ALVES

ESTUDO DOS MECANISMOS ENVOLVIDOS NO EFEITO DO DIABETES  
MELLITUS MATERNO SOBRE A FUNÇÃO MICROVASCULAR DE RATOS  
ADULTOS: INFLUÊNCIA DA IDADE.

Dissertação apresentada ao Departamento  
de Fisiologia e Farmacologia do Centro  
de Ciências Biológicas da Universidade  
Federal de Pernambuco, para obtenção  
do grau de Mestre em Bioquímica e  
Fisiologia.

ORIENTADOR:  
Prof. Dr. Fabiano Elias Xavier

RECIFE  
2012

Catalogação na fonte  
Elaine Barroso  
CRB 1728

**Alves, Fernanda Elizabeth dos Ramos**

Exposição à hiperglicemia materna induz disfunção endotelial em artérias de resistência da prole adulta. Papel das prostaglandinas derivadas da COX-2/ Fernanda Elizabeth dos Ramos Alves- Recife: O Autor, 2013.

97 folhas : il., fig., tab.

Orientador: Fabiano Elias Xavier

Dissertação (mestrado) – Universidade Federal de Pernambuco, Centro de Ciências Biológicas, Bioquímica e Fisiologia, 2013.

Inclui bibliografia

1. Diabetes 2. Hipertensão 3. Feto I. Xavier, Fabiano Elias (orientador) II. Título

616.462      CDD (22.ed.)      UFPE/CCB- 2013- 290

FERNANDA ELIZABETHE DOS RAMOS ALVES

ESTUDO DOS MECANISMOS ENVOLVIDOS NO EFEITO DO DIABETES  
MELLITUS MATERNO SOBRE A FUNÇÃO MICROVASCULAR DE RATOS  
ADULTOS: INFLUÊNCIA DA IDADE.

Dissertação apresentada para o  
cumprimento parcial das  
exigências para obtenção do título de  
Mestre em Bioquímica e Fisiologia pela  
Universidade Federal de Pernambuco.

Aprovado por:

---

Prof. Dr. Fabiano Elias Xavier

---

Profa. Dra. Glória Isolina Boente Pinto Duarte

---

Profa. Dra. Dayane Aparecida Gomes

---

Profa. Dra. Luciana Venturini Rossoni

---

Data: 29/08/2012

*Aos meus pais Rosana e Fernando,  
meus maiores exemplos  
de dedicação e perseverança.*

## AGRADECIMENTOS

A Deus que sempre tem me orientado em minhas escolhas.

A meus pais que como referências em superação e dedicação, me ajudaram a manter meus objetivos.

Ao Professor Fabiano Elias pela confiança e oportunidade depositadas em mim.

À professora Glória por todo apoio e pela oportunidade de aprendizado diário.

Ao José Antônio que forneceu o auxílio técnico, o qual foi indispensável para a produção deste trabalho.

A Diego que acompanhou e me auxiliou na confecção todo o trabalho.

Aos amigos do laboratório, Luciana, Juliana Rocha, Francine, Thayanne, Rebeca, Odair, Leylliane, Marcelo, Hicla, Carolina, Mayara, Maíra, professora Cristina e Alex, pessoas especiais que foram fundamentais em diversos momentos.

A Tiago, pelo companheirismo.

A Isabel, Prisse e Delba, pela amizade e admiração.

À Juliana Dantas, uma amiga sempre disposta a escutar e apoiar minhas escolhas.

A todos os professores e amigos da pós-graduação em Bioquímica e Fisiologia.

Aos membros da banca examinadora, por terem aceitado o convite e pela análise do trabalho.

À FACEPE (Fundação de Amparo à Ciência e Tecnologia do estado de Pernambuco) pelo apoio financeiro.

## RESUMO

O objetivo do presente estudo foi determinar como o diabetes durante a gestação afeta a função vascular na prole adulta, a influência da idade e se a ativação das vias das ciclooxygenases (COX) está envolvida neste efeito. O diabetes materno foi induzido por estreptozotocina no dia 7º de gestação em ratas Wistar. Estudos de reatividade vascular foram realizados para analisar o relaxamento à acetilcolina (ACh) e a contração à noradrenalina (NA) em artérias mesentéricas de resistência (AMR) de ratos machos provenientes de ratas controle (CON) e diabéticas (STZ) aos 3, 6 e 12 meses de idade (3M, 6M e 12M, respectivamente). A liberação de TxB<sub>2</sub>, PGE<sub>2</sub> e PGF<sub>2α</sub> foi determinada por imunoensaio enzimático. A expressão de COX-1 e COX-2 foi analisada por *Western blot*. A prole de ratas diabéticas desenvolveu hipertensão a partir dos 6 meses de idade comparada à prole de ratas controle. Em ratos STZ, o relaxamento à ACh foi reduzido em todas as idades estudadas enquanto à contração à NA foi aumentada no grupo STZ6M e STZ12M. Tanto a resposta à ACh, quanto à NA foram restauradas na presença do inibidor da COX-2 (NS-398). O bloqueio do receptor TP (SQ29548) restaurou o relaxamento à ACh nas artérias dos animais STZ3M, enquanto que no grupo STZ6M, somente o bloqueio dos receptores TP e EP (SQ29548+AH6809) foi capaz de restaurar o relaxamento à ACh e a contração à NA. No grupo STZ12M, as respostas à ACh e à NA foram restauradas quando os receptores TP, EP e FP foram bloqueados simultaneamente (SQ29548+AH6809+AL8810). A liberação de TxB<sub>2</sub> estimulada por ACh ou NA foi aumentada nos ratos STZ em todas as idades estudadas. O aumento da liberação de PGE<sub>2</sub> foi detectado nos vasos dos animais com 6 e 12 meses de idade, enquanto a produção de PGF<sub>2α</sub> aumentou apenas no grupo STZ12M. A expressão da COX-2, mas não da COX-1, foi aumentada nas artérias dos ratos STZ comparadas ao grupo controle. Estes resultados indicam a influencia da idade sobre a regulação da COX-2 em artérias de resistências de ratos adultos provenientes de ratas diabéticas, associada a uma produção aumentada de prostanoídes vasoconstritores. Este efeito desempenha um papel chave na patogênese da disfunção endotelial nestas artérias, que por sua vez poderia contribuir para a progressão da disfunção vascular nestes ratos.

**Palavras chave:** Diabetes; hipertensão; programação fetal; disfunção endotelial; ciclooxygenase; prostanoídes.

## ABSTRACT

The present study was designed to investigate how diabetes during pregnancy affects vascular function in adult offspring, the influence of age and the role of cyclooxygenase (COX)-derived prostaglandins. Blood pressure, relaxation to acetylcholine (ACh) and vasoconstriction to noradrenaline (NA) were analyzed in the offspring of control (O-CR) and diabetic (O-DR) mothers at 3, 6 and 12 months of age. O-DR developed hypertension from 6 months of age compared to O-CR. ACh elicited relaxation that was impaired in mesenteric resistance arteries (MRA) from 3, 6 and 12-month old O-DR. Contraction to NA was increased in MRA from 6 and 12-month old O-DR. COX-2 inhibition restored ACh and NA responses in all O-DR groups. TP receptor blockade (SQ29548) restored ACh relaxation in MRA from 3-month-old O-DR. In 6-month-old O-DR TP and EP receptors blockade (SQ29548+AH6809) was required to restore the relaxation to ACh and the contraction to NA. In 12-month-old O-DR, these responses were restored only when TP, EP and FP receptors were blocked (SQ29548+AH6809+AL8810). ACh- or NA-stimulated TxB<sub>2</sub> were higher in all O-DR. Release of PGE<sub>2</sub> increased in MRA from 6- and 12-month-old O-DR while PGF<sub>2α</sub> was increased only in MRA from 12-month-old O-DR. COX-2 expression was higher in MRA from O-DR than O-CR. In O-DR, COX-2 expression increased with age, reaching higher levels at 12 months of age. COX-1 expression was similar in all groups. Results indicate an age-dependent up-regulation of COX-2 coupled to an enhanced formation of vasoconstrictor prostanoids in MRA from O-DR. This increased formation of vasoconstrictor prostanoids plays a key role in the pathogenesis of endothelial dysfunction which in turn could contribute to progression of vascular dysfunction in these rats.

**Keywords:** Diabetes; hypertension; fetal programming; endothelial dysfunction; cyclooxygenase; prostanoids.

## **LISTA DE FIGURAS**

Figura 1. Metabolismo do ácido araquidônico e ação dos prostanóides sobre as células do músculo liso, plaquetas e células endoteliais.

Figura 2. Mecanismos de ação dos prostanóides.

## SUMÁRIO

1. INTRODUÇÃO	9
2. FUNDAMENTAÇÃO TEÓRICA	11
2.1. Diabetes gestacional.....	11
2.2. Fisiopatologia das alterações cardiovasculares associadas à exposição fetal à hiperglicemia materna.....	14
2.3. Sistema vascular e artérias de resistência na hipertensão e no diabetes .....	16
2.4 Controle endotelial do tônus vascular .....	20
2.4.1. Produtos derivados da via da ciclooxigenase .....	20
<i>Tromboxano A<sub>2</sub> (TXA<sub>2</sub>)</i> .....	23
<i>Prostaglandina E<sub>2</sub> (PGE<sub>2</sub>)</i> .....	24
<i>Prostaglandina I<sub>2</sub> (PGI<sub>2</sub>)</i> .....	25
<i>Prostaglandina F<sub>2α</sub> (PGF<sub>2α</sub>)</i> .....	25
2.5. Hipótese de trabalho .....	26
3. OBJETIVOS	28
4. REFERÊNCIAS BIBLIOGRÁFICAS	29
5. ARTIGO 1	53
6. ARTIGO 2	64
7. CONCLUSÕES	97

## 1. INTRODUÇÃO

Estudos epidemiológicos e experimentais têm fornecido evidências substanciais de que a exposição intrauterina à hiperglicemia materna tem efeitos sobre a descendência, incluindo aumento do risco de obesidade, de diabetes tipo 2 e de doenças cardiovasculares (CLAUSEN *et al.*, 2009; SIMEONI & BARKER, 2009).

O conceito de “programação fetal” (*fetal programming*) sugere que o feto pode ser “programado” durante o período intrauterino para desenvolver doenças na vida adulta, e tem sido utilizado como uma das bases para o conhecimento da origem de algumas doenças crônicas, como é o caso das doenças cardiovasculares (BARKER *et al.*, 1994; 1998). Trabalhos publicados pelo grupo do Dr. David Barker foram os primeiros a demonstrar o envolvimento de distúrbios ocorridos durante a fase intrauterina no desenvolvimento de doenças cardiovasculares na vida adulta (BARKER *et al.*, 1986; 1989; 1994; 1998). Barker e seus colaboradores observaram que a distribuição geográfica da taxa de mortalidade neonatal na Inglaterra e no País de Gales no início do século XIX era próxima da distribuição da taxa de mortalidade por doenças cardiovasculares setenta anos mais tarde. Devido ao fato de que a mortalidade neonatal no início do século XIX era atribuída ao baixo peso após o nascimento, Barker sugeriu que fatores iniciados durante a vida fetal, e que retardam o crescimento, poderiam “programar” ou definitivamente alterar a estrutura e/ ou a fisiologia de sistemas ligados ao desenvolvimento de doenças cardiovasculares na idade adulta (BARKER, 1998). Além disso, com base na associação entre peso corporal após o nascimento e a pressão arterial na vida adulta publicada em 1985 por Wadsworth *et al.*, Barker propôs que influências do ambiente fetal poderiam também alterar a pressão arterial na vida adulta (BARKER *et al.*, 1989). Vários estudos epidemiológicos atualmente suportam essa relação inversa entre peso corporal após o nascimento e a hipertensão arterial (LAW *et al.*, 1996; HUXLEY *et al.*, 2000). Além disso, uma elevação da pressão arterial também é encontrada em crianças com baixo peso corporal (LAW *et al.*, 2002). Estas observações serviram como base de sustentação da “Hipótese de Barker” que postula que condições adversas ocorridas no útero, como a desnutrição, podem desencadear um retardamento do crescimento e uma “programação fetal” da hipertensão arterial e de outras doenças cardiovasculares. Portanto, a programação fetal pode ser definida como um

fenômeno que ocorre através estímulos ou insultos no ambiente uterino durante um período considerado “janela crítica de desenvolvimento” podendo trazer alterações na estrutura e função de órgãos e tecidos ao longo da vida adulta (LEON *et al.*, 1998; SIMEONI & BARKER, 2009).

Dentre as alterações mais comuns que podem resultar em programação fetal está o consumo de baixa quantidade de calorias (BLONDEAU *et al.*, 2002) ou de proteínas (PINHEIRO *et al.*, 2008) durante a gestação e/ou aleitamento, a insuficiência placentária (GATFORD *et al.*, 2010), a exposição durante a gestação aos glicocorticóides (XU *et al.*, 2011) e o diabetes gestacional (BLONDEAU *et al.*, 2010; SILVERMAN *et al.*, 1991).

## 2. FUNDAMENTAÇÃO TEÓRICA

### 2.1. Diabetes gestacional

Segundo a *American Diabetes Association* (ADA), o diabetes mellitus gestacional (DMG) é comumente identificado como o surgimento de intolerância à glicose que inicia, ou é reconhecido pela primeira vez, durante a gravidez. O resultado desse processo, quando não tratado, é o aparecimento de efeitos adversos sobre o feto, que incluem macrossomia, sofrimento fetal, restrição de crescimento intrauterino e prematuridade (MANDERSON *et al.*, 2002; ROGERS & VELTEN, 2011).

O DMG é uma doença que afeta de 1 a 14% das mulheres, variando de acordo com a população estudada e os critérios diagnósticos utilizados (BRODY *et al.*, 2003, RIBEIRO *et al.*, 2011). No Brasil, os dados referentes a esta doença referem-se à última década, onde a estimativa era de que cerca de 3 a 4% das gestantes desenvolveriam DMG, o que significa mais de 200.000 novos casos por ano (MAGANHA *et al.*, 2003; RIBEIRO *et al.*, 2011; SCHMIDT *et al.*, 2000). Devido ao aumento da prevalência de obesidade, estima-se que o número de mulheres com DMG cresça substancialmente nos próximos anos, tanto nos países desenvolvidos como naqueles em desenvolvimento (KIM *et al.*, 2011b). Em países, como os Estados Unidos, aproximadamente 4-6% das gestações apresentam complicações relacionadas ao diabetes mellitus (ALBRECHT *et al.*, 2011; KIM *et al.*, 2011a). A maioria dos casos é diagnosticada pela primeira vez durante a gravidez e o restante dos casos é considerado como pré-gestacionais (DM tipo 1 ou tipo 2) (ALBRECHT *et al.*, 2011).

A gestação consiste em um período no qual a fêmea é transitoriamente submetida a alterações metabólicas que são finamente reguladas a fim de fornecer suprimento adequado ao feto. Uma vez que a unidade feto-placentária encontra-se sob a ação de diversos hormônios envolvidos no metabolismo da glicose, como a prolactina e os esteróides, e é capaz de secretar substâncias como a leptina (AKERMAN *et al.*, 2002), a resistina (LAPPAS *et al.*, 2005), e os fatores de crescimento semelhantes à insulina (SFERRUZZI-PERRI *et al.*, 2011), é possível observar que durante a gestação e a lactação ocorre uma complexa relação entre as alterações maternas e o controle da homeostase glicêmica no feto/ neonato.

Evidências de que a hiperglicemia materna é capaz de resultar em doenças na prole adulta foram inicialmente sugeridas através de estudos com uma população indígena da América do Norte, os índios Pima do Arizona (PETTIT *et al.*, 1983). Esta população apresenta elevada prevalência de diabetes tipo 2 durante a infância e a vida adulta (FRANKS *et al.*, 2006), sendo este fato associado à exposição aos altos níveis glicêmicos durante a vida intrauterina. Além disso, tem-se demonstrado que nesta população há risco aumentado de obesidade, hipertensão e dislipidemias na vida adulta (DABELEA *et al.*, 1993; MANDERSON *et al.*, 2002; SILVERMAN *et al.*, 1991).

Além de fatores ambientais, fatores genéticos têm sido considerados como uma possível explicação para a associação entre exposição perinatal a altas concentrações de glicose e o aparecimento de doenças metabólicas e cardíacas. De acordo com essa hipótese, mães com início precoce de diabetes ou com DMG podem ter um genótipo específico que transmite alta suscetibilidade à diabetes mellitus tipo 2 para a prole (SIMEONI & BARKER, 2009). Entretanto, dados disponíveis indicam também um papel importante do ambiente intrauterino sobre estas alterações. Trabalhos realizados na população de índios Pima sugerem esta conclusão. A prevalência de diabetes mostrou ser semelhante em mães que desenvolveram o diabetes durante a gravidez em relação àquelas com diabetes pré-existente (PETTIT *et al.*, 1998). Além disso, a frequência para o desenvolvimento de diabetes tem se mostrado maior em filhos de mãe diabética do que em descendentes de homens diabéticos (LINDSAY *et al.*, 2000). Portanto, a susceptibilidade genética para o diabetes poderá agir apenas como um fator predisponente ou agravante na determinação do risco de diabetes sobre a geração, quando adulta (SIMEONI & BARKER 2009; STRIDE *et al.*, 2002).

Mulheres com diabetes pré-existente apresentam risco aumentado para complicações da gestação, incluindo retardo no crescimento fetal e morte perinatal (DUNNE *et al.*, 2003). Embora os mecanismos a cerca dessas associações não sejam completamente compreendidos, tem sido mostrado que o controle da glicemia no início da gestação é capaz de reduzir tais complicações (KITZMILLER *et al.*, 2008). Por outro lado, mulheres com diabetes gestacional que se mantêm hiperglicêmicas durante toda a gestação apresentam complicações mais severas comparadas àquelas que apresentam o diabetes durante o segundo ou terceiro trimestre de gestação, (CASEY *et al.*, 1997; KIM *et al.*, 2011b). Dessa forma, admite-

se que o diabetes pré-existente e o diabetes gestacional podem promover efeitos distintos sobre o desenvolvimento fetal e em longo prazo da prole, sendo necessário investigar as associações entre estas duas condições separadamente. Além disso, na mãe, o diabetes durante a gestação promove um risco adicional para desenvolvimento de doenças cardiovasculares e renais como, a hipertensão arterial, os acidentes vasculares e a insuficiência renal (PARADISI *et al.*, 2002; RADENKIVIC *et al.*, 2009).

A glicose é o principal nutriente que passa para o feto através de difusão facilitada (RUDGE *et al.*, 2011). Dessa forma, adaptações nas células de contato entre a circulação materna e a fetal podem ocorrer em resposta ao abundante suprimento de glicose. Como demonstrado por Taricco *et al.*, (2003), placenta de mães diabéticas mostram diversas modificações estruturais quando comparadas com placenta de mães não diabéticas, além disso, em outros estudos é possível observar imaturidade das vilosidades (DASKALAKIS *et al.*, 2008), aumento de peso placentário e endoarterite quando em condições de diabetes (RUDGE *et al.*, 2011; SALOMÓN *et al.*, 2012).

Embora o mecanismo da relação entre a condição metabólica materna e o metabolismo da glicose na prole não esteja completamente esclarecido, pelo menos uma parte do efeito é atribuído à exposição intrauterina à elevada glicose materna. A glicose cruza livremente a placenta e estimula o pâncreas fetal a produzir insulina (FREINKEL, 1980) acarretando alterações potenciais sobre este órgão e sobre a sensibilidade dos tecidos à insulina. Estudos *in vitro* com fetos humanos oriundos de mães diabéticas demonstraram redução da secreção de insulina regulada por estimulação adrenérgica (REIHER *et al.*, 1983). Em ratos, a exposição à hiperglicemia materna tem sido associada ao desenvolvimento anormal das células beta-pancreáticas e redução da absorção de glicose pelo músculo esquelético (BOLOKER *et al.*, 2002).

Nesse sentido, com a finalidade de estudar a programação do desenvolvimento intrauterino durante a exposição à hiperglicemia materna, modelos animais têm sido desenvolvidos e têm sido relacionados ao aparecimento de alterações metabólicas, cardiovasculares e renais (GILL-RANDALL *et al.*, 2004; FETITA *et al.*, 2006; WEST *et al.*, 2011). Estudos deste porte, em humanos são limitados por questões éticas e pelo aparecimento de variáveis não controladas que podem modificar o ambiente intrauterino como, o comportamento alimentar e

socioeconômico, o *status* nutricional e os fatores genéticos (MOSES, 2012; NOMURA *et al.*, 2012). Um método comumente utilizado para induzir hiperglicemia durante a gravidez é a injeção de estreptozotocina (STZ) (WARD *et al.*, 2001) que é uma glicosamina-nitrosuréia capaz de promover efeitos tóxicos sobre as células  $\beta$ -pancreáticas (JUNOD *et al.*, 1967). Nesta situação, observa-se risco aumentado de complicações sobre o feto, como má formação congênita, aborto e morte perinatal (CARRAPATO & MARCELINO, 2001; COX, 1994; YESSOUFOU & MOUTAIROU, 2011).

Quanto às alterações sobre o peso corpóreo da prole de ratas diabéticas pela injeção de STZ, observa-se tanto microssomia, na qual se observa baixo peso e retardo no crescimento (AERTS *et al.*, 1990), macrossomia (OH *et al.*, 1990; GELARDI *et al.*, 1990), caracterizada por excesso de peso, ou ainda peso e crescimento normais (GERBER *et al.*, 2000). O baixo peso ao nascimento, como consequência de alterações do ambiente intrauterino pode resultar tanto da desnutrição materna induzida por restrição protéica, como do aporte inadequado de nutrientes para o feto (HUXLEY *et al.*, 2000; LAW *et al.*, 1996). Neste sentido, Canavan & Goldspink (1988) demonstraram que a hiperglicemia durante a gestação induz supressão do crescimento fetal, o qual está associado com redução da síntese de proteína. Entretanto, este efeito está diretamente relacionado ao grau de elevação da glicemia materna. Assim, como consequência de uma hiperglicemia leve observa-se filhos com maior peso, enquanto que em um processo hiperglicêmico mais grave a prole apresenta baixo peso ao nascimento como consequência da maior redução na síntese proteica (SEGAR *et al.*, 2009).

Portanto, estes modelos têm permitido investigar, em longo prazo, os efeitos da exposição intrauterina à hiperglicemia e suas consequências sobre a prole adulta. Embora, este tipo "programação fetal", no que se refere às doenças cardiovasculares tenha atraído muita atenção, os mecanismos envolvidos ainda não foram completamente elucidados.

## **2.2. Fisiopatologia das alterações cardiovasculares associadas à exposição fetal à hiperglicemia materna.**

As doenças cardiovasculares são as principais causas de morte no mundo ocidental e o diabetes mellitus representa o maior fator de risco para o desenvolvimento destas doenças (CUGNET-ANCEAU *et al.*, 2009).

Há diversos indícios epidemiológicos e estudos em animais indicando que a exposição perinatal ao diabetes materno está associada com o desenvolvimento de diabetes tipo 2 e de doenças cardiovasculares durante a vida adulta (METZGER *et al.*, 2002; SIMEONI & BARKER 2009). Além disso, tem-se verificado que o prejuízo no metabolismo da glicose adquirido durante a “janela de desenvolvimento” pode ser transmitido às gerações subseqüentes, possivelmente através de alterações epigenéticas sobre a expressão gênica (PINHEIRO *et al.*, 2008).

Sabe-se que as consequências citadas anteriormente são resultados da combinação entre mecanismos que atuam em níveis molecular, celular e tecidual. Órgãos ou sistemas, como o rim ou os vasos sanguíneos, alcançam o desenvolvimento total no período perinatal, estando mais suscetíveis aos danos ocorridos durante este período (SIMEONI & BARKER, 2009). Amri *et al.* (1999) demonstraram que em ratos a exposição do feto à hiperglicemia materna diminui a nefrogênese, fato que contribui para o desenvolvimento de insuficiência renal crônica e hipertensão arterial na vida adulta. Estudos de Rocha *et al.* (2005) e Cavanal *et al.* (2007) demonstraram que ratos adultos normoglicêmicos provenientes de ratas diabéticas apresentam prejuízo da função renal e hipertensão arterial. Outros trabalhos têm ainda mostrado que filhos de mães diabéticas apresentam elevada incidência de intolerância à glicose, resistência à insulina e obesidade (BOLOKER *et al.*, 2002; BLONDEAU *et al.*, 2010), o que também predispõe o indivíduo ao desenvolvimento de diabetes e de doenças cardiovasculares na vida adulta.

Utilizando modelos de diabetes materno severo, estudos mostram que descendentes de ratas hiperglicêmicas apresentam disfunção endotelial quando adultos. Segar *et al.*, (2010) desenvolveram um modelo de hiperglicemia em ratos durante a segunda semana de gestação (13º dia), onde foi possível observar disfunção endotelial e outras complicações cardiovasculares, tróficas e metabólicas na prole adulta. Utilizando modelo semelhante, Reinking *et al.* (2009) demonstraram que animais oriundos de ratas diabéticas apresentam cardiomiopatia hipertrófica, similar à encontrada em humanos descendentes de mães diabéticas. Além disso, Manderson *et al.* (2002) demonstraram que na prole de ratas diabéticas ocorre aumento plasmático na concentração de moléculas de adesão, incluindo a E-selectina, associado a uma maior predisposição a doenças vasculares.

Na última década, diversos trabalhos indicam que a hiperglicemia altera a angiogênese na prole em vários modelos experimentais, possivelmente através de uma redução na proliferação e aumento da apoptose de células endoteliais, estando estes mecanismos sob regulação do fator angiogênico VEGF (Fator de crescimento vascular) (LARGER *et al.*, 2004; PINTER *et al.*, 2001). Neste sentido, Ingram *et al.*, (2008) descreveram que células progenitoras endoteliais isoladas de filhos de mães diabéticas apresentam funções angiogênicas alteradas, com redução da renovação e da capacidade de formação vascular. O estudo mostrou ainda que as células progenitoras endoteliais de descendentes de mães diabéticas apresentam um processo de senescência celular acelerado, o que pode predispor o indivíduo ao aparecimento de disfunção endotelial e de outras complicações vasculares (ACOSTA *et al.*, 2011; INGRAM *et al.*, 2008). Dessa forma, é possível concluir que uma disfunção endotelial precoce pode contribuir posteriormente para o surgimento de outras alterações vasculares e, consequentemente, de hipertensão arterial na descendência de mães diabéticas.

### **2.3. Sistema vascular e artérias de resistência na hipertensão e no diabetes**

Hoje se sabe que existe uma grande heterogeneidade na reatividade dos diferentes territórios vasculares, a qual varia de acordo com os tipos e densidade de receptores farmacológicos e com os mecanismos de transporte de íons no músculo liso (MULVANY & ASLHJAEER, 1990). Além disso, há uma série de trabalhos demonstrando que a liberação de fatores endoteliais pode variar dependendo do tamanho e localização dos vasos sanguíneos (CLARK & FUCHS, 1997; LAGAUD *et al.*, 1999).

A parede arterial é composta por três camadas claramente distintas: 1). A camada íntima, constituída por células endoteliais diferenciadas e arranjadas ao longo do eixo axial do vaso e na direção do fluxo sanguíneo, uma camada subendotelial contendo tecido conjuntivo e de uma membrana basal; 2). A camada média, constituída de células musculares lisas, as quais se encontram circumferencialmente arranjadas ao longo do eixo arterial e de uma matriz extracelular que inclui lâminas de fibras elásticas, fibras colágenas e proteoglicanos e 3). A camada adventícia, formada por tecido conjuntivo, contendo basicamente

fibras de colágeno, fibras elásticas e componentes celulares como fibroblastos e mastócitos (PEARSON, 1976).

As artérias de resistência podem ser definidas como vasos pré-arteriolares que contribuem passivamente e ativamente para a manutenção da resistência basal e para o controle do fluxo sanguíneo durante alterações hemodinâmicas (FEIH *et al.*, 2008). A dissipação da energia necessária para superar a resistência vascular, evidenciada por um acentuado declínio na pressão arterial ocorre, principalmente, nas arteríolas pré-capilares e nas pequenas artérias, ou seja, nos vasos com diâmetro inferior a 350 µm (BORDERS & GRANGER, 1986; CHRISTENSEN & MULVANY, 2001; MULVANY & AALKJAER, 1990).

Alterações funcionais e estruturais nas artérias de resistência têm sido bem documentadas na hipertensão essencial e secundária (AGABITI, 2003; HEAGERTY, 2007) e também no diabetes mellitus tipos 1 e 2 (GREENSTEIN *et al.*, 2009; LEVY *et al.*, 2008). Estas alterações vasculares representam um dos principais fatores para o aumento da resistência vascular periférica, contribuindo, portanto, para o estabelecimento da hipertensão arterial. Essas alterações incluem espessamento da parede arterial e redução do diâmetro interno nas artérias de resistência (100 - 350 µm de diâmetro interno). Tais mudanças estruturais tem sido observadas em diversos leitos vasculares como em tecidos subcutâneo, omental, mesentérico e cerebral de ratos hipertensos (RIZZONI *et al.*, 2001; RIZZONI *et al.*, 2009). Entretanto, está cada vez mais evidente que as alterações nestes vasos não são ocasionadas apenas pela elevação crônica da pressão arterial. Outros fatores estão possivelmente envolvidos nas interações entre hipertensão e diabetes, incluindo a ativação do sistema nervoso simpático e do sistema renina-angiotensina, resistência à insulina, aumento dos níveis de leptina, disfunção endotelial e estresse oxidativo (BELIN DE CHANTEMÈLE *et al.*, 2009; CHANG *et al.*, 2012; GRASSI & DIEZ, 2009; NAKAJINMA *et al.*, 2010).

A relação entre o diabetes e as complicações cardiovasculares é complexa, mas uma característica marcante consiste no prejuízo da função endotelial. A exposição crônica das proteínas plasmáticas e das membranas celulares à hiperglicemia implica em aumento da via dos polióis, glicosilação não-enzimática de proteínas, com aumento na produção dos produtos finais da glicação avançada (AGEs), sendo estes últimos conhecidos por inativarem o óxido nítrico, levando como consequência ao prejuízo da vasodilatação dependente do endotélio

(BUCALA *et al.*, 1991). Nas proteínas como o colágeno, os AGEs causam ligações cruzadas entre os polipeptídios da membrana, levando ao aprisionamento das proteínas plasmáticas ou intersticiais (KALOUSOVÁ *et al.*, 2002). Nos vasos de resistência e nos mais calibrosos, o aprisionamento da lipoproteína de baixa densidade (LDL) retarda seu efluxo a partir da parede vascular, acelerando a deposição do colesterol no endotélio e estimulando o processo aterogênico (JAKUS & RIETBROCK, 2004). Adicionalmente, nos capilares, a membrana basal glicosilada provoca a fixação de proteínas plasmáticas como a albumina, caracterizando a microangiopatia diabética (TAYLOR & POSTON, 1994).

Sabe-se que a hiperglicemia aguda ou crônica é capaz de causar inúmeras alterações sobre a função de pequenas e de grandes artérias, dentre elas: diminuição do relaxamento dependente do endotélio (DE VRIESE *et al.*, 2000; OZYAZGAN *et al.*, 2000; BROUWERS *et al.*, 2010), aumento da resposta contrátil do músculo liso vascular (SHI *et al.*, 2007; SHI & VANHOUTTE, 2008; XAVIER *et al.*, 2003) e predisposição ao desenvolvimento de eventos inflamatórios, trombóticos e ateroscleróticos (SOWERS 1990; VILAHUR *et al.*, 2009).

Um dos mecanismos responsáveis pela alteração vascular observada no diabetes inclui a reduzida disponibilidade do óxido nítrico (NO), seja por sua menor liberação/ biodisponibilidade (HAIDARA *et al.*, 2006) ou redução da sensibilidade do músculo liso (FÉLÉTOU *et al.*, 2010). Estas alterações normalmente, estão associadas com uma superprodução de espécies reativas de oxigênio (EROs), peroxidação lipídica e elevada produção de moléculas de adesão (DE VRIESE *et al.*, 2000; HINK *et al.*, 2001; MAZZONE *et al.*, 2008). Além disso, a formação de produtos de glicação avançada e o diacilglicerol (DAG) juntamente com os ânions superóxidos contribuem para o desacoplamento da via da NOS (óxido nítrico sintase) endotelial através da depleção de tetrabiopterina ( $BH_4$ ), um co-fator essencial para a síntese de NO (FÉLÉTOU *et al.*, 2010).

Evidências crescentes sugerem que a disfunção endotelial em estados hiperglicêmicos é paralela ao estado de resistência insulínica, estado no qual há resposta inadequada dos tecidos à molécula, acompanhada de elevados níveis plasmáticos deste hormônio (HITOMI *et al.*, 2011; VAZZANA *et al.*, 2012). A insulina, em concentrações fisiológicas, atua como um vasodilatador e estimula a produção endotelial de NO (PESSIN & SALTIEL, 2000). Entretanto, na presença da resistência à insulina, é possível verificar prejuízo na captação periférica da glicose e na

vasodilatação dependente do endotélio mediada pelo hormônio (HITOMI *et al.*, 2011; TAGUCHI *et al.*, 2012). Diversos mecanismos inter-relacionados contribuem para a disfunção endotelial relacionada ao estado de resistência, tais como baixos níveis de colesterol HDL, aumento nos níveis de LDL e elevação de marcadores de inflamação (CHYU *et al.*, 2011; FAGAN & DEEDWANIA, 1998; TOGASHI *et al.*, 2012).

Diante dessa perspectiva, é possível deduzir que as alterações cardiovasculares observadas em animais expostos à hiperglicemia materna podem ser consequência de uma somatória de fatores, incluindo aqueles relacionados ao metabolismo da insulina. A maioria dos estudos em proles de ratas diabéticas demonstra hiperinsulinemia, a qual surge como um mecanismo adaptativo a fim de compensar o ambiente hiperglicêmico materno (BLONDEAU *et al.*, 2002; BLONDEAU *et al.*, 2010; GUZMÁN-GUTIÉRREZ *et al.*, 2011). Entretanto, estes animais apresentam, além da hiperinsulinemia, resistência à insulina.

Estudos prévios realizados em nosso laboratório demonstraram uma relação entre o aparecimento de desordens metabólicas e a hipertensão arterial em animais da prole de ratas diabéticas tipo 1. Os resultados desse estudo demonstraram que ratos adultos provenientes de ratas diabéticas desenvolvem alterações na homeostase da glicose, caracterizadas por intolerância à glicose e resistência à insulina. Além disso, observou-se que os animais com 6 e 12 meses de idade apresentaram hipertensão arterial, que foi acompanhada de disfunção endotelial. Os experimentos de reatividade vascular utilizando artérias de condutância (aorta e artéria mesentérica superior) revelaram um prejuízo do relaxamento dependente do endotélio aos 6 e 12 meses de idade e aumento da contratilidade à fenilefrina. Além disso, os resultados revelaram que esse desequilíbrio estaria associado a uma menor biodisponibilidade do óxido nítrico, em decorrência de uma maior produção de radicais livres acompanhada de uma liberação aumentada de substâncias vasoconstritoras derivadas da ciclooxygenase (de QUEIROZ, 2010).

Sendo assim, as complicações vasculares em indivíduos expostos a estados hiperglicêmicos em fases iniciais da vida podem contribuir para o aumento da morbimortalidade das doenças cardiovasculares observado nas últimas décadas. Os mecanismos envolvidos no desenvolvimento destas doenças são complexos e parcialmente compreendidos, mas se sabe que são em grande parte desencadeados por um desequilíbrio na função do endotélio.

## 2.4. Controle endotelial do tônus vascular

As células endoteliais produzem diversas substâncias capazes de modular o tônus vascular. Dentre as substâncias vasoconstritoras se destacam os derivados do metabolismo do ácido araquidônico pela via das ciclooxygenases (SIMMONS *et al.*, 2004; VIRDIS *et al.*, 2010), os ânions superóxido (PARAVICINI & TOUYZ 2007; WENCESLAU *et al.*, 2011), a endotelina-1 (BARTON, 2011) e a angiotensina II (YOU *et al.*, 2005). Entre os fatores vasodilatadores, destacam-se o óxido nítrico (NO) (FURCHGOTT, 1983; GILL *et al.*, 2007), a prostaciclina (PGI<sub>2</sub>) (TRACHTE, 1986) e o fator hiperpolarizante derivado do endotélio (EDHF) (CHEN *et al.*, 1991; FÉLÉTOU & VANHOUTTE *et al.*, 2006 ).

A integridade endotelial é essencial para a regulação do tônus vascular. Em condições fisiológicas existe um equilíbrio entre a liberação de fatores vaso relaxantes (EDRF) e vasoconstritores (EDCF). No entanto, em diversas condições patológicas, como na hipertensão arterial e no diabetes, esse equilíbrio é alterado, levando a um aumento da produção de EDCFs. Nos últimos anos, diversos estudos demonstram haver uma associação entre o aumento da produção de Prostaglandina H<sub>2</sub> (PGH<sub>2</sub>) e tromboxano A<sub>2</sub>, derivados do metabolismo do ácido araquidônico pela COX, e o estabelecimento da disfunção endotelial (ALFRANCA *et al.*, 2006; FÉLÉTOU *et al.*, 2010; PANNIRSELVAM *et al.*, 2005).

### 2.4.1. Produtos derivados da via da ciclooxygenase

As prostaglandinas, potentes mensageiros bioativos, são produzidas a partir do ácido araquidônico (AA), e foram extraídas a partir de vesículas seminais por Goldblatt e von Euler em 1930, quando foi evidenciada a ação contratil destas substâncias. A biossíntese das prostaglandinas depende basicamente da liberação do ácido araquidônico a partir da ação da fosfolipase A<sub>2</sub> sobre os fosfolipídeos da membrana (HAMBERG & SAMUELSSON, 1973; SIMMONS *et al.*, 2004).

O ácido araquidônico pode ser oxidado pelas ciclooxygenases para produzir o endoperóxido cíclico (PGH<sub>2</sub>) que servirá de substrato para a produção das prostaglandinas E<sub>2</sub> (PGE<sub>2</sub>), F<sub>2α</sub> (PGF<sub>2</sub>), da PGI<sub>2</sub> e do tromboxano A<sub>2</sub> (TxA<sub>2</sub>) (Figura 1). Além disso, ele pode ainda ser oxidado pelas lipoxigenases e originar o ácido 15-s-hidroxieicosatetraenóico (15-HETE), uma substância que é capaz de produzir contração vascular (VANHOUTTE & EBER, 1991). Adicionalmente, foi demonstrado por Miller & Vanhoutte (1985) que o próprio ácido araquidônico tem a capacidade de

induzir contração dependente do endotélio em veias femorais de cães e que esta resposta é bloqueada por inibidores das ciclooxigenases.

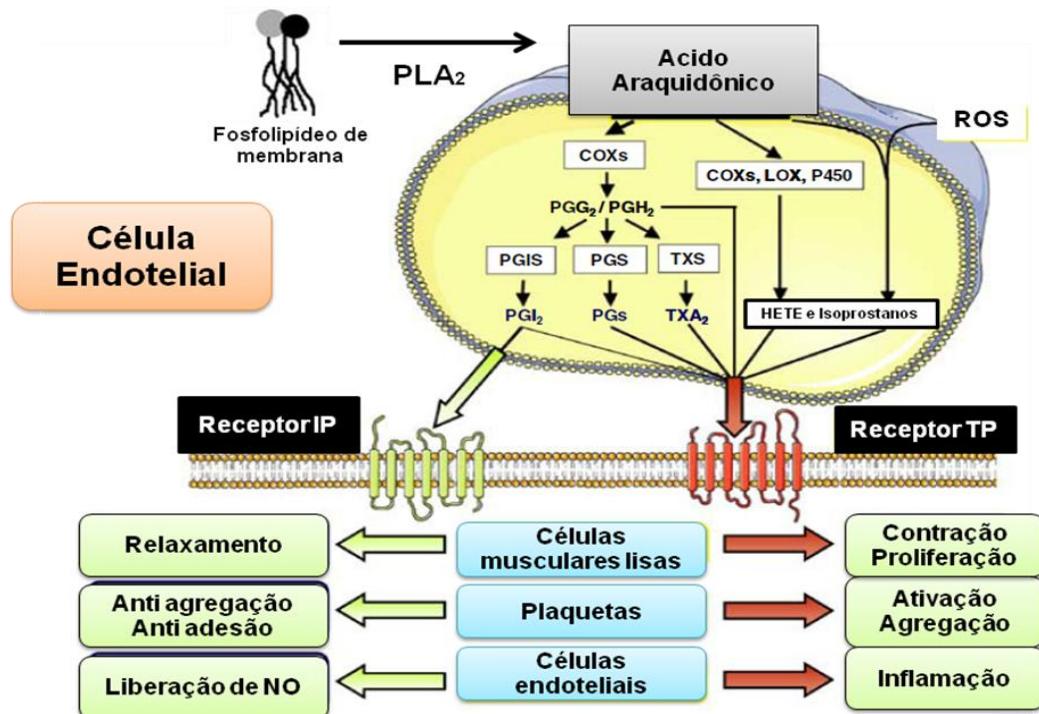


Figura 1: Metabolismo do ácido araquidônico e ação dos prostanóides sobre as células do músculo liso, plaquetas e células endoteliais. Envolvimento dos receptores IP e TP sobre a disfunção vascular. COX 1 e 2 catalisam a conversão do ácido araquidônico a endoperóxido cíclico PGH<sub>2</sub> e PGG<sub>2</sub> através de reações sequenciais de ciclooxigenação e hidroperoxidação. PGH<sub>2</sub> é metabolizado pelas sintases específicas para produzir prostanoides: PGS Sintases de prostaglandinas, PGIS Prostaglandina sintase, TXS Tromboxano sintase, PGs prostaglandinas, PGG<sub>2</sub> prostaglandina G<sub>2</sub>, PGH<sub>2</sub> prostaglandina H<sub>2</sub>, PGI<sub>2</sub> prostaglandina I<sub>2</sub> (prostaciclina), TXA<sub>2</sub> tromboxano A<sub>2</sub>. O ácido araquidônico também pode ser oxidado pelo citocromo P450, e pelas Lipooxigenases. PGH<sub>2</sub> prostaglandina H<sub>2</sub>, PGI<sub>2</sub> prostaglandina I<sub>2</sub> (prostaciclina), TXA<sub>2</sub> tromboxano A<sub>2</sub>, HETE ácido hidroxieicosatetraenoico (Adaptado FÉLETOU et al., 2010).

Além das prostaglandinas citadas, a ativação da COX também pode levar à formação de espécies reativas de oxigênio, como os ânions superóxido, e de isoprostanos, já que estas enzimas são capazes de co-oxidar outras substâncias como a molécula de NADPH (nicotinamida-adenina dinucleotídeo fosfatado) (TANG et al., 2007; WATKINS et al., 1999), e de modificar através de oxidação não enzimática, ácidos graxos poliinsaturados. Espécies reativas de oxigênio diminuem a biodisponibilidade de óxido nítrico (RUBANYI & VANHOUTTE, 1986) e como um ciclo de retroalimentação positiva, há estimulação da COX produzindo mais prostanoides (MORITA, 2002). Tanto os ânions superóxido, como os isoprostanos

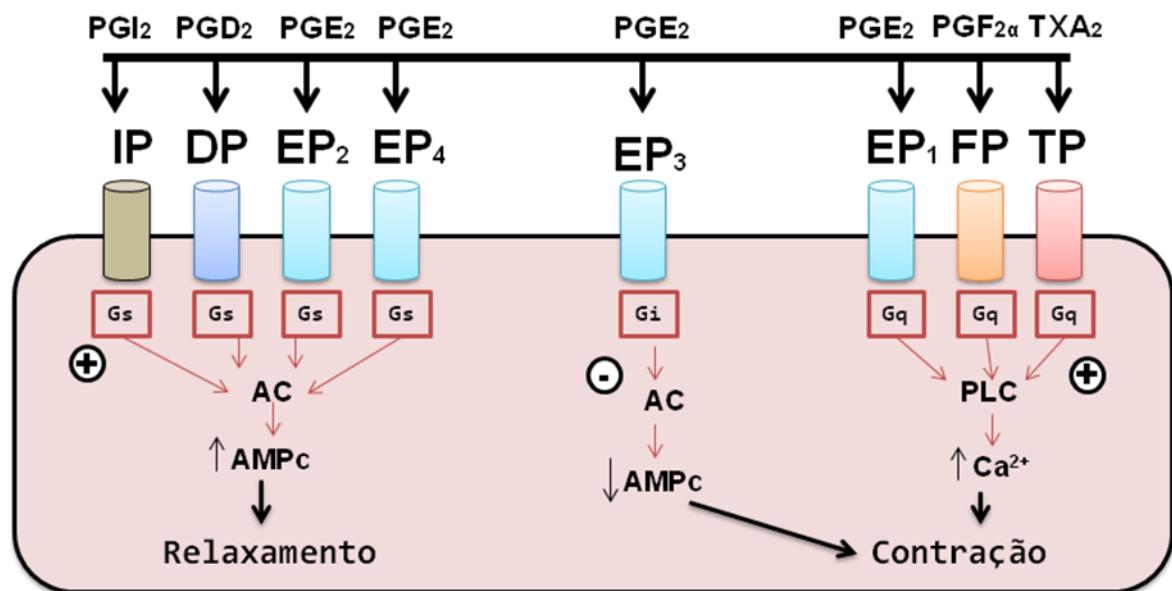
podem ser considerados biomarcadores de estresse oxidativo e têm sido associados a diversas doenças cardiovasculares (HAMILTON *et al.*, 2001; PRATICO *et al.*, 2001).

Existem duas isoformas da ciclooxygenase: a COX-1 e a COX-2. Ambas são heme-proteínas que apresentam potência semelhante para oxidar o ácido araquidônico em endoperóxido (GARAVITO & DE WITT, 1999), embora, baixas concentrações de ácido araquidônico são preferencialmente metabolizadas pela COX-2, enquanto que em níveis mais elevados o AA é metabolizado pela COX-1 (MORITA, 2002). Além disso, a COX-2 pode metabolizar outros substratos como, os ácidos graxos e o 2-araquidonil glicerol (SMITH & SONG, 2002) levando, portanto, a uma diferenciada produção de metabólitos entre as duas isoformas.

Considera-se geralmente que a COX-1 na maioria dos tecidos é expressa constitutivamente enquanto que a COX-2 é induzida principalmente nos locais de inflamação (FELETOU *et al.*, 2010). Alguns pesquisadores consideram que as prostaglandinas derivadas através do metabolismo da COX-1 apresentam papel positivo, visto que possuem funções fisiológicas importantes como controle da hemostasia, integridade da mucosa gastrointestinal e fluxo sanguíneo renal. Já as originadas da COX-2 têm papel negativo, pois estão envolvidas em reações inflamatórias e são responsáveis por fenômenos como dor, rubor, edema capilar e vasodilatação (PARENTE & PERRETTI, 2003). No entanto, a COX-2 também é expressa constitutivamente em vários tipos celulares, incluindo nas células endoteliais onde sua expressão é regulada pela tensão de cisalhamento (TOPPER *et al.*, 1996), e em diversas partes do néfron (ZHANG *et al.*, 1997). Na parede vascular, tanto células endoteliais como células musculares lisas contêm COX, porém, em vasos sanguíneos saudáveis, a proporção é maior nas células da camada íntima (DE WITT *et al.*, 1983). Sob determinadas condições como, por exemplo, no processo de envelhecimento e inflamação, a COX-2, pode ser expressa em células endoteliais e do músculo liso vascular, participando das contrações dependentes do endotélio (SHI *et al.*, 2008).

Uma relação entre o aumento da expressão das ciclooxygenases e o estabelecimento de disfunção endotelial tem sido demonstrada em modelos de hipertensão e diabetes. Em aortas de animais espontaneamente hipertensos (SHR), as contrações dependentes do endotélio são associadas ao aumento na expressão de COX-1, e a uma maior liberação de prostaciclina e tromboxano A<sub>2</sub> (FÉLÉTOU *et*

al., 2009). Adicionalmente, em modelos de diabetes tipo 2, o aumento do tônus arteriolar tem sido atribuído a uma maior expressão de COX-2, levando a uma produção aumentada de prostanoídes vasoconstritores (BAGI *et al.*, 2005; GUO *et al.*, 2005). Estes agentes contráteis interagem com receptores acoplados à proteína G, que são classificados em três subtipos: EP, FP, e TP, de acordo com a sensibilidade às prostaglandinas  $E_2$ ,  $F_{2\alpha}$  e tromboxano  $A_2$ , respectivamente (ALFRANCA *et al.*, 2006; FELETOU *et al.*, 2010; TSUBOI *et al.*, 2002) (Figura 2).



**Figura 2.** Mecanismos de ação dos prostanoídes. AC: adenilato ciclase, AMPc: Monofosfato cíclico de adenosina, DP: receptor da PGD<sub>2</sub>, EP: receptor de PGE, FP: receptor de PGF<sub>2α</sub>, IP: receptor de PGI<sub>2</sub>, TP: receptor de TxA<sub>2</sub>, PLC: fosfolipase C. (Adaptado BREYER *et al.*, 2011; YAGAMI, 2006).

### Tromboxano $A_2$ ( $TxA_2$ )

O  $TxA_2$  é sintetizado a partir da ação da enzima  $TxA_2$ -sintetase sobre o  $PGH_2$ . Ele é considerado um dos prostanoídes vasoconstritores mais importantes, também apresenta ação agregante plaquetária, promove a expressão de moléculas de adesão e favorece a infiltração de macrófagos através da modulação de receptores do tipo tirosina quinase (FONLUPT *et al.*, 1991; NAKAHATA, 2008). Sua ação é mediada através do receptor para tromboxano (receptor TP) nas células musculares lisas, onde a transdução do sinal envolve uma proteína Gq resultando em estimulação da fosfolipase C, o que promove aumento na concentração intracelular

de  $\text{Ca}^{2+}$  induzindo vasoconstrição (MAYEUX *et al.*, 1989) (Figura 2). Em humanos, mas não em roedores, duas isoformas para o receptor TP têm sido descritas, TP $\alpha$  e TP $\beta$ , as quais diferem apenas nas suas porções C-terminais (HIRATA *et al.*, 1996).

A liberação do TXA<sub>2</sub>, como de outras prostaglandinas pelas células endoteliais, ocorre através da ação de agonistas vasoconstritores (noradrenalina, serotonina, fenilefrina, angiotensina II, endotelina-1, etc), vasodilatadores (acetilcolina, bradicinina, etc) e por estímulos mecânicos (TADDEI & VANHOUTTE, 1993).

Estudos recentes tem mostrado que espécies reativas de oxigênio são capazes de aumentar a densidade de receptores TP na membrana de células musculares lisas (VALENTIN *et al.*, 2004; WILSON *et al.*, 2009), além disso, em células endoteliais, a ativação de receptores TP reduz a produção de óxido nítrico (LIU *et al.*, 2009, XAVIER *et al.*, 2009), fato que pode, portanto, contribuir para o estabelecimento de disfunção endotelial. Neste sentido, é possível compreender a importância deste prostanóide no desenvolvimento de desordens vasculares.

Embora o tromboxano A<sub>2</sub> seja o ligante preferencial para o receptor TP, outras prostaglandinas, inclusive o PGH<sub>2</sub>, também podem ativar este receptor (GLUAIS *et al.*, 2005). Além disso, isoprostanos são também potentes agonistas endógenos dos receptores TP (WATKINS *et al.*, 1999) (Figura 2).

### *Prostaglandina E<sub>2</sub> (PGE<sub>2</sub>)*

A PGE<sub>2</sub> corresponde ao prostanóide mais abundante do organismo (SERHAN & LEVY, 2003), sendo produzido por diversos tipos celulares, como fibroblastos e células endoteliais. A ampla capacidade de promover efeitos pró e antiinflamatórios consiste na ativação de diferentes tipos e receptores, denominados EP<sub>1</sub>, EP<sub>2</sub>, EP<sub>3</sub> e EP<sub>4</sub>.

No sistema vascular, os subtipos EP<sub>1</sub> e EP<sub>3</sub> promovem vasoconstrição, a qual ocorre através da ativação da via do IP<sub>3</sub>/Ca<sup>2+</sup> (receptor EP<sub>1</sub>) ou da diminuição dos níveis intracelulares de AMPc (receptor EP<sub>3</sub>) por inibição da adenilil ciclase (COLEMAN *et al.*, 1994; FUNK *et al.*, 1993). A ligação da PGE<sub>2</sub> aos receptores EP<sub>2</sub> provoca vasodilatação através do aumento nos níveis de AMPc via ativação da adenilil ciclase (COLEMAN *et al.*, 1994), enquanto que através do receptor EP<sub>4</sub> há ativação da via do NO (HRISTOVSKA *et al.*, 2007) (Figura 2).

Portanto, uma diminuição na vasodilatação mediada pelos receptores EP<sub>2</sub>/EP<sub>4</sub> ou um aumento da sinalização através dos receptores EP<sub>1</sub>/EP<sub>3</sub> pode estar envolvido no desequilíbrio da função vascular e na fisiopatologia da hipertensão. Guan *et al.* (2007) demonstraram que a ativação do receptor EP<sub>1</sub> contribui para a ação vasoconstritora da angiotensina II. Além disso, tem sido demonstrado que a PGE<sub>2</sub> desempenha um papel chave no desenvolvimento da doença renal hipertensiva (SUGANAMI *et al.*, 2003).

#### *Prostaciclina (PGI<sub>2</sub>)*

A prostaciclina é o principal metabólito do ácido araquidônico produzido pelas ciclooxigenases em células endoteliais (MONCADA & VANE, 1979). Ao ativar os receptores IP no músculo liso vascular produz relaxamento, quando em vasos saudáveis (EDWARDS *et al.*, 2010). Dependendo da artéria, uma hiperpolarização pode ocorrer, a qual envolve a abertura de um ou mais tipos de canais para potássio (FÉLÉTOU & VANHOUTTE, 2006b).

Esta prostaglandina apresenta atividade vasodilatadora e antiagregante plaquetária (RAMLI *et al.*, 2011). Através da estimulação dos receptores IP, a PGI<sub>2</sub> promove a ativação da adenilil cilase levando ao aumento dos níveis de AMPc, este, por sua vez induz a ativação da PKA, a qual induz inibição da contração (HATHAWAY *et al.*, 1981) (Figura 2). No entanto, estudos recentes demonstram que a PGI<sub>2</sub> é capaz também de induzir vasoconstrição, a qual é mediada através dos receptores TP (GLUAIS *et al.*, 2005, XAVIER *et al.*, 2009) (Figura 1). Através de sua ação nesse receptor, este prostanóide tem sido identificado como um importante contribuinte para a disfunção endotelial em artérias de animais normotensos *Wistar Kyoto* e hipertensos SHR tratados com aldosterona (BLANCO-RIVERO *et al.*, 2005; XAVIER *et al.*, 2008). Assim, dependendo das circunstâncias, esta prostaglandina pode ser considerada um EDCF. Somado a isto, hoje se sabe que a prostaciclina é capaz de regular as ações cardiovasculares do tromboxano A<sub>2</sub>, graças a uma interrelação entre os receptores TP e IP ainda não elucidada (FETALVERO *et al.*, 2007).

#### *Prostaglandina F<sub>2α</sub> (PGF<sub>2α</sub>)*

A PGF<sub>2α</sub> participa da modulação do tônus vascular elevando os níveis intracelulares de cálcio no músculo liso vascular promovendo contração

(ABRAMOVITZ *et al.*, 1994). O receptor FP está acoplado à proteína G<sub>q</sub>, e quando estimulado ativa a via do IP<sub>3</sub>/ Ca<sup>2+</sup> produzindo contração da musculatura lisa (PIERCE *et al.*, 1999). Sua ação vasoconstritora também pode ser mediada através de sua ligação aos receptores TP (WONG *et al.*, 2009) (Figuras 1 e 2).

Kimura *et al.* (1994) demonstraram um aumento da resposta contrátil induzida pela PGF<sub>2α</sub> em artérias e veias mesentéricas de resistência, na condição de diabetes, sendo este efeito relacionado à microangiopatia nesses leitos. Corroborando com estes dados, Georgescu & Popov (2003) demonstraram que aumentos nos níveis plasmáticos de glicose, colesterol ou proteína C reativa estão associados ao aumento da resposta vasoconstritora ao prostanoide. Adicionalmente, um outro estudo (YU *et al.*, 2009) demonstraram haver uma relação entre a ativação dos receptores FP e o surgimento de hipertensão arterial e aterosclerose. Portanto, é possível concluir que um aumento na produção deste eicosanóide pode estar envolvido na gênese do processo hipertensivo, bem como das comorbidades relacionadas.

## **2.5. Hipótese de trabalho**

Inicialmente, grande parte dos estudos sobre a “programação fetal” dava maior ênfase ao efeito da desnutrição materna sobre o surgimento de algumas doenças na vida adulta (LAW *et al.*, 2002; SILVERMAN *et al.*, 1991). Entretanto, hoje existem inúmeros trabalhos demonstrando que a exposição fetal à hiperglicemia materna tem uma contribuição importante para o aparecimento de distúrbios metabólicos, neurais e principalmente cardiovasculares no período pós-natal (BRINCIOTTI *et al.*, 2011; HAY, 2012; NOLD & GEORGIEFF, 2004).

Com o objetivo de investigar o impacto do diabetes materno sobre o sistema cardiovascular da prole, vários estudos têm sido realizados (RADENKOVIC *et al.*, 2009; STANLEY *et al.*, 2009; WICHI *et al.*, 2005). Embora tenha sido encontrada uma relação positiva direta entre o ambiente intrauterino hiperglicêmico e o aumento da susceptibilidade para complicações cardiovasculares na vida adulta (CAVANAL *et al.*, 2007; ROCHA *et al.*, 2005; SEGAR *et al.*, 2010), estudos sobre a contribuição do sistema vascular, incluindo a participação de mecanismos endoteliais e componentes da maquinaria contrátil do músculo liso ainda são muito escassos.

Nos últimos anos, estudos com vasos de resistência têm obtido maior destaque, principalmente no que diz respeito aos processos envolvidos no

estabelecimento/ manutenção da pressão arterial elevada e da disfunção endotelial decorrentes de estados hiperglicêmicos (RIZZONI *et al.*, 2011; SCHIFFRIN, 2012). Tal importância deve-se ao fato de que esse tipo de artérias representam, na patogênese de muitas doenças cardiovasculares e em particular na hipertensão arterial, um papel determinante para o desenvolvimento e progressão dos distúrbios observados (JIN *et al.*, 2011; MULVANY, 1987), e alguns autores têm inclusive sugerido que o desenvolvimento de anormalidades nestes vasos poderia ser a causa primária do processo hipertensivo (DENG *et al.*, 1995; MULVANY, 2012).

Sendo assim, o efeito da hiperglicemia materna sobre as artérias de resistência da prole adulta pode representar uma das causas de desenvolvimento de doenças cardiovasculares nesses indivíduos, fato que torna de grande relevância o entendimento dos mecanismos responsáveis por estas alterações.

### **3. OBJETIVOS**

Avaliar o papel da idade e das prostaglandinas derivadas da ciclooxigenase sobre as possíveis alterações do relaxamento dependente do endotélio e da resposta contrátil induzida por estimulação alfa-adrenérgica em artérias de resistência da prole de ratas diabéticas tipo-1.

#### 4. REFERÊNCIAS BIBLIOGRÁFICAS

1. ABRAMOVITZ M.; BOIE Y.; NGUYEN T.; RUSHMORE T. H.; BAYNE M. A. Cloning and expression of a cDNA for the human prostanoid FP receptor. *Journal Biological Chemistry*, v. 269, p. 2632–2636, 1994.
2. ACOSTA J. C.; HAAS D. M.; SAHA C. K.; DIMEGLIO L. A.; INGRAM D. A.; HANELINE L. S. Gestational diabetes mellitus alters maternal and neonatal circulating endothelial progenitor cell subsets. *American Journal Obstetric Gynecology*, v. 204, n. 3, p. 254-258, 2011.
3. AMERICAN DIABETES ASSOCIATION. Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care*, v. 27, n. 1, p. 5-10, 2004.
4. AMERICAN DIABETES ASSOCIATION. Gestational Diabetes Mellitus. *Diab Care* v. 26, p. S103–S105, 2003.
5. AERTS L.; HOLEMANS k.; VAN ASSCHE F.A. Maternal diabetes during pregnancy: Consequences for the offspring. *Diabetes/Metabolism Reviews*, v. 6, n. 3, p. 147–167, 1990.
6. AGABITI-ROSEI E. Structural and functional changes of the microcirculation in hypertension: influence of pharmacological therapy. *Drugs*, v. 63, n.1, p. 19-29, 2003.
7. AKERMAN F.; LEI Z.M.; RAO C.V. Human umbilical Cord and fetal membrane co-express leptin and its receptor genes. *Gynecological Endocrinol*. v. 16, p. 299-306, 2002.
8. ALBRECHT S.S.; KUKLINA E.V.; BANSIL P.; JAMIESON D.J.; WHITEMAN M.K.; KOURTIS A.P.; POSNER S.F.; CALLAGHAN W.M.; Diabetes trends among delivery hospitalizations in the U.S., 1994–2004. *Diabetes Care*, v. 33, n. 4, p. 768–773, 2010.
9. ALFRANCA A.; IÑIGUEZ M. A.; FRESCO M.; REDONDO J. M. Prostanoid signal transduction and gene expression in the endothelium: role in cardiovascular diseases. *Cardiovascular Research*, v. 70, n. 3, p. 446-456, 2006.
10. AMRI K.; FREUND N.; VILAR J.; MERLET-BÉNICHOU C.; LELIÈVRE-PÉGORIER M. Adverse effects of hyperglycemia on kidney development in rats: *in vivo* and *in vitro* studies. *Diabetes*, v. 48, n. 11, p. 2240-2245, 1999.

11. BAGI Z.; ERDEI N.; TOTH A.; LI W.; HINTZE T. H.; KOLLER A. Type 2 diabetic mice have increased arteriolar tone and blood pressure: enhanced release of COX-2-derived constrictor prostaglandins. *Arteriosclerosis, thrombosis, and vascular biology*, v. 25, p. 1610–1616, 2005.
12. BARKER D. J. P. *Mothers, Babies, and Disease in Later Life*. British Medical Association, 1994.
13. BARKER D. J. In utero programming of chronic disease. *Clinical science*, v. 95, p. 115-128, 1998.
14. BARKER D. J.; OSMOND C. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. *Lancet*, v. 1, p. 1077-1081, 1986.
15. BARKER D. J.; OSMOND C.; GOLDING J.; KUH D.; WADSWORTH M. E. Growth in utero, blood pressure in childhood and adult life, and mortality from cardiovascular disease. *British medical journal / British Medical Association*, v. 298, p. 564-567, 1989.
16. BARTON M. The discovery of endothelium-dependent contraction: the legacy of Paul M. Vanhoutte. *Pharmacological Research*, v. 63, n. 6, p. 455-462, 2011.
17. BELIN DE CHANTEMÈLE E. J.; VESSIÈRES E.; GUIHOT A. L.; TOUTAIN B.; MAQUIGNAU M.; LOUFRANI L.; HENRION D. Type 2 diabetes severely impairs structural and functional adaptation of rat resistance arteries to chronic changes in blood flow. *Cardiovascular Research* v. 81, n. 4, p.788-796, 2009.
18. BIAN, K.; DOURSOUT F. Vascular system: role of nitric oxide in cardiovascular diseases. *Journal Clinical Hypertension*, v.10, n.4, p.304-310, 2008.
19. BLANCO-RIVERO J.; CACHOFEIRO V.; LAHERA V.; ARAS-LOPEZ R.; MÁRQUEZ-RODAS I.; SALAICES M. Participation of prostacyclin in endothelial dysfunction induced by aldosterone in normotensive and hypertensive rats. *Hypertension*, v. 46, p. 107–112, 2005.

20. BLONDEAU B.; AVRIL I.; DUCHENE B.; BREANT B. Endocrine pancreas development is altered in foetuses from rats previously showing intra-uterine growth retardation in response to malnutrition. *Diabetologia* v.45, p. 394–401, 2002.
21. BLONDEAU B.; JOLY B.; PERRET C.; PRINCE S.; BRUNEVAL P.; LELIÈVRE-PÉGORIER M.; FASSOT C.; DUONG VAN HUYEN J. P. Exposure in utero to maternal diabetes leads to glucose intolerance and high blood pressure with no major effects on lipid metabolism. *Diabetes Metabolism Journal*, v. 37, n. 3, p. 245-51, 2010.
22. BOLOKER J.; GERTZ S. J.; SIMMONS R. A. Gestational diabetes leads to the development of diabetes in adulthood in the rat. *Diabetes* v. 51, p. 1499–1506, 2002.
23. BORDERS J. L.; GRANGER H. J. Power dissipation as a measure of peripheral resistance in vascular networks. *Hypertension*, v. 8, p. 184–191, 1986.
24. BREYER R. M.; BAGDASSARIAN C. K.; MYERS S. A.; BREYER M. D. Prostanoid receptors: subtypes and signaling. *Annual review of pharmacology and toxicology*, v. 41, p. 661–690, 2001.
25. BRINCIOTTI M.; NAPOLI A.; MITTICA A.; BITTERMAN O.; MATRICARDI M. Cortical evoked potentials in children of diabetic mothers. *Experimental Diabetes Research*, 2011.
26. BRODY S. C.; HARRIS R.; LOHR K. Screening for gestational diabetes: a summary of the evidence for the U.S. Preventive Services Task Force. *Obstetrics and Gynecology*, v. 101, n. 2, p. 380-392, 2003.
27. BROUWERS O.; NIJESSEN P. M.; HAENEN G.; MIYATA T.; BROWNLEE M.; STEHOUWER D. C.; MEY J. G.; SCHALKWIJK C. G. Hyperglycaemia-induced impairment of endothelium-dependent vasorelaxation in rat mesenteric arteries is mediated by intracellular methylglyoxal levels in a pathway dependent on oxidative stress. *Diabetologia*, v. 53, p. 989–1000, 2010.
28. BUCALA R.; TRACEY K. J.; CERAMI A. Advanced glycosylation products quench nitric oxide and mediate defective endothelium-dependent vasodilatation in experimental diabetes. *Journal of Clinical Investigation*, v. 87, n. 2, p. 432– 438, 1991.

29. BUSH N. C.; CHANDLER-LANEY P. C.; ROUSE D. J.; GRANGER W. M.; OSTER R. A.; GOWER B.A. Higher maternal gestational glucose concentration is associated with lower offspring insulin sensitivity and altered beta-cell function. *Journal of clinical endocrinology and metabolism*, v. 96, n. 5, p. 803-809, 2011.
30. BUZZARD C. J.; PFISTER S. L.; CAMPBELL W. B. Endothelium-dependent contractions in rabbit pulmonary artery are mediated by thromboxane A2. *Circulation Research*, v. 72, n. 5, p. 1023-1034, 1993.
31. CANAVAN J. P.; GOLDSPINK D. F. Maternal diabetes in rats. II. Effects on fetal growth and protein turnover. *Diabetes*, v. 37, n. 12, p. 1671-1677, 1988.
32. CARRAPATO M. R.; MARCELINO F. The infant of the diabetic mother. The critical developmental windows. *Early Pregnancy*, v. 5, p. 57-58, 2001.
33. CASEY B. M.; LUCAS M. J.; MCINTIRE D.; LEVENO K. J. Pregnancy outcomes in women with gestational diabetes compared with the general obstetric population. *Obstetrics and Gynecology*, v. 90, n. 6, p. 869-873, 1997.
34. CAVANAL M. E. F.; GOMES G. N.; FORTI A. L.; ROCHA S. O.; FRANCO M. C.; FORTES Z. B.; GIL F. Z. The influence of L-arginine on blood pressure, vascular nitric oxide and renal morphometry in the offspring from diabetic mothers. *Pediatric Research*, v. 62, p. 145-150, 2007.
35. CHANG C. M.; HSIEH C. J.; HUANG J. C.; HUANG I. C. Acute and chronic fluctuations in blood glucose levels can increase oxidative stress in type 2 diabetes mellitus. *Acta Diabetology*, 2012.
36. CHEN G.; YAMAMOTO Y.; MIWA K.; SUZUKI H. Hyperpolarization of arterial smooth muscle induced by endothelial humoral substances. *American Journal Physiology*, v. 260, n. 6, p. 1888-1892, 1991.
37. CHRISTENSEN K. L.; MULVANY M. J. Vasodilatation, not hypotension, improves resistance vessel design during treatment of essential hypertension: a literature survey. *Journal Hypertension*, v. 19, n. 6, p. 1001-1006, 2001.

38. CHYU K. Y.; PETER A.; SHAH P. K. Progress in HDL-based therapies for atherosclerosis. *Current Atherosclerosis Report*, v. 13, n. 5, p. 405-412, 2011.
39. CLARK S. G; FUCHS L. C. Role of nitric oxide and Ca<sup>2+</sup>-dependent K<sub>1</sub> channels in mediating heterogeneous microvascular responses to acetylcholine in different vascular beds. *Journal of pharmacology and experimental therapeutics*, v.282, p. 1473–1479, 1997.
40. CLAUSEN T. D.; MATHIESEN E. R.; HANSEN T. Overweight and the metabolic syndrome in adult offspring of women with diet-treated gestational diabetes mellitus or type 1 diabetes. *Journal Clinical Endocrinology Metabolism*, v.94, p. 2464–2470, 2009
41. COLEMAN R. A.; SMITH W. L.; NARUMIYA S. International Union of Pharmacology classification of prostanoid receptors: properties, distribution, and structure of the receptors and their subtypes. *Pharmacology Review*, v. 46, p. 205-229, 1994.
42. COX N. J. "Maternal component in NIDDM transmission: how large an effect?" *Diabetes*, v. 43, n. 1, p. 166–168, 1994.
43. CUGNET-ANCEAU C.; BA/UDUCEAU B. Glycaemic control and cardiovascular morbi-mortality: the contribution of the 2008 studies. *Ann Endocrinol (Paris)*, v. 70, p. 48–54, 2009.
44. DABELEA D.; HANSON R. L.; BENNETT P. H.; ROUMAIN J.; KNOWLER W. C.; PETTITT D. J. Increasing prevalence of type II diabetes in American Indian children. *Diabetologia*, v. 41, p. 904– 910, 1998.
45. DASKALAKIS G.; MARINOPoulos S.; KRIELESI V.; PAPAPANAGIOTOU A.; PAPANTONIOU N.; MESOGITIS S.; ANTSAKLIS A. Placental pathology in women with gestational diabetes. *Acta obstetricia et gynecologica Scandinavica*, v. 87, p. 403-407, 2008.
46. DENG L. Y.; LI J. S; SCHIFFRIN E. L. Endothelium-dependent relaxation of small arteries from essential hypertensive patients: mechanisms and comparison with normotensive subjects and with responses of vessels from spontaneously hypertensive rats. *Clinical science*, v. 88, p. 611–622, 1995.

47. DE QUEIROZ, DIEGO BARBOSA. Alterações vasculares em ratos expostos ao diabetes materno: Contribuição das prostaglandinas derivadas da COX-2 e sua repercussão em diferentes idades. Recife: UFPE, 2010. Dissertação (mestrado) – Programa de Pós-Graduação em Bioquímica e Fisiologia, Universidade Federal de Pernambuco, Recife, 2010.
48. DE VRIESE A. S.; VERBEUREN T. J.; VAN DE VOORDE J.; LAMEIRE N. H.; VANHOUTTE P. M. Endothelial dysfunction in diabetes. *British Journal Pharmacology*, v. 130, p.963–974, 2000.
49. DE WITT D. L.; DAY J. S.; SONNENBURG W. K.; SMITH W. L. Concentrations of prostaglandin endoperoxide synthase and prostaglandin I<sub>2</sub> synthase in the endothelium and smooth muscle of bovine aorta. *Journal Clinical Invest* v. 72, p. 1882–1888, 1983.
50. DUNNE F.; BRYDON P.; SMITH K.; GEE H. Pregnancy in women with Type 2 diabetes: 12 years outcome data 1990– 2002. *Diabetic Medicine*, v. 20, n. 9, p. 734–738, 2003.
51. EDWARDS G.; FÉLÉTOU M.; WESTON A. H. Endothelium-derived hyperpolarising factors and associated pathways: a synopsis. *Pflügers Archiv : European journal of physiology*, v. 459, n. 6, p. 863-879, 2010.
52. FAGAN T. C.; DEEDWANIA P. C. The cardiovascular dysmetabolic syndrome. *American Journal Medicine*, v. 105, p. 77S-82S, 1998.
53. FÉLÉTOU M.; VANHOUTTE P. M (a). Endothelium-derived hyperpolarizing factor: where are we now? *Arteriosclerosis, thrombosis, and vascular biology*, v. 26, n. 6, p. 1215–1225, 2006.
54. FÉLÉTOU M.; VANHOUTTE P. M. (b) EDHF: the complete story. Boca Raton, Taylor & Francis CRC press, p. 1–298, 2006.
55. FÉLÉTOU M.; HUANG Y.; VANHOUTTE P. M. Vasoconstrictor prostanoids. *Pflugers Archviev*, v.459, n. 6, p. 941-950, 2010.
56. FÉLÉTOU M.; VERBEUREN T. J.; VANHOUTTE P. M. Endothelium-dependent contractions in SHR: a tale of prostanoid TP and IP receptors. *Briitish Journal Pharmacology*, v. 156, p. 563–574, 2009.

57. FETALVERO K. M.; MARTIN K. A.; HWA J. Cardioprotective prostacyclin signaling in vascular smooth muscle. *Prostaglandins & other lipid mediators*, v. 82, p. 109–118, 2007.
58. FETITA L. S.; SOBNGWI E.; SERRADAS P.; CALVO F.; GAUTIER J. F. Consequences of fetal exposure to maternal diabetes in offspring. *Journal Clinical Endocrinology Metabolism*, v. 91, n. 10, p. 3718-3724, 2006.
59. JIN X.; OTONASHI-SATOH Y.; ZAMAMI Y.; KOYAMA T.; SUN P.; KITAMURA Y.; KAWASAKI H. Endothelial modulation of agonist-induced vasoconstriction in mesenteric microcirculation. *Journal of the Pharmaceutical Society of Japan*, v. 130, n. 5, p. 723-728, 2010.
60. FONLUPT P.; CROSET M.; LAGARDE M. 12-HETE inhibits the binding of PGH<sub>2</sub>/TXA<sub>2</sub> receptor ligands in human platelets. *Thrombosis research*, v. 63, p. 239–248, 1991.
61. FRANKS P. W.; LOOKER H. C.; KOBES S.; TOUGER L.; TATARANNI P. A.; HANSON R. L.; KNOWLER W. C. Gestational glucose tolerance and risk of type 2 diabetes in young Pima Indian offspring. *Diabetes*, v. 55, p. 460–465, 2006.
62. FREINKEL N. Of pregnancy and progeny. *Diabetes* v. 29, p. 1023–1035, 1980.
63. FUNK C. D.; FURCI L.; FITZGERALD G. A.; GRYGORCZYK R.; ROCHELLE C.; BAYNE M. A.; ABRAMOVITZ M.; ADAM M.; METTERS K. M. Cloning and expression of a cDNA for the human prostaglandin E receptor EP<sub>1</sub> subtype. *Journal Biology Chemistry*, v. 268, p. 26767-26772, 1993.
64. FURCHGOTT R. F. Role of endothelium in responses of vascular smooth muscle. *Circulation Research*, v. 53, n. 5, p. 557-573, 1983.
65. GARAVITO R. M.; DE WITT D. L. The cyclooxygenase isoforms: Structural insights into the conversion of arachidonic acid to prostaglandins. *Acta biochimica et biophysica Sinica*, v. 1441, p. 278-287, 1999.
66. GATFORD K. L.; SIMMONS R. A.; DE BLASIO M. J.; ROBINSON J. S; OWENS J. A. Review: placental programming of postnatal diabetes and impaired insulin action after IUGR. *Placenta*, v.31, p. 60–65, 2010.

67. GELARDI N. L.; CHA C. M.; OH W. Glucose metabolism in adipocytes of obese offspring of mild hyperglycemic rats. *Pediatric Research*, v. 28, p. 641–645, 1990.
68. GEORGESCU A.; POPOV D. The contractile response of the mesenteric resistance arteries to prostaglandin F<sub>2alpha</sub>; effects of simultaneous hyperlipemia-diabetes. *Fundamental & clinical pharmacology*, v. 17, n. 6, p. 683-689, 2003.
69. GERBER R. T.; HOLEMANS K.; O'BRIEN-COKER I.; MALLET A. I.; VAN BREE R.; VAN ASSCHE F. A.; POSTON, L. Increase of the isoprostane 8-isoprostaglandin F<sub>2α</sub> in maternal and fetal blood of rats with streptozotocin-induced diabetes: Evidence of lipid peroxidation. *American Jornal Obstetricia Gynecology*, v. 183, p. 1035–1040, 2000.
70. GILL R. M.; BRAZ J. C.; JIN N.; ETGEN GJ, SHEN W. Restoration of impaired endothelium-dependent coronary vasodilation in failing heart: role of eNOS phosphorylation and cGMP/cGK-I signaling. *American Journal Physiology Heart Circulation Physiology*, v. 292, n. 6, p. 2782-2790, 2007.
71. GILL-RANDALL R.; ADAMS D.; OLLERTON R. L.; LEWIS M.; ALCOLADO J. C. Type 2 diabetes mellitus—genes or intrauterine environment? An embryo transfer paradigm in rats. *Diabetologia*, v. 47, p. 1354–1359, 2004.
72. GLUAIS P.; LONCHAMPT M.; MORROW J. D.; VANHOUTTE P. M.; FÉLÉTOU M. Acetylcholine-induced endothelium-dependent contractions in the SHR aorta: the Janus face of prostacyclin. *British Journal Pharmacology* v. 146, p. 834–845, 2005.
73. GRASSI G.; DIEZ J. Obesity-related cardiac and vascular structural alterations: beyond blood pressure overload. *Journal Hypertension*, v.27, p. 1750–1752, 2009.
74. GREENSTEIN A. S.; PRICE A.; SONOYAMA K.; PAISLEY A.; KHAVANDI K.; WITHERS S. Eutrophic remodeling of small arteries in type 1diabetes mellitus is enabled by metabolic control: a 10-year follow-up study. *Hypertension*, v. 54, p. 134–141, 2009.
75. GRILL V.; JOHANSSON B.; JALKANEN P.; ERIKSSON U. J. Influence of severe diabetes mellitus early in pregnancy in the rat: effects on insulin sensitivity and insulin secretion in the offspring. *Diabetologia*, v.34, p. 373-378, 1991.

- 76.GUAN Y, ZHANG Y, WU J, QI Z, YANG G, DOU D. Antihypertensive effects of selective prostaglandin E2 receptor subtype 1 targeting. *J Clin Invest* 2007;117:2496–2505.
- 77.GUO Z, SU W, ALLEN S, PANG H, DAUGHERTY A, SMART E. COX-2 up-regulation and vascular smooth muscle contractile hyperreactivity in spontaneous diabetic db/db mice. *Cardiovasc Res* 2005;67:723–735.
- 78.GUZMÁN-GUTIÉRREZ E.; ABARZÚA F.; BELMAR C.; NIEN J. K.; RAMÍREZ M. A.; ARROYO P.; SALOMÓN C.; WESTERMEIER F.; PUEBLA C.; LEIVA A.; CASANELLO P.; SOBREVIA L. Functional link between adenosine and insulin: a hypothesis for fetoplacental vascular endothelial dysfunction in gestational diabetes. *Current vascular pharmacology*, 2011.
- 79.HADI H. A.; SUWAIDI J. A. Endothelial dysfunction in diabetes mellitus. *Vascular health and risk management*, v. 3, n. 6, p. 853-876, 2007.
- 80.HAIDARA M. A.; YASSIN H. Z.; RATEB M.; AMMAR H.; ZORKANI M. A. Role of Oxidative Stress in Development of Cardiovascular Complications in Diabetes Mellitus. *Current Vascular Pharmacology*, v. 4, n. 3, p. 215-227, 2006.
- 81.HAMBERG M.; SAMUELSSON B. Detection and isolation of an endoperoxide intermediate in prostaglandin biosynthesis. *Proceedings of the National Academy of Sciences of the United States of America*, v. 70, p. 899–903, 1973.
- 82.HAMILTON C. A.; BROSNAN M. J.; MCINTYRE M.; GRAHAM D.; DOMINICZAK A. F. Superoxide excess in hypertension and aging: a common cause of endothelial dysfunction. *Hypertension*, v. 37, p. 529–534, 2001.
- 83.HAO L.; NISHIMURA T.; WO H.; FERNANDEZ-PATRON C. Vascular responses to alpha1-adrenergic receptors in small rat mesenteric arteries depend on mitochondrial reactive oxygen species. *Arteriosclerosis, thrombosis, and vascular biology*, v.26, p. 819–825, 2006.
- 84.HATHAWAY D. R.; EATON C. R.; ADELSTEIN R. S. Regulation of human platelet myosin light chain kinase by the catalytic subunit of cyclic AMP-dependent protein kinase. *Nature*, v. 291, p. 252-256, 1981.

85. HAY W. W Jr. Care of the infant of the diabetic mother. *Currente Diabetes Reports*, v. 2, n. 1, p. 4-15, 2012.
86. HEAGERTY A. M. Predicting hypertension complications from small artery structure. *Journal Hypertension*, v. 25, p. 939–940, 2007.
87. HINK U.; LI H.; MOLLNAU H.; OELZE M.; MATHEIS E.; HARTMANN M.; SKATCHKOV M.; THAISS F.; STAHL R. A.; WARNHOLTZ A.; MEINERTZ T.; GRIENDLING K.; HARRISON D. G.; FORSTERMANN U.; MUNZEL T. Mechanisms underlying endothelial dysfunction in diabetes mellitus. *Circulation Research*, v. 88, n. 2, p. E14–E22, 2001.
88. HIRATA T.; USHIKUBI F.; KAKIZUKA A.; OKUMA M.; NARUMIYA S. Two thromboxane A<sub>2</sub> receptor isoforms in human platelets. Opposite coupling to adenylate cyclase with different sensitivity to Arg60 to Leu mutation. *Journal Clinical Investigation*, v. 97, p. 949–956, 1996.
89. HITOMI H.; KAIFU K.; FUJITA Y.; SOFUE T.; NAKANO D.; MORIWAKI K.; HARA T.; KIYOMOTO H.; KOHNO M.; KOBORI H.; NISHIYAMA A. Angiotensin II shifts insulin signaling into vascular remodeling from glucose metabolism in vascular smooth muscle cells. *American Journal Hypertension*, v. 24, n.10, p. 1149-1155, 2011.
90. HOLEMANS K.; GERBER R. T.; MEURRENS K.; DE CLERCK F.; POSTON L.; VAN ASSCHE F. A. Streptozotocin diabetes in the pregnant rat induces cardiovascular dysfunction in adult offspring. *Diabetologia* v.42, p. 81-89, 1999.
91. HOROWITZ A.; MENICE C. B.; LAPORTE R.; MORGAN K. G. Mechanisms of smooth muscle contraction. *Physiology Review*, v. 76, p. 967-1003, 1996.
92. HRISTOVSKA A. M.; RASMUSSEN L. E.; HANSEN P. B.; NIELSEN S. S.; NÜSING R. M.; NARUMIYA S. Prostaglandin E2 induces vascular relaxation by E-prostanoid 4 receptor-mediated activation of endothelial nitric oxide synthase. *Hypertension*, v. 50, p. 525–530, 2007.
93. HUXLEY R. R.; SHIELL A. W.; LAW C. M. The role of size at birth and postnatal catch-up growth in determining systolic blood pressure: a systematic review of the literature. *Journal Hypertension*, v. 18, n. 7, p. 815-831, 2000.

94. INGRAM D. A.; LIEN I. Z.; MEAD L. E.; ESTES M.; PRATER D. N.; DERR-YELLIN E.; DIMEGLIO L. A; HANELINE L. S. In vitro hyperglycemia or a diabetic intrauterine environment reduces neonatal endothelial colony-forming cell numbers and function. *Diabetes*, v. 57, p. 724–731, 2008.
95. JAKUS V.; RIETBROCK N. Advanced glycationend-products and the progress of diabetic vascular complications. *Physiology Research*, v. 53, p. 131-142, 2004.
96. JIN X.; SATOH-OTONASHI Y.; ZAMAMI Y.; TAKATORI S.; HASHIKAWA-HOBARA N.; KITAMURA Y.; KAWASAKI H. New molecular mechanisms for cardiovascular disease: contribution of endothelium-derived hyperpolarizing factor in the regulation of vasoconstriction in peripheral resistance arteries. *Journal Pharmacology Science*, v. 116, n. 4, p. 332-336, 2011.
97. JUNOD A.; LAMBERT E. A.; STAUFFACHER W.; RENOLD A. E. Diabetogenic action of streptozotocin: relationship of dose to metabolic response. *Journal of Clinical Investigation*, v. 48, n. 11, p. 2129–2139, 1969.
98. KALOUSOVÁ M.; KRHA J.; ZIMA T. Advanced glycation end-products and advanced oxidation protein products in patients with diabetes Mellitus. *Physiology Research*, v. 51, p. 597-604, 2002.
99. KIM S. Y.; ENGLAND L.; WILSON H. G.; BISH C. A.; SATTEN G. A.; DIETZ P. (a) Percentage of gestational diabetes mellitus attributable to overweight and obesity. *American Journal of Public Health*, v. 100, n. 6, p. 1047–1052, 2010.
100. KIM S. Y.; ENGLAND J. L.; SHARMA J. A.; NJOROGE T. (b) Gestational diabetes mellitus and risk of childhood overweight and obesity in offspring: a systematic review. *Experimental Diabetes Reserach*, 2011.
101. KIMURA I.; HATA Y.; ISLAM M. A.; KIMURA M. Diabetes mellitus-induced enhancement of prostaglandin F2 alpha-responses is inhibited by lipoxygenase- but not cyclooxygenase-inhibitors in mesenteric veins and arteries of mouse and rat. *Japanese journal of pharmacology*, v. 64, p. 65–70, 1994.
102. KITZMILLER J. L.; BLOCK J. M.; BROWN F. M.; CATALANO P. M.; CONWAY D. L.; COUSTAN D. R.; GUNDERSON E. P.; HERMAN W. H.;

HOFFMAN L. D.; INTURRISI M.; JOVANOVIC L. B.; KJOS S.; KNOPP R. H.; MONTORO M. N.; OGATA E. S.; PARAMSOTHY P.; READER D. M.; ROSENN B. M.; THOMAS A. M.; KIRKMAN M. S. Managing preexisting diabetes for pregnancy: summary of evidence and consensus recommendations for care. *Diabetes Care*, v. 31, n. 5, p. 1060–1079, 2008.

103. LAGAUD G. J.; SKARSGARD P. L.; LAHER I.; VAN BREEMEN C. Heterogeneity of endothelium-dependent vasodilation in pressurized cerebral and small mesenteric resistance arteries of the rat.. *Journal Pharmacology and Experimental Therapeutics. Journal Pharmacology Experimental Therapy*. v. 290, n. 2, p. 832-9, 1999.
104. LAPPAS M.; YEE K.; PERMEZEL M.; RICE G. Relase amd regulation of leptin, resistin and adiponectin from human placenta, fetal membranes, ans maternal adipose tissue and skeletal muscle from normal and gestational diabetes mellitus complicated pregnancies. *Journal Endocrinology*, v. 186, p. 457-465, 2005.
105. LARGER E.; MARRE M.; CORVOL P.; GASC J. M. Hyperglycemia-induced defects in angiogenesis in the chicken chorioallantoic membrane model. *Diabetes*, v.53, p. 752–61, 2004.
106. LAW C. M. Fetal and infant influences on non-insulin-dependent diabetes mellitus (NIDDM). *Diabetic Medicine*,v. 13, n. 9,p. 49-52, 1996.
107. LAW C. M. Significance of birth weight for the future. *Archives of disease in childhood. Fetal and neonatal edition*, v. 86, p. 7-8, 2002.
108. LEON D. A.; LITHELL H. O.; VAGERÖ D.; KOUPILOVÁ I.; MOHSEN R.; BERGLUND L.; LITHELL U-B.; MCKEIGUE P. M. Reduced fetal growth rate and increased risk of death from ischaemic heart disease: cohort study of 15,000 Swedish men and women born 1915-1929. *British Medical Journal*, v. 317, p. 241–245, 1998.
109. LEVY B. I.; AMBROSIO G.; PRIES A. R. Microcirculation in hypertension: a new target for treatment? *Circulation*, v. 104:735–740, 2001.
110. LEVY B. I.; SCHIFFRIN E. L.; MOURAD J. J.; AGOSTINI D.; VICAUT E.; SAFAR M. E.; STRUIJKER-BOUDIER H. A. Impaired tissue perfusion: a pathology common to hypertension, obesity, and diabetes mellitus. *Circulation*, v. 118, p. 968–976, 2008.

111. LINDSAY R. S.; DABELEA D.; ROUMAIN J.; HANSON R. L.; BENNETT P. H.; KNOWLER W. C. Type 2 diabetes and low birth weight: the role of paternal inheritance in the association of low birth weight and diabetes. *Diabetes*, v. 49, p. 445–449, 2000.
112. LIU Y.; LI H.; BUBOLZ A. H.; ZHANG D. X.; GUTTERMAN D. D. Endothelial cytoskeletal elements are critical for flow-mediated dilation in human coronary arterioles. *Medical & biological engineering & computing*, v. 46, p. 469–78, 2008.
113. LIU C. Q.; LEUNG F. P.; WONG S. L.; WONG W. T.; LAU C. W.; LU L. Thromboxane prostanoid receptor activation impairs endothelial nitric oxide-dependent vasorelaxations: the role of Rho kinase. *Biochemistry Pharmacology*, v. 78, p. 374–381, 2009.
114. MAGANHA C. A.; VANNI D. G. B. S.; BERNARDINI M. A.; ZUGAIB M. Tratamento do Diabetes Melito Gestacional. *Revista da Associação Médica Brasileira*, v. 49, p. 330–334, 2003.
115. MANDERSON J. G.; MULLAN B.; PATTERSON C. C.; HADDEN D. R.; TRAUB A. I.; CANCE D. R. Cardiovascular and metabolic abnormalities in the offspring of diabetic pregnancy. *Diabetologia*, v. 45, p. 991–996, 2002.
116. MAYEUX P. R.; MAIS D. E.; CARR C.; HALUSHKA P. V. Human erythroleukemia cells express functional thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptors. *Journal Pharmacology Experimental Therapy*, v. 250, p. 923–927, 1989.
117. MAZZONE T.; CHAIT A.; PLUTZKY J. Cardiovascular disease risk in type 2 diabetes mellitus: insights from mechanistic studies. *Lancet*, v. 371, p. 1800–1809, 2008.
118. METZGER B. E.; LOWE L. P.; DYER A. R.; TRIMBLE E. R.; CHAOVARINDR U.; COUSTAN D. R.; HADDEN D. R.; MCCANCE D. R.; HOD M.; MCINTYRE H. D.; OATS J. J.; PERSSON B.; ROGERS M. S.; SACKS D. A. Hyperglycemia and adverse pregnancy outcomes. *The New England journal of medicine*, v. 358, p. 1991–2002, 2008.
119. MILLER V. M.; VANHOUTTE P. M. Endothelium-dependent contractions to arachidonic acid are mediated by products of cyclooxygenase. *American Journal Physiology*, v. 248, p.H432-437, 1985.

120. MONCADA S.; VANE J. R. Pharmacology and endogenous roles of prostaglandin endoperoxides, thromboxane A<sub>2</sub> and prostacyclin. *Pharmacology Review*, v. 30, p. 293–331, 1979.
121. MORGADO M.; CAIRRÃO E.; SANTOS-SILVA A. J.; VERDE I. Cyclic nucleotide-dependent relaxation pathways in vascular smooth muscle. *Cellular and molecular life sciences*, v. 69, n. 2, p. 247-266, 2012.
122. MORITA I. Distinct functions of COX-1 and COX-2. *Prostaglandins Other Lipids Mediators*. 68,p. 165–175, 2002.
123. MOSES R. G. Gestational diabetes mellitus: implications of an increased frequency with IADPSG criteria. , v. 35, n. 3, p. 461-462, 2012.
124. MULVANY M. J. The fourth Sir George Pickering memorial lecture. The structure of the resistance vasculature in essential hypertension. *Journal Hypertension*, v. 5, n. 2, p. 129-136, 1987.
125. MULVANY M. J. Small artery remodelling in hypertension. Basic and clinical pharmacology and toxicology, v. 110, n. 1, p. 49-55, 2012.
126. MULVANY M. J.; AALKJÆR C. Structure and function of small arteries. *Physiol Review*, v. 70, p. 921–961, 1990.
127. NAKAHATA N. Thromboxane A<sub>2</sub>: physiology/pathophysiology, cellular signal transduction and pharmacology. *Pharmacology Therapy*, v. 118, p. 18–35, 2008.
128. NAKAJIMA H.; ONISHI K.; KURITA T.; ISHIDA M.; NAGATA M.; KITAGAWA K.; DOHI K.; NAKAMURA M.; SAKUMA H.; ITO M. Hypertension impairs myocardial blood perfusion reserve in subjects without regional myocardial ischemia. *Hypertension Research*, v. 33, n. 11, p. 1144-1149, 2010.
129. NOLD J. L.; GEORGIEFF M. K. Infants of diabetic mothers. *Pediatric Clinical North American*, v. 51, n. 3, p. 619-637, 2004.
130. NOMURA Y.; MARKS D. J.; GROSSMAN B.; YOON M.; LOUDON H.; STONE J.; HALPERIN J. M. Exposure to gestational diabetes mellitus and low socioeconomic status: effects on neurocognitive development and risk

of attention-deficit/hyperactivity disorder in offspring. Archives of pediatrics & adolescent medicine, v. 166, n. 4, p. 337-343, 2012.

131. OH W.; GELARDI N. L.; CHA C. M. The crossgeneration effect of neonatal macrosomia in rat pups of streptozotocin induced diabetes. Pediatric Research, v. 29, p. 606–610, 1991.
132. OZYAZGAN S.; UNLUCERCI Y.; BEKPINAR S.; AKKAN A. G. Impaired relaxation in aorta from streptozotocin-diabetic rats: effect of aminoguanidine (AMNG) treatment. International journal of experimental diabetes research, v. 1, n. 2, p. 145-153, 2000.
133. PANNIRSELVAM M.; WIEHLER W. B.; ANDERSON T.; TRIGGLE C. R. Enhanced vascular reactivity of small mesenteric arteries from diabetic mice is associated with enhanced oxidative stress and cyclooxygenase products. British Journal Pharmacology, Archives of pediatrics & adolescent medicine, v. 144, n. 7, p. 953-960, 2005.
134. PARADISO G.; BIAGGI A.; FERRAZZANI S.; DE CAROLIS S.; CARUSO A. Abnormal carbohydrate metabolism during pregnancy: association with endothelial dysfunction. Diabetes Care, v. 25, p. 560–564, 2002.
135. PARAVICINI T. M.; TOUYZ RM. Redox signaling in hypertension. Cardiovascular Research, v. 5, n. 71(2):247-258, 2006.
136. PARENTE L.; PERRETTI M. Advances in the pathophysiology of constitutive and inducible cyclooxygenases: two enzymes in the spotlight. Biochemistry Pharmacology, v. 15, n. 65, p. 153-159, 2003.
137. PEARSON J. D. Lipid metabolism in cultured aortic smooth muscle cells and comparison with other cell types. Part I. Composition of cells grown in hyperlipemic serum. Atherosclerosis, v. 24, n. 2, p. 233-242, 1976.
138. PESSIN J. E.; SALTIEL A. R. Signaling pathways in insulin action: molecular targets of insulin resistance. Journal Clinical Investigation, v. 106, p. 165-169, 2000.
139. PETTITT D. J.; BAIRD H. R.; ALECK K. A.; BENNETT P. H.; KNOWLER W. C. Excessive obesity in offspring of Pima Indian women

with diabetes during pregnancy. *The New England journal of medicine*, v. 308, p. 242–245, 1983.

140. PETTITT D. J.; ALECK K. A.; BAIRD H. R.; CARRAHER M. J.; BENNETT P. H.; KNOWLER W. C. Congenital susceptibility to NIDDM. Role of intrauterine environment. *Diabetes*, v. 37, p. 622–628, 1988.
141. PIERCE K. L.; FUJINO H.; SRINIVASAN D.; REGAN J. W. Activation of FP prostanoid receptor isoforms leads to Rho-mediated changes in cell morphology and in the cell cytoskeleton. *Journal of chemical biology*, v. 10, n. 274, p. 35944-35949, 1999.
142. PINHEIRO A. R.; SALVUCCI I. D.; AGUILA M. B.; MANDARIM-DELACERDA C. A. Protein restriction during gestation and/or lactation causes adverse transgenerational effects on biometry and glucose metabolism in F1 and F2 progenies of rats. *Clinical Science (Lond)*, v. 114, p. 381–392, 2008.
143. PINTER E.; HAIGH J.; NAGY A.; MADRI J. A. Hyperglycemia-induced vasculopathy in the murine conceptus is mediated via reductions of VEGF-A expression and VEGF receptor activation. *American Journal Pathology*, v. 158, p. 1199–1206, 2001.
144. PRATICO D.; LAWSON J. A.; ROKACH J.; FITZGERALD G. A. The isoprostanes in biology and medicine. *Trends in endocrinology and metabolism*, v. 12, p. 243–247, 2001.
145. PRIES A. R.; SECOMB T. W.; GAEHTGENS P. The endothelial surface layer. *Pflugers Arch*, v. 440, n. 5, p. 653-666, 2000.
146. RADENKOVIC M.; RADUNOVIC N.; MOMCILOV P.; GRBOVIC L. Altered response of human umbilical artery to 5-HT in gestational diabetic pregnancy. *Pharmacology Report*, v. 61, p. 520-528, 2009.
147. RAMLI J.; CALDERON ARTERO P.; BLOCK R. C.; MOUSA S. A. Novel therapeutic targets for preserving a healthy endothelium: strategies for reducing the risk of vascular and cardiovascular disease. *Cardiology Journal*, v. 18, n. 4, p. 352-363, 2011.
148. REIHER H.; FUHRMANN K.; NOACK S.; WOLTANSKI K. P.; JUTZI E.; HAHN VON DORSCHE H.; HAHN H. J. Age-dependent insulin secretion of

the endocrine pancreas in vitro from fetuses of diabetic and nondiabetic patients. *Diabetes Care*, v. 6, p. 446–451, 1983.

149. REINKING B. E.; WEDEMEYER E. W.; WEISS R. M.; SEGAR J.L.; SCHOLZ T. D. Cardiomyopathy in offspring of diabetic rats is associated with activation of the MAPK and apoptotic pathways. *Cardiovascular Diabetology*, v. 8, p. 43, 2009.
150. REUSENS B.; REMACLE C. Programming of the endocrine pancreas by the early nutritional environment. *International journal of biochemistry and cell biology*, v. 38, p. 913–922, 2006.
151. RIBEIRO M. C.; NAKAMURA M. U.; ABDO C. H. N.; TORLONI M. R.; M. T. SCANAVINO; MATTAR R. Pregnancy and Gestational Diabetes: a prejudicial combination to female sexual function? *Revista Brasileira Ginecologia e Obstetrícia*, v. 33, n. 5, p. 219-224, 2011.
152. RIZZONI D.; PORTERI E.; GUELFI D.; MUIESAN M. L.; VALENTINI U.; CIMINO A.; GIRELLI A.; RODELLA L.; BIANCHI R.; SLEIMAN I.; ROSEI E. A. Structural alterations in subcutaneous small arteries of normotensive and hypertensive patients with non insulin dependent diabetes mellitus. *Circulation*, v. 103, p. 1238–1244, 2001.
153. RIZZONI D.; DE CIUCEIS C.; PORTERI E.; PAIARDI S.; BOARI G. E.; MORTINI P.; CORNALI C.; CENZATO M.; RODELLA L. F.; BORSANI E.; RIZZARDI N.; PLATTO C.; REZZANI R.; ROSEI E A. Altered structure of small cerebral arteries in patients with essential hypertension. *Journal Hypertension*, v. 27, p. 838–845, 2009.
154. RIZZONI D.; DE CIUCEIS C.; PORTERI E.; SEMERARO F.; ROSEI E A. Structural alterations in small resistance arteries in obesity. *Basic Clinical Pharmacology Toxicology*, v. 110, n. 1, p. 56-62, 2011.
155. ROCHA S. O.; GOMES G. N.; FORTI A. L.; DO CARMO PINHO FRANCO M.; FORTES Z. B.; DE FÁTIMA CAVANAL M.; GIL F. Z. Long-term effects of maternal diabetes on vascular reactivity and renal function in the rat male offspring. *Pediatric Research*, v. 58, p. 1274-1279, 2005.
156. ROGERS LK & VELTEN M. F. Maternal inflammation, growth retardation, and preterm birth: Insights into adult cardiovascular disease. *Life Science*, v. 26, n. 89, p. 417-21, 2011.

157. RUBANYI GM, VANHOUTTE PM. Superoxide anions and hyperoxia inactivate endothelium-derived relaxing factor. *American Journal Physiology*, v. 250, p. H222–H227, 1986.
158. RUDGE M. V.; LIMA C. P.; DAMASCENO D. C.; SINZATO Y. K.; NAPOLI G.; RUDGE C. V.; GALLEGOS F. Q.; CALDERON I. M. Histopathological placental lesions in mild gestational hyperglycemic and diabetic women. *Diabetology Metabolism Syndrome*, v. 10, n. 3, p. 19, 2011.
159. SALOMÓN C.; WESTERMEIER F.; PUEBLA C.; ARROYO P.; GUZMÁN-GUTIÉRREZ E.; PARDO F.; LEIVA A.; CASANELLO P.; SOBREVIA L. Gestational diabetes reduces adenosine transport in human placental microvascular endothelium, an effect reversed by insulin. *PLoS One*, n.7, p. 40578, 2012.
160. SCHIFFRIN E. L. Effect of treatment on cardiac small resistance arteries in hypertension and diabetes: trick or treat. *Journal Hypertension*, v. 30, n. 2, p. 271-272, 2012.
161. SCHMIDT M. I.; MATOS M. C.; REICHELT A. J.; FORTI A. C.; DE LIMA L.; DUNCAN B. B. Prevalence of gestational diabetes mellitus - do the new WHO criteria make a difference? *Brazilian Gestational Diabetes Study Group. Diabetic Medicine*, v. 17, p. 376–380, 2000.
162. SEGAR E. M.; NORRIS A. W.; YAO J. R.; HU S.; KOPPENHAFER S. L.; ROGHAIR R. D.; SEGAR J.L.; SCHOLZ T. D. Programming of growth, insulin resistance and vascular dysfunction in offspring of late gestation diabetic rats. *Clinical Science (Lond)*, v. 117, n. 3, p. 129–138, 2010.
163. SERHAN C. N.; LEVY, B. Success of prostaglandin E2 in structure-function is a challenge for structure-based therapeutics. *Proceedings of the National Academy of Sciences of the USA*, v. 100, n. 15, p. 8609-8611, 2003.
164. SFERRUZZI-PERRI A. N.; OWENS J. A.; PRINGLE K. G.; ROBERTS C. T. The neglected role of insulin-like growth factors in the maternal circulation regulating fetal growth. *Journal of Physiology*, v. 1, n. 589, p. 7-20, 2011.

165. SHI Y.; SO K. F.; MAN R. Y.; VANHOUTTE P. M. Oxygen-derived free radicals mediate endothelium-dependent contractions in femoral arteries of rats with streptozotocin-induced diabetes. *British Journal Pharmacology* 152: 1033–1041, 2007.
166. SHI Y.; VANHOUTTE P. M. Oxidative stress and COX cause hyper-responsiveness in vascular smooth muscle of the femoral artery from diabetic rats. *British Journal Pharmacology*, v. 154, p. 639-651, 2008.
167. SIMEONI U.; BARKER D. J. Offspring of diabetic pregnancy: Long-term outcomes. *Seminars in Fetal & Neo Medicine*, v. 14, p. 119–124, 2009.
168. SIMMONS D. L.; BOTTING R. M. ; TIMOTHY H. L. A. Cyclooxygenase isozymes: the biology of prostaglandin synthesis and inhibition. *Pharmacology Review*, v. 56, n. 3, p. 387-437, 2004.
169. SILVERMAN B. L.; RIZZO T.; GREEN O. C.; CHO N. H.; WINTER R. J.; OGATA E. S.; RICHARDS G. E.; METZGER B. E. Long-term prospective evaluation of offspring of diabetic mothers. *Diabetes*, v. 40, n. 2, p. 121–125, 1991.
170. SMITH W. L.; SONG I. The enzymology of prostaglandin endoperoxide H synthases 1- and –2. *Prostaglandins Other Lipid Mediators*, v. 68–69, p. 115–128, 2002.
171. SOOTHILL P. W. Diagnosis of intrauterine growth retardation and its fetal and perinatal consequences. *Acta Paediatric*, v. 399, p. 55-58, 1994.
172. SOWERS J. R. Relationship between hypertension and subtle and overt abnormalities of carbohydrate metabolism. *Journal American Society Nephrology*, v. 1, p. 39-47, 1990.
173. STANLEY J. L.; ASHTON N.; TAGGART M. J.; DAVIDGE S. T.; BAKER P. N. Uterine artery function in a mouse model of pregnancy complicated by diabetes. *Vascular Pharmacology*, v. 50, n. 2, p. 8-13, 2009.
174. STRIDE A.; SHEPHERD M.; FRAYLING T. M.; BULMAN M. P.; ELLARD S.; HATTERSLEY A. T. Intrauterine hyperglycemia is associated with an earlier diagnosis of diabetes in HNF-1alpha gene mutation carriers. *Diabetes Care*, v. 25, p. 2287–2291, 2002.

175. SUGANAMI T.; MORI K.; TANAKA I.; MUKOYAMA M.; SUGAWARA A.; MAKINO H. Role of prostaglandin E receptor EP1 subtype in the development of renal injury in genetically hypertensive rats. *Hypertension*, v. 42, p. 1183–1190, 2003.
176. SUN D.; HUANG A.; KALEY G. Mechanical compression elicits NOdependent increases in coronary flow. *American Journal Physiology Heart Circulation Physiology*, v. 287, p. 2454–2460, 2004.
177. TADDEI S.; VANHOUTTE P. M. Endothelium-dependent contractions to endothelin in the rat aorta are mediated by thromboxane A<sub>2</sub>. *Journal Cardiovascular Pharmacology*, v. 22, n. 8, p. 328-331, 1993.
178. TAGUCHI K.; MATSUMOTO T.; KAMATA K.; KOBAYASHI T. Akt/eNOS pathway activation in endothelium-dependent relaxation is preserved in aortas from female, but not from male, type 2 diabetic mice. *Pharmacology Research*, v. 65, n. 1, p. 56-65, 2012.
179. TANG E. H.; LEUNG F. P.; HUANG Y.; FÉLÉTOU M.; SO K. F., MAN R. Y. Calcium and reactive oxygen species increase in endothelial cells in response to releasers of endothelium-derived contracting factor. *British Journal Pharmacology*, v. 151, p. 15–23, 2007.
180. TANG E. H.; JENSEN B. L.; SKOTT O.; LEUNG G. P.; FELETOU M.; MAN R. Y.; VANHOUTTE P. M. The role of prostaglandin E and thromboxane-prostanoid receptors in the response to prostaglandin E2 in the aorta of Wistar Kyoto rats and spontaneously hypertensive rats. *Cardiovascular Research*, v. 1, n. 7, p. 130-138, 2008.
181. TARICCO E.; RADAELLI T.; NOBILE DE SANTIS M. S.; CETIN I. Fetal and placental weight in relation to maternal characteristics in gestational diabetes. Histological placental lesions in women with recurrent preterm delivery. *Placenta*, v. 24, p. 343-347, 2003.
182. TAYLOR PD; POSTON L. The effect of hyperglycaemia on function of rat isolated mesenteric resistance artery. *British Journal of Pharmacology*, v. 113, n. 3, p. 801–808, 1994.
183. TOGASHI N.; MAEDA T.; YOSHIDA H.; KOYAMA M.; TANAKA M.; FURUHASHI M.; SHIMAMOTO K.; MIURA T. Angiotensin II receptor

activation in youth triggers persistent insulin resistance and hypertension--a legacy effect? *Hypertension Research*, v. 35, n. 3, p. 334-340, 2012.

184. TOPPER J. N; CAI J.; FALB D.; GIMBRONE M. A. Identification of vascular endothelial genes differentially responsive to fluid mechanical stimuli: cyclooxygenase-2, manganese superoxide dismutase, and endothelial cell nitric oxide synthase are selectively up-regulated by steady laminar shear stress. *Proceedings of the National Academy of Sciences of the USA*, v. 93, p. 10417–10422, 1996.
185. TRACHTE G. J. Prostacyclin mediates arachidonic acid-induced relaxation of rabbit isolated mesenteric arteries. *Journal Cardiovascular Pharmacology*, v. 8, n. 4, p. 758-764, 1986.
186. TSUBOI K.; SUGIMOTO Y.; ICHIKAWA A. Prostanoid receptor subtypes. *Prostaglandins & other lipid mediators*, v. 69, p. 535–556, 2002.
187. VALENTIN F.; FIELD M. C.; TIPPINS J. R. The mechanism of oxidative stress stabilization of the thromboxane receptor in COS-7 cells. *Journal Biology Chemistry*, v. 279, p. 8316–8324, 2004.
188. VANHOUTTE, P. M.; EBER, B. Endothelium-derived relaxing and contracting factors. *Wiener klinische Wochenschrift*, v. 103, n. 14, p. 405-411, 1991.
189. VANHOUTTE P. M.; FÉLÉTOU M.; TADDEI S. Endothelium-dependent contractions in hypertension. *British Journal Pharmacology*, v. 144, p. 449-458, 2005.
190. VAZZANA N.; RANALLI P.; CUCCURULLO C.; DAVÌ G. Diabetes mellitus and thrombosis. *Thrombosis Research*, v. 129, n. 3, p. 371-377, 2012.
191. VILAHUR G.; CASANI L.; BADIMON L. A thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> receptor antagonist (S18886) shows high antithrombotic efficacy in an experimental model of stent-induced thrombosis. *Journal of thrombosis and haemostasis*, v. 98, p. 662–669, 2007.
192. VIRDIS A.; GHIADONI L.; TADDEI S. Human endothelial dysfunction: EDCFs. *European Journal Physiology*, v. 459, p. 1015-1023, 2010.

193. XU D.; CHEN M.; PAN X. L.; XIA L. P.; WANG H. Dexamethasone induces fetal developmental toxicity through affecting the placental glucocorticoid barrier and depressing fetal adrenal function. *Environmental toxicology and pharmacology*, v. 32, n. 3, p. 356-363, 2011.
194. WARD D. T.; YAU S. K.; MEE A. P.; MAWER E. B., MILLER C. A.; GARLAND H. O.; RICCARDI D. Functional, molecular, and biochemical characterization of streptozotocin-induced diabetes. *Journal American Society Nephrology*, v. 12, p. 779–790, 2001.
195. WATKINS M. T.; PATTON G. M.; SOLER H. M.; ALBADAWI H.; HUMPHRIES D. E.; EVANS J. E.; KADOWAKI K. Synthesis of 8-epi-prostaglandinF2 $\alpha$  by human endothelial cells: role of prostaglandin H2 synthase. *Biochemistry Journalal*, v. 344, p. 747–775, 1999.
196. WENCESLAU C. F.; DAVEL A. P.; XAVIER F. E.; ROSSONI L. V. Long-term ouabain treatment impairs vascular function in resistance arteries. *Journal Vascular Research*, v. 48, n. 4, p. 316-26, 2011.
197. WEST N. A.; CRUME T. L.; MALIGIE M. A.; DABELEA D. Cardiovascular risk factors in children exposed to maternal diabetes in utero. *Diabetologia*, v. 54, n. 3, p. 504-507, 2011.
198. WICHI R. B.; SOUZA S. B.; CASARINI D. E.; MORRIS M.; BARRETO-CHAVES M. L.; IRIGOYEN M. C. Increased blood pressure in the offspring of diabetic mothers. *American Journal Physiology Regulation*, v. 288, p. 1129-1133, 2005.
199. WILLIAMS S. A.; BOOLELL M.; MAC GREGOR G. A.; SMAJE L. H.; WASSERMAN S. M.; TOOKE J. E. Capillary hypertension and abnormal pressure dynamics in patients with essential hypertension. *Clinical Science*, v. 79, p. 5-8, 1990.
200. WILSON S. J.; CAVANAGH C. C.; LESHER A. M.; FREY A. J.; RUSSELL S. E.; SMYTH E. M. Activation-dependent stabilization of the human thromboxane receptor: role of reactive oxygen species. *Journal Lipid Research*, v. 50, p. 1047–1056, 2009.

201. WOLIN M. S. Interactions of oxidants with vascular signaling systems. *Arteriosclerosis, thrombosis, and vascular biology*, v. 20, p. 1430–1442, 2000.
202. WONG S. L.; LEUNG F. P.; LAU C. W.; AU C. L.; YUNG L. M.; YAO X.; CHEN Z. Y.; VANHOUTTE P. M.; GOLLASCH M.; HUANG Y. Cyclooxygenase-2-derived prostaglandin F2alpha mediates endothelium-dependent contractions in the aortae of hamsters with increased impact during aging. *Circulation Research*, v. 104, p. 228–235, 2009.
203. XAVIER F. E.; BLANCO-RIVERO J.; FERRER M.; BALFAGÓN G. Endothelium modulates vasoconstrictor response to prostaglandin I2 in rat mesenteric resistance arteries: interaction between EP1 and TP receptors. *British Journal Pharmacology*, v. 158, n. 7, p. 1787-1795, 2009.
204. XAVIER F. E.; DAVEL A. P.; ROSSONI L. V.; VASSALLO D. V. Time-dependent hyperreactivity to phenylephrine in aorta from untreated diabetic rats: role of prostanoids and calcium mobilization. *Vascular Pharmacology*, v. 40, p. 67-76, 2003.
205. XAVIER F. E.; ARAS-LÓPEZ R.; ARROYO-VILLA I.; CAMPO L. D.; SALAICES M.; ROSSONI L. V. Aldosterone induces endothelial dysfunction in resistance arteries from normotensive and hypertensive rats by increasing thromboxane A2 and prostacyclin. *British Journal Pharmacology*, v. 154, p. 1225–1235, 2008.
206. YAGAMI T. Cerebral Arachidonate Cascade in Dementia: Alzheimer's Disease and Vascular Dementia. *Current Neuropharmacology*, 4, n. 1, p. 87–100, 2006.
207. YESSOUFOU A.; MOUTAIROU K. Maternal diabetes in pregnancy: early and long-term outcomes on the offspring and the concept of "metabolic memory". *Experimental Diabetes Research*, 2011.
208. YOU D.; LOUFRANI L.; BARON C.; LEVY B. I.; WIDDOP R. E.; HENRION D. High blood pressure reduction reverses angiotensin II type 2 receptor-mediated vasoconstriction into vasodilation in spontaneously hypertensive rats. *Circulation*, v. 111, n. 8, p. 1006-1011, 2005.
209. YU Y.; LUCITT M. B.; STUBBE J.; CHENG Y.; FRIIS U. G.; HANSEN PB. Prostaglandin F2alpha elevates blood pressure and promotes atherosclerosis. *Proceedings of the National Academy of Sciences of the USA*, v. 106, p. 7985–7990, 2009.

210. ZHANG, M. Z.; WANG, J. L.; CHENG, H. F.; HARRIS, R. C; MCKANNA JA. Cyclooxygenase-2 in rat nephron development. American Journal Physiology, v. 273, p. 994-1002, 1997.

## 5. ARTIGO 1

# RESEARCH PAPER

## Effect of age and COX-2-derived prostanoids on the progression of adult vascular dysfunction in the offspring of diabetic rats

FE Ramos-Alves\*, DB de Queiroz\*, J Santos-Rocha, GP Duarte and FE Xavier

*Departamento de Fisiologia e Farmacologia, Universidade Federal de Pernambuco, Recife, Brazil*

### Correspondence

Fabiano E. Xavier, Departamento de Fisiologia e Farmacologia, Centro de Ciências Biológicas, Universidade Federal de Pernambuco, Avenida Professor Moraes Rêgo, Cidade Universitária, 50670-901, Recife Brazil. E-mail: fabianoxavier@ufpe.br, fabiano.exavier@gmail.com

\*Both authors have contributed equally to this study.

### Keywords

diabetes; hypertension; fetal programming; endothelial dysfunction; cyclooxygenase; prostanoids; insulin resistance

### Received

8 November 2011

### Revised

9 February 2012

### Accepted

5 March 2012

### BACKGROUND AND PURPOSE

The present study was designed to determine how diabetes in pregnancy affects vascular function in their offspring, the influence of age and whether COX activation is involved in this effect.

### EXPERIMENTAL APPROACH

Relaxation responses to ACh were analysed in mesenteric resistance arteries from the offspring of control rats (O-CR) and those of diabetic rats (O-DR) at 3, 6 and 12 months of age. TxB<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2α</sub> release were determined by enzyme immunoassay. COX-1 and COX-2 expression were measured by Western blot analysis.

### KEY RESULTS

O-DR developed hypertension from 6 months of age compared with O-CR. In O-DR, relaxation responses to ACh were impaired in all ages studied and were restored by COX-2 inhibition. TP receptor blockade (SQ29548) restored ACh relaxation in arteries from 3-month-old O-DR while TP and EP receptor blockade (SQ29548 + AH6809) was required to restore it in 6-month-old O-DR. In 12-month-old O-DR, ACh relaxation was restored when TP, EP and FP receptors were blocked (SQ29548 + AH6809 + AL8810). ACh-stimulated TxB<sub>2</sub> was higher in all O-DR. ACh-stimulated PGE<sub>2</sub> release was increased in arteries from 6- and 12-month-old O-DR, whereas PGF<sub>2α</sub> was increased only in 12-month-old O-DR. COX-2, but not COX-1, expression was higher in O-DR than O-CR.

### CONCLUSIONS AND IMPLICATIONS

The results indicate an age-dependent up-regulation of COX-2 coupled to an enhanced formation of vasoconstrictor prostanoids in resistance arteries from O-DR. This effect plays a key role in the pathogenesis of endothelial dysfunction, which in turn could contribute to the progression of vascular dysfunction in these rats.

### Abbreviations

AH6809, 6-isopropoxy-9-oxoanthene-2-carboxylic acid; AL8810, 9α,15R-dihydroxy-11β-fluoro-15-(2,3-dihydro-1H-inden-2-yl)-16,17,18,19,20-pentanor-prosta-5Z,13E-dien-1-oic acid; EP, prostaglandin E<sub>2</sub> receptors; FP, prostaglandin F<sub>2α</sub> receptors; KHS, Krebs-Henseleit solution; NS-398, N-(2-cyclohexyloxy-4-nitrophenyl) methansulfonamide; O-CR, offspring of control rats; O-DR, offspring of diabetic rats; PGE<sub>2</sub>, prostaglandin E<sub>2</sub>; PGF<sub>2α</sub>, prostaglandin F<sub>2α</sub>; SC-560, 5-(4-chlorophenyl)-1-(4-methoxyphenyl)-3-trifluoromethylpyrazole; SQ29548, [1S-[1a,2a(Z),3a,4a]]-7-[3-[[2-(phenylamino) carbonyl]hydrazino]methyl]-7-oxabicyclo [2.2.1] hept-2-yl]-5-heptanoic acid; TP, thromboxane A<sub>2</sub> receptors; TxA<sub>2</sub>, thromboxane A<sub>2</sub>; U46619, (15)-hydroxy-11,9 -(epoxymethano)prosta-5,13-dienoic acid

## Introduction

In the past few decades, the incidence of cardiovascular diseases has been increasing worldwide. In the early 20th century, it was responsible for less than 10% of deaths around the entire world, while at the beginning of the 21st century it accounted for almost 50% and 25% of deaths in developed and developing countries, respectively [World Health Organization (WHO), 2002]. In most countries, the increased incidence of cardiovascular diseases has been attributed in part to environmental factors such as diet, smoking and reduced physical exercise (WHO, 2002). On the other hand, the susceptibility to cardiovascular diseases may also be acquired *in utero* through changes in the uterine environment. Fetal programming refers to the observations that disturbances during the critical period of development can cause lifelong changes in the structure and function of the organism leading to diseases in later life (Barker, 2004). This concept comes from epidemiological studies by Barker and colleagues who obtained evidence for an inverse relationship between low weight at birth and development of cardiovascular diseases in adulthood (Barker, 1995; 2004; Barker *et al.*, 2002). However, other maternal conditions that produce an adverse environment to fetal development, including chronic hyperglycaemia, also increase the risk of metabolic and cardiovascular diseases in the offspring (Simeoni and Barker, 2009).

Fetal exposure to maternal hyperglycaemia can result in insulin resistance, changes in glucose metabolism and increased risk of hypertension in adult life (Nehiri *et al.*, 2008; Segar *et al.*, 2009; Simeoni and Barker, 2009; Chen *et al.*, 2010; Blondeau *et al.*, 2011). This hypertension has been partially attributed to renal mechanisms. Amri *et al.* (1999) demonstrated that exposure to maternal diabetes impairs the offspring's nephrogenesis, reducing the number of nephron, which could be a risk factor for the development of chronic renal disease and hypertension in adulthood. In addition, baroreflex dysfunction and increased activity of the renin-angiotensin system have also been reported in adult offspring of diabetic mothers (Wichi *et al.*, 2005).

Other studies have shown that maternal diabetes promotes alterations in vascular reactivity in their offspring. Rocha *et al.* (2005) demonstrated that offspring of streptozotocin-induced diabetic rats had reduced endothelium-dependent relaxation and hypertension in adulthood. Likewise, Holemans *et al.* (1999) observed impaired endothelial function in resistance arteries from offspring of diabetic rats, but without changes in BP. The reason for this discrepancy with regard to BP is not fully known, but the age of animals used may have influenced the results. Rocha *et al.* (2005) used 12-month-old rats while Holemans *et al.* (1999) used animals that were only 3 months old, which suggest that age may influence the development of hypertension in these animals. The results of Nehiri *et al.* (2008) confirm this hypothesis; these authors demonstrated an increased BP in the offspring of diabetic rats when they were only 6 months-old. The contribution of the vascular system to these changes in BP as well as the mechanisms involved remain to be elucidated.

Several studies have demonstrated that the balance between vasodilator and vasoconstrictor prostanoids is modified in hypertension (Alvarez *et al.*, 2005; Virdis *et al.*, 2009; Féretou *et al.*, 2011) and diabetes (Bagi *et al.*, 2005; Féretou

*et al.*, 2011), and may involve the inducible form of the enzyme, COX-2. In healthy arteries, most prostanoids are produced by the constitutive isoform of COX (COX-1). However, these products may also be synthesized by the COX-2 that is usually expressed at undetectable levels in the vascular wall but can be up-regulated by inflammatory and physical stimuli (Féretou *et al.*, 2011). COX-2-derived prostanoids are associated with the development of vascular complications under conditions of insulin resistance and cardiovascular risk (Helmersson *et al.*, 2004; Bagi *et al.*, 2005; Elmarakby and Imig, 2010; Retailleau *et al.*, 2010). On the basis of these findings, we hypothesized that endothelial dysfunction in resistance arteries from the offspring of insulin-resistant diabetic rats is the result of enhanced COX-2 expression/metabolism, which increases the formation of contractile prostanoids.

In the present study, we tested our hypothesis and specifically investigated the effects of COX inhibitors on ACh-induced relaxation of mesenteric resistance arteries from the offspring of control and diabetic rats. Secondly, we identified the prostanoids released by ACh in arteries from the offspring of diabetic rats and compared their concentrations with those produced in arteries from the offspring of control rats. The influence of age on these parameters was also investigated.

## Methods

### Experimental animals

Male and female Wistar rats (250–300 g) were obtained from colonies maintained in the animal quarters of the Universidade Federal de Pernambuco (UFPE). Rats were housed at a constant room temperature, humidity and light cycle (12 h light/dark) and had free access to tap water and standard rat chow *ad libitum*. This study was approved by the Institutional Animal Care and Use Committee of UFPE (approval reference number: 23076.015755/2008-96), and conforms to the *Guide for Care and Use of Laboratory Animals* published by the National Institutes of Health (NIH publication 85-23, revised 1996). The results of all studies involving animals are reported in accordance with the ARRIVE guidelines (Kilkenny *et al.*, 2010; McGrath *et al.*, 2010).

On the 7th day of pregnancy, diabetes was induced by a single injection of streptozotocin (50 mg·kg<sup>-1</sup>; i.p.). The diabetes was confirmed by measuring plasma glucose concentrations (ACCU-CHEK®, Roche Diagnostics, Mannheim, Germany). After birth, each litter was reduced to six pups and restricted to male offspring only. The unwanted pups were killed by CO<sub>2</sub> inhalation followed by cervical dislocation. When the number of males was not enough to complete six, females were used but discarded at weaning. The offspring were divided into two groups: (i) O-CR – offspring of control rats and (ii) O-DR – offspring of diabetic rats. In this study, we used O-CR and O-DR at 3, 6 and 12 months of age.

### Glucose tolerance and insulin sensitivity

The oral glucose tolerance test was performed according to a standard protocol. After a 10 h fast, a single oral dose (2 g·kg<sup>-1</sup> of body weight) of glucose was delivered. Blood glucose was

then measured from the tail vein just before, and 30, 60, 90 and 120 min after glucose injection, using test strips and reader (ACCU-CHEK®, Roche Diagnostics). After 48 h, the animals were subjected to a new 10 h fast for assessment of insulin sensitivity by insulin tolerance test. For this, regular insulin was administered i.p. at the dose of  $1.5 \text{ U} \cdot \text{kg}^{-1}$  body weight. Blood glucose was determined before and 15, 30, 45 and 60 min after insulin administration.

### Arterial BP measurement

Rats were anaesthetized with a mixture of ketamine, xylazine and acetopromazin (64.9, 3.2 and  $0.78 \text{ mg} \cdot \text{kg}^{-1}$ , respectively, i.p.). The right carotid artery was cannulated with a polyethylene catheter (PE-50) that was exteriorized in the mid scapular region. Adequacy of anaesthesia was assessed by monitoring withdrawal reflexes. After 24 h, arterial pressure was measured in conscious, freely moving rats. The arterial cannula was connected to a transducer and pressure signals were recorded for a 60 min period using an interface and software for computer data acquisition (ADIInstruments Pty Ltd, Castle Hill, New South Wales, Australia).

### Vessel preparation

Rats were anaesthetized with ketamine, xylazine and acetopromazin mixture (64.9, 3.2 and  $0.78 \text{ mg} \cdot \text{kg}^{-1}$ , respectively, i.p.) and killed by exsanguination. The mesenteric vascular bed was removed and placed in cold ( $4^\circ\text{C}$ ) Krebs-Henseleit solution (KHS; in mM: 115 NaCl, 2.5 CaCl<sub>2</sub>, 4.6 KCl, 1.2 KH<sub>2</sub>PO<sub>4</sub>, 1.2, MgSO<sub>4</sub>.7H<sub>2</sub>O, 25 NaHCO<sub>3</sub>, 11.1 glucose and 0.03 EDTA). For reactivity experiments, the third-order branch of the mesenteric arcade was dissected and cut into segments of approximately 2 mm in length. Segments of mesenteric resistance arteries were mounted in a small vessel chamber myograph (Danish Myo Technology A/S, Aarhus, Denmark) to measure isometric tension according to the method described by Mulvany and Halpern (1977).

### Experimental protocols

After a 45 min equilibration period, each arterial segment was exposed to KCl (120 mM) to assess its maximum contractility. After a washout period, the presence of the vascular endothelium was confirmed by the ability of 1  $\mu\text{M}$  ACh to relax segments precontracted with noradrenaline at a concentration that produced approximately 50–70% of the contraction induced by KCl. The segments were rinsed with KHS for 1 h and then a cumulative ACh concentration-response curve (0.1 nM to 3  $\mu\text{M}$ ) was obtained in the noradrenaline-precontracted segments. Endothelium-independent relaxation was studied by evaluating relaxation to sodium nitroprusside (1 nM–10  $\mu\text{M}$ ) in arteries previously contracted with noradrenaline. The possible role of COX-derived metabolites was investigated in segments from O-CR and O-DR rats. Arteries were pre-incubated with either indomethacin (a COX-1 and COX-2 inhibitor, 10  $\mu\text{M}$ ), SC-560 (a COX-1 inhibitor, 1  $\mu\text{M}$ ), NS-398 (COX-2 inhibitor, 10  $\mu\text{M}$ ), SQ29548 [Tx<sub>A</sub><sub>2</sub> receptor (TP) antagonist, 1  $\mu\text{M}$ ], AH6809 [PGE<sub>2</sub> receptor (EP<sub>1</sub>, EP<sub>2</sub> and EP<sub>3</sub>) antagonist, 30  $\mu\text{M}$ ] or AL8810 [PGF<sub>2α</sub> receptor (FP) antagonist, 10  $\mu\text{M}$ ], before generating concentration-response curves to ACh. All drugs were added 30 min before the concentration-response curve to ACh.

In another set of experiments, the vasoactive responses to the TP receptor agonist U46619 (1 nM–10  $\mu\text{M}$ ), PGE<sub>2</sub> (10 nM–10  $\mu\text{M}$ ) or PGF<sub>2α</sub> (1 nM to 3  $\mu\text{M}$ ), were analysed in quiescent arteries from all groups.

### Prostanoid production

To measure the release of Tx<sub>A</sub><sub>2</sub>, PGE<sub>2</sub>, PGF<sub>2α</sub> and PGI<sub>2</sub>, we used specific enzyme immunoassay kits (Cayman Chemical Company, Ann Arbor, MI, USA) for each prostanoid. The second, third and fourth branches of mesenteric artery were pre-incubated for 45 min in 200  $\mu\text{L}$  of gassed KHS at  $37^\circ\text{C}$ . Afterwards, three washout periods of 10 min in a bath of 200  $\mu\text{L}$  of KHS were run before incubation with ACh (0.1 nM–3  $\mu\text{M}$ ). The different assays were performed following the manufacturer's instructions. Results are expressed as  $\text{pg} \cdot \text{mL}^{-1} \text{ mg}^{-1}$  wet tissue.

### Western blot analysis of COX-1 and COX-2

Mesenteric resistance arteries from O-CR and O-DR rats were homogenized; proteins were electrophoretically separated (30  $\mu\text{g}$  per lane) and transferred to polyvinyl difluoride membranes (GE Healthcare do Brasil Ltda, São Paulo, SP, Brazil). Membranes were incubated with antibodies (Cayman Chemical Company) against COX-1 (1:1000 dilution), COX-2 (1:500 dilution) or α-actin (1:3000 dilution) proteins and individual horseradish peroxidase-conjugated secondary antibodies (GE Healthcare do Brasil Ltda) in blocking buffer. Immunoreactive proteins were detected by chemiluminescence with ECL Plus (GE Healthcare do Brasil Ltda) and subjected to autoradiography (Hyperfilm ECL, GE Healthcare do Brasil Ltda). Signals on the immunoblot were quantified using a computer programme (NIH Image V1.56, Bethesda, MD, USA). Ram seminal vesicle microsomes and mouse macrophage microsomes protein (Cayman Chemical Company) were used as positive control for COX-1 and COX-2, respectively.

### Drugs

Drugs used were noradrenaline hydrochloride, ACh chloride, sodium nitroprusside, indomethacin (Sigma, St. Louis, MO, USA), NS-398, SQ29548, AH6809, AL8810, PGE<sub>2</sub>, PGF<sub>2α</sub> (Cayman Chemical Company) and U46619 (Calbiochem-Novabiochem GmbH, Darmstadt, Germany). Stock solutions of ACh and sodium nitroprusside were made in distilled water; noradrenaline was dissolved in an NaCl (0.9%)–ascorbic acid (0.01% w v<sup>-1</sup>) solution; indomethacin, SQ29548, PGE<sub>2</sub> and PGF<sub>2α</sub> were dissolved in ethanol; and AH6809, AL8810 and U46619 were dissolved in dimethyl sulfoxide. These solutions were kept at  $-20^\circ\text{C}$  and appropriate dilutions were made on the day of the experiment. Drug/molecular target nomenclature conforms to the *British Journal of Pharmacology's Guide to Receptors and Channels* (Alexander *et al.*, 2011).

### Statistical analysis

Relaxation responses to ACh and sodium nitroprusside are expressed as a percentage of the maximum contractile response induced by noradrenaline. U46619, PGE<sub>2</sub> and PGF<sub>2α</sub> contractile responses are expressed as a percentage of the maximum response produced by KCl.

All values are expressed as means  $\pm$  SEM of the number of animals used in each experiment. Results were analysed using Student's *t*-test or by one-way or two-way ANOVA for comparison between groups. When ANOVA showed a significant treatment effect, Bonferroni's *post hoc* test was used to compare individual means. Differences were considered statistically significant at  $P < 0.05$ .

## Results

Dams injected with streptozotocin had severe hyperglycaemia on gestational days 14 and 21 compared with control dams (control  $852 \pm 24$  vs. diabetic  $4820 \pm 225$  mg·L $^{-1}$ , *t*-test:  $P < 0.05$ ). Gestation occurred normally, and the rats delivered spontaneously at term (21 days of gestation). Diabetic dams gave birth to fewer pups than the controls (control:  $10 \pm 1$  vs. diabetic  $6 \pm 2$  pups per litter; *t*-test:  $P < 0.05$ ). As shown in Table 1, mean body weight was significantly lesser in O-DR than O-CR. Blood glucose levels were similar in O-CR and O-DR (Table 1).

Oral glucose tolerance test was performed at 3 and 12 months of age. Blood glucose levels were higher in both 3 and 12-month-old O-DR at 30 min compared with O-CR rats (results not shown) and remained increased until the time of 120 min (3-month-old rats: O-CR,  $1050 \pm 49$  vs. O-DR,  $1280 \pm 45$  mg·L $^{-1}$ , *t*-test:  $P < 0.05$ ; 12-month-old rats: O-CR,  $1060 \pm 32$  vs. O-DR,  $1420 \pm 23$  mg·L $^{-1}$ , *t*-test:  $P < 0.05$ ). Results from the insulin tolerance test demonstrated significant insulin resistance among the O-DR rats, as they presented a higher blood glucose from 15 min to 60 min after an insulin injection (blood glucose 60 min after the insulin injection; 3-month-old rats: O-CR,  $350 \pm 40$  vs. O-DR,  $470 \pm 23$  mg·L $^{-1}$ , *t*-test:  $P < 0.05$ ; 12-month-old rats: O-CR,  $330 \pm 15$  vs. O-DR,  $572 \pm 32$  mg·L $^{-1}$ , *t*-test:  $P < 0.05$ ).

O-DR presented higher BP in adulthood. Although the mean arterial pressure of 3-month-old rats was similar in both groups (O-CR:  $97.5 \pm 2.54$  vs. O-DR:  $104 \pm 8.40$  mmHg, *t*-test,  $P > 0.05$ ), it was significantly increased in O-DR at both 6 (O-CR:  $105 \pm 4.70$  vs. O-DR:  $132 \pm 5.30$  mmHg, *t*-test,  $P > 0.05$ ) and 12 months (O-CR:  $102 \pm 5.10$  vs. O-DR:  $149 \pm 3.70$  mmHg, *t*-test,  $P > 0.05$ ) compared with O-CR. The heart rate was similar in all O-DR groups compared with their respective age-matched O-CR (results not shown).

### Vascular function in adult diabetic offspring

KCl (120 mM) evoked similar contractions in vessels from both diabetic and age-matched control offspring rats

(3-month-old rats, O-CR:  $3.15 \pm 0.04$  vs. O-DR:  $3.19 \pm 0.07$  mN·mm $^{-1}$ ; 6-month-old rats, O-CR:  $3.28 \pm 0.12$  vs. O-DR:  $3.19 \pm 0.09$  mN·mm $^{-1}$ ; 12-month-old rats, O-CR:  $3.21 \pm 0.02$  vs. O-DR:  $3.25 \pm 0.13$  mN·mm $^{-1}$ ; ANOVA,  $P > 0.05$ ).

ACh induced cumulative concentration- and endothelium-dependent relaxations of noradrenaline-contracted arteries from 3-, 6- and 12-month-old O-CR (Figure 1). However, in mesenteric arteries from O-DR, ACh induced a biphasic response, characterized by a relaxing effect at concentrations equal or below 0.1  $\mu$ M, which was lower than that in age-matched O-CR, and by a contractile response at concentrations above 0.3  $\mu$ M that was absent in arteries from O-CR (Figure 1A). Further deterioration of this relaxation was noted with aging (compare Figure 1A–C). Mesenteric resistance arteries from 3-month-old O-DR relaxed 86% to ACh while 6- and 12-month-old O-DR showed 59% and 37% relaxation, respectively (ANOVA:  $P < 0.05$ ). In arteries from O-CR, relaxation to ACh did not change with age (Figure 1). In all groups, relaxation induced by sodium nitroprusside was comparable (Figure 2).

### Effects of COX-1 and COX-2 inhibition and TP, EP and FP receptor antagonism on endothelium-dependent relaxation

In 3-, 6- and 12-month-old O-CR, ACh-induced vasodilatation was not modified by indomethacin or NS-398 (Figure 3A,C,E). In contrast, the altered response to ACh in arteries from 3-, 6- and 12-month-old O-DR was normalized by either indomethacin or NS-398, but not by SC-560, (Figure 3B,D,F).

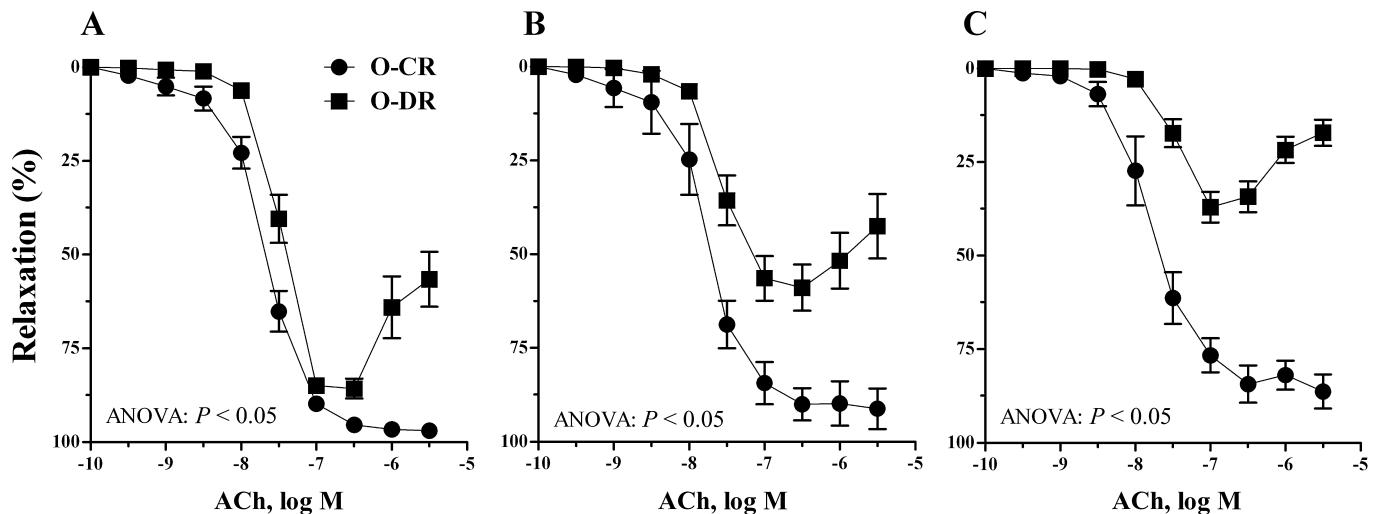
In vessels from 3-month-old O-DR, the response to ACh was also enhanced by SQ29548 to similar values to those obtained with COX-2 blockade (Figure 4A). When applied concomitantly with SQ29548, AH6809 or AH6809 plus AL8810 failed to further increase ACh-induced relaxation (Figure 4A). In 6-month-old O-DR, SQ29548 increased the relaxation to ACh, but its effect was reduced compared to that obtained with NS-398 (Figure 4B). When co-incubated with SQ29548, AH6809 increased the response to ACh to similar values to those obtained with NS-398. AL8810 failed to enhance the effect of SQ29548 plus AH6809 (Figure 4B). In arteries from 12-month-old O-DR, SQ29548 partly increased the response to ACh (Figure 4C). In arteries from this group, although co-incubation with AH6809 increased the effect of SQ29548, the normalization of the ACh response was achieved only when arteries were treated with SQ29548 in combination with AH6809 and AL8810 (Figure 4C).

**Table 1**

Body weights (BW) and blood glucose (BG) at the times of vascular testing

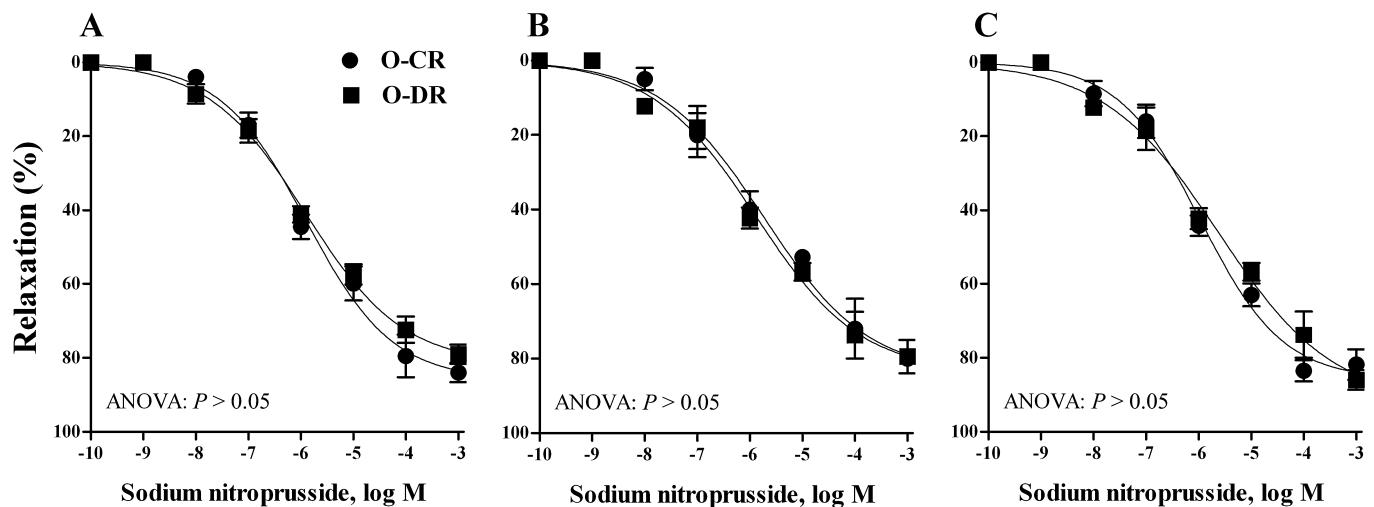
	3 months		6 months		12 months	
	BW (g)	BG (mg·L $^{-1}$ )	BW (g)	BG (mg·L $^{-1}$ )	BW (g)	BG (mg·L $^{-1}$ )
O-CR	$318 \pm 7.23$	$957 \pm 34$	$422 \pm 6.18$	$952 \pm 27$	$467 \pm 0.40$	$942 \pm 41$
O-DR	$295 \pm 9.40^*$	$994 \pm 13$	$375 \pm 11.7^*$	$925 \pm 30$	$443 \pm 2.50^*$	$908 \pm 21$

Values are means  $\pm$  SEM. ANOVA: \* $P < 0.05$  compared with O-CR at the same time point.  
O-CR, offspring of control rats; O-DR, offspring of diabetic rats.



**Figure 1**

Endothelium-dependent relaxation induced by ACh in mesenteric resistance arteries from (A) 3-, (B) 6- and (C) 12-month-old offspring of control (O-CR) and diabetic (O-DR) rats. Each point represents the mean of 7–8 experiments  $\pm$  SEM.



**Figure 2**

Endothelium-independent relaxation induced by sodium nitroprusside in mesenteric resistance arteries from (A) 3-, (B) 6- and (C) 12-month-old offspring of control (O-CR) and diabetic (O-DR) rats. Each point represents the mean of 7–8 experiments  $\pm$  SEM.

### Vascular response to U46619, PGE<sub>2</sub> and PGF<sub>2α</sub>

U46619, PGE<sub>2</sub> and PGF<sub>2α</sub> caused cumulative concentration-dependent contractions in quiescent arteries. These contractions were not significantly different (ANOVA:  $P > 0.05$ ) between preparations of O-CR and O-DR (results not shown).

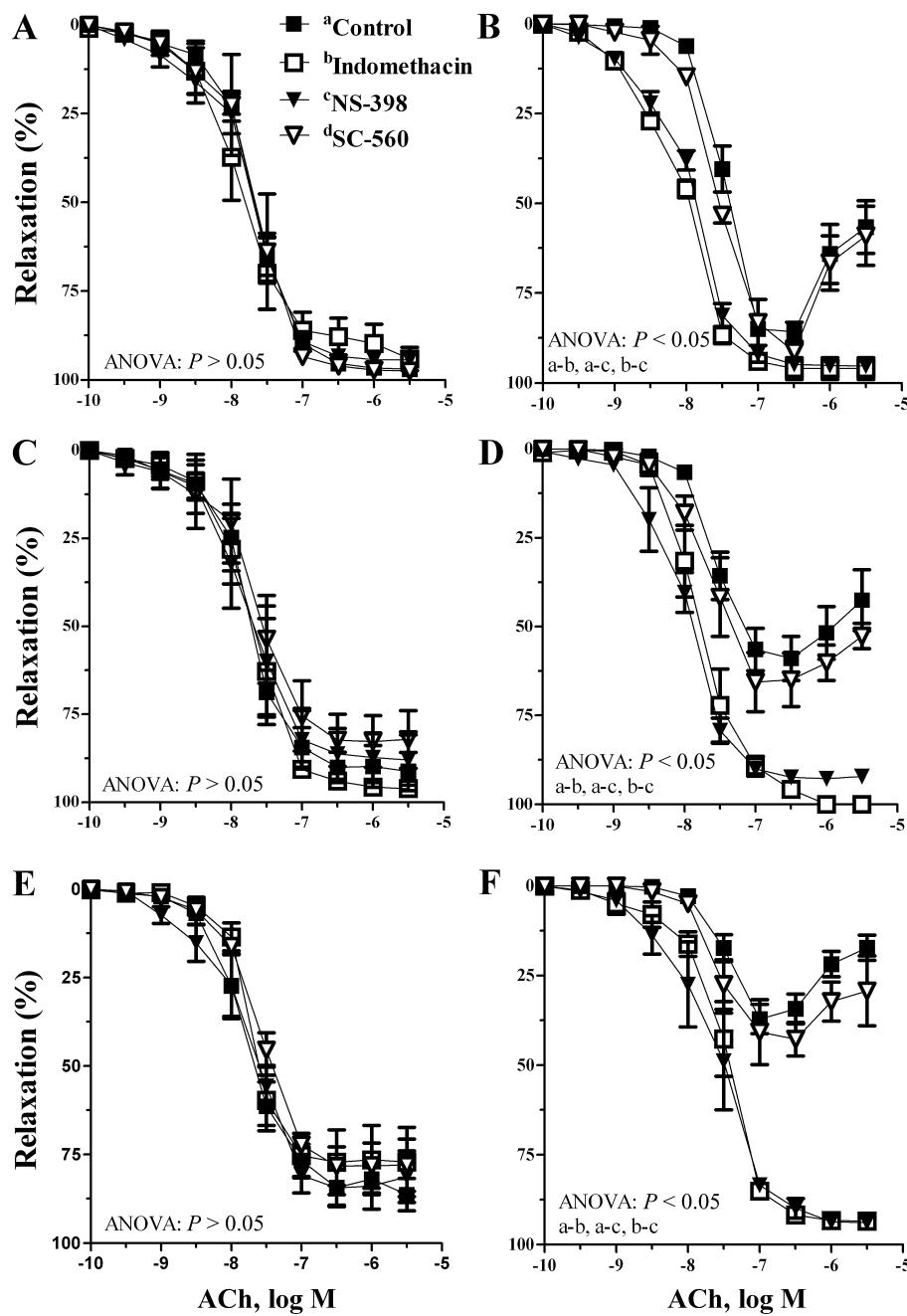
### Prostanoids production

In mesenteric resistance arteries from all groups, ACh increased the production of Tx<sub>B2</sub>, PGE<sub>2</sub> and PGF<sub>2α</sub> (Figure 5). The ACh-stimulated Tx<sub>B2</sub> levels were higher in arteries from both 3-, 6- and 12-month-old O-DR compared with their respective age-matched O-CR (Figure 5A). In arteries from 6-

and 12-month-old O-DR, the ACh-stimulated levels of PGE<sub>2</sub> were higher, while in 3-month-old O-DR they were not modified (Figure 5B). The ACh-stimulated PGF<sub>2α</sub> levels were higher only in arteries from 12-month-old O-DR (Figure 5C).

### COX-1 and COX-2 protein expression

Western blot analysis showed a basal protein expression of COX-2 in mesenteric arteries from O-CR and O-DR (Figure 6). This expression was higher in arteries from O-DR and increased with age, reaching higher levels in 12-month-old O-DR (Figure 6). COX-1 expression was similar in all groups (Figure 6).

**Figure 3**

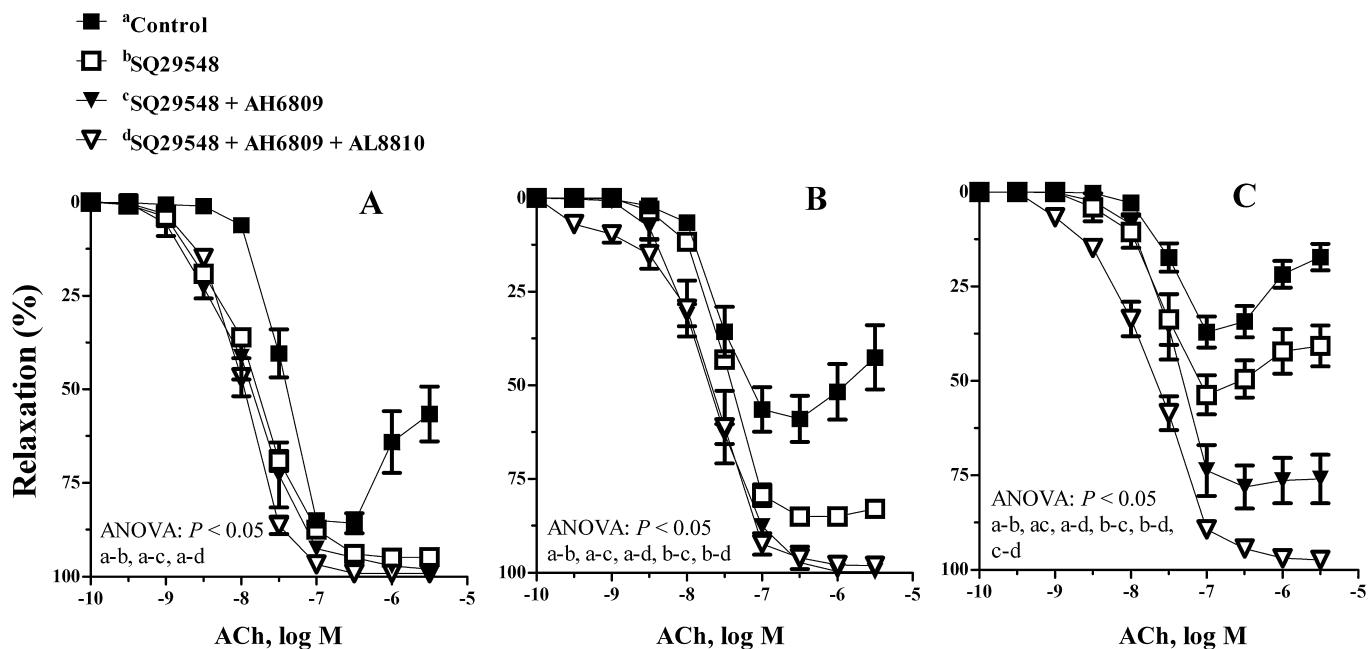
Effect of indomethacin (10  $\mu$ M) or NS-398 (10  $\mu$ M) on the concentration-dependent relaxation to ACh in segments of mesenteric resistance arteries from (A) 3-month-old O-CR, (B) 3-month-old ODR, (C) 6-month-old O-CR, (D) 6-month-old ODR, (E) 12-month-old O-CR and (F) 12-month-old ODR. Results are expressed as mean  $\pm$  SEM;  $n = 7$ –8 animals in each group.

## Discussion and conclusions

In the present study, we investigated the long-term vascular consequences of *in utero* exposure to maternal hyperglycaemia in rats. Our major novel finding is that this exposure is associated with an age-dependent COX-2 up-regulation, coupled with an enhanced formation of contracting prostanooids that contribute to the impairment of endothelial

function in mesenteric resistance arteries from adult offspring of diabetic rats.

Hyperglycaemia in pregnancy leads to insulin resistance in the offspring (Segar *et al.*, 2009; Blondeau *et al.*, 2011), leading to vasoconstriction, inflammation and thrombosis, which occasionally produces hypertension (Chahwala and Arora, 2009). In the current study, although no offspring group developed pre-diabetes or diabetes during the



**Figure 4**

Effect of SQ29548 (1  $\mu$ M) alone or in association with AH6809 (30  $\mu$ M) or AH6809 plus AL8810 (10  $\mu$ M) on the concentration-dependent relaxation to ACh in segments of mesenteric resistance arteries from (A) 3-, (B) 6- and (C) 12-month-old offspring of diabetic (O-DR) rats. Results are expressed as mean  $\pm$  SEM;  $n = 7$ -8 animals in each group.

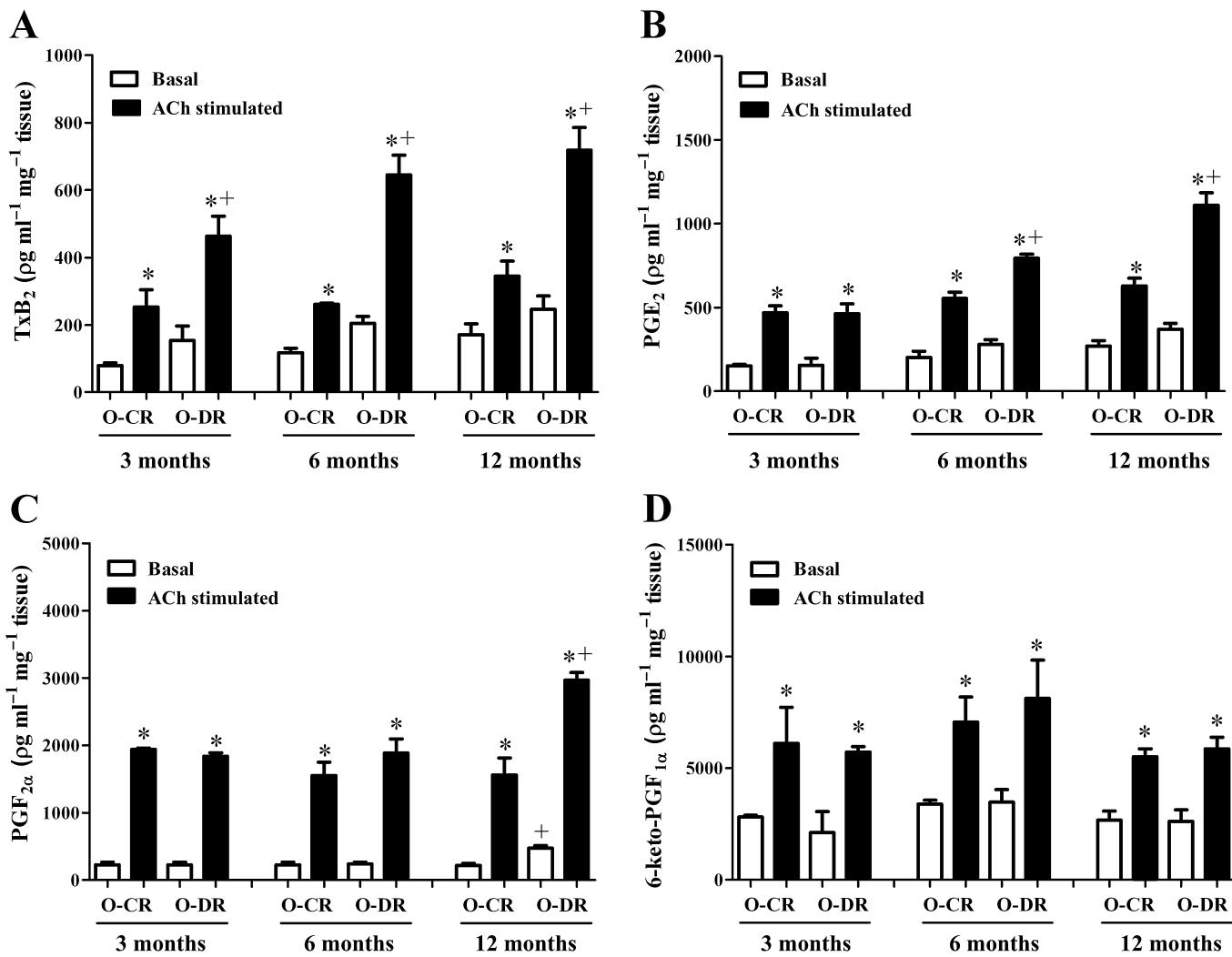
follow-up period, O-DR showed insulin resistance and glucose intolerance at 3 months of age and a further impairment of these parameters at 12 months of age. This effect did not seem to be related to a toxic effect of streptozotocin on the fetal endocrine pancreas as: (i) streptozotocin has a very short (30 min) half-life (Schein and Loftus, 1968); (ii) beta-cell development in the rat occurs mainly during the last week of fetal growth (Blondeau and Breant, 2005); (iii) streptozotocin presented no cytotoxic effect on fetal pro-islets (Liu *et al.*, 1994); and (iv) insulin resistance in O-DR occurs irrespective of the time of streptozotocin administration.

The mechanisms of the hyperglycaemia-programmed hypertension are complex and involve renal, neural and vascular effects (Amri *et al.*, 1999; Holemans *et al.*, 1999; Rocha *et al.*, 2005; Wichi *et al.*, 2005; Nehiri *et al.*, 2008; Segar *et al.*, 2009; Chen *et al.*, 2010). Previous studies have demonstrated a reduced endothelium-dependent relaxation response to dilators in both conductance (Segar *et al.*, 2009; Porto *et al.*, 2010) and resistance (Holemans *et al.*, 1999; Rocha *et al.*, 2005) arteries from O-DR compared with O-CR. This response has been accompanied, or not, by hypertension. Holemans *et al.* (1999) demonstrated impaired relaxation to ACh and bradykinin in arteries from 3-month-old offspring of diabetic dams, but without changes in BP. On the other hand, Rocha *et al.* (2005) demonstrated increased BP in 12-month-old diabetic offspring associated with impaired endothelial and renal functions. Therefore, it is possible to postulate that the phenotype of the offspring of diabetic rats tends to vary according to age at the time of analysis.

The results obtained here show that although the BP in 3-month-old rats was similar in both groups, it was increased in the 6- and 12-month-old rats in O-DR compared with

those in the O-CR group, as previously reported by Nehiri *et al.* (2008). Our results also demonstrate that in mesenteric resistance arteries from 3-, 6- and 12-month-old O-DR, the relaxation to ACh, but not to sodium nitroprusside, was impaired compared with that observed in O-CR. In O-DR arteries, this relaxation was progressively impaired with age while in O-CR it remained unmodified. The importance of abnormal endothelium-dependent dilation, as observed in O-DR, lies in the possible pathological consequences. Reduced endothelial function may contribute to coagulation, inflammation and atherosclerosis, as well as to increased peripheral resistance and hypertension (Félétou and Vanhoutte, 2006). Our results also revealed that the endothelial dysfunction in O-DR precedes hypertension, suggesting that this mechanism could be involved in the genesis of hypertension in these animals. Furthermore, a greater impairment of the endothelium-dependent relaxation in arteries from 6- and 12-month-old O-DR was associated with a higher BP in these rats. The mechanism by which maternal diabetes impairs the endothelial function is not fully known. Insulin resistance, perhaps as a result of raised plasma cholesterol and triglycerides, is implicated in endothelial dysfunction (Johnstone *et al.*, 1993). Transient hyperglycaemia (Holemans *et al.*, 1997), oxidative stress (Siman and Eriksson, 1997) or the synthesis of advanced glycosylation end-products (Vlassara *et al.*, 1992) *in utero* have also been suggested as possible mechanisms involved in decreased endothelium-dependent vasodilatation in the adult offspring.

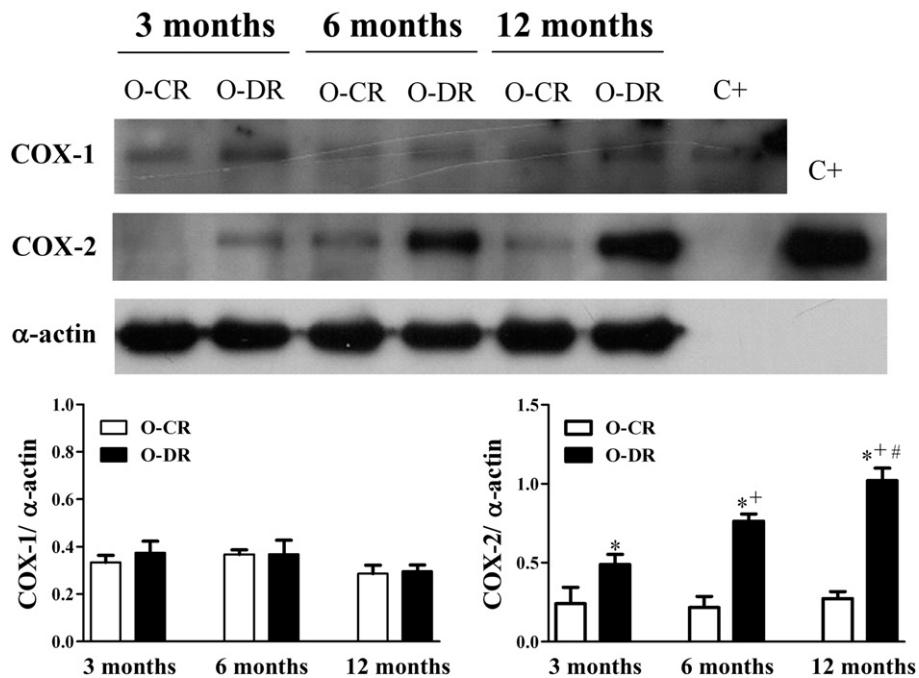
Insulin resistance is associated with a pro-inflammatory state of the vascular wall leading to vascular remodelling and endothelial dysfunction in systemic arteries (Chahwala and Arora, 2009). Various studies have shown that COX-2 is

**Figure 5**

Basal and ACh-stimulated production of TxB<sub>2</sub> (A), PGE<sub>2</sub> (B), PGF<sub>2α</sub> (C) and prostacyclin (6-keto-PGF<sub>1α</sub>, D) in mesenteric resistance segments from 3-, 6- and 12-month-old offspring of control (O-CR) or diabetic (O-DR) rats.  $n = 6-7$  animals in each group. ANOVA: \* $P < 0.05$  ACh-stimulated vs. basal; + $P < 0.05$  O-DR vs. age-matched O-CR.

induced during some inflammatory process and that prostanoids produced by this COX isoform are responsible for many inflammatory symptoms (Parente and Perretti, 2003). In this sense, COX-2-derived prostanoids have been shown to be associated with the development of vascular complications under conditions of insulin resistance and cardiovascular risk (Helmersson *et al.*, 2004; Bagi *et al.*, 2005; Virdis *et al.*, 2009; Elmarakby and Imig, 2010; Retailleau *et al.*, 2010; Féletalou *et al.*, 2011), which in turn could contribute to elevation in BP (Tian *et al.*, 2011). The O-DRs used in the present study were insulin resistant and presented endothelial dysfunction in adulthood. To investigate the participation of COX-derived products in the ACh responses, we used the non-selective COX inhibitor indomethacin, the selective COX-1 inhibitor SC-560 and the selective COX-2 inhibitor NS-398. In arteries from 3-, 6- and 12-month-old O-DR, indomethacin or NS-398, but not SC-560, restored the ACh-induced relaxation to similar levels to that in O-CR, indicating that contractile prostanoids from COX-2, but not from COX-1, contribute to the endothelial dysfunction in these rats. In keeping with these functional results, Western blot analysis revealed an up-regulation of COX-2, but not COX-1, in arteries from all O-DR, reaching higher levels at 12 months of age. In O-CR arteries, inhibition of COX-1 or COX-2 did not affect the response to ACh, suggesting that in these rats COX isoenzymes do not have a significant effect on the ACh-induced relaxation, as previously reported (Xavier *et al.*, 2008).

Activation of TP receptors allows vasoconstrictor prostanoids such as TxA<sub>2</sub> to contribute to endothelial dysfunction in different cardiovascular disorders (Alvarez *et al.*, 2005; Blanco-Rivero *et al.*, 2005; Xavier *et al.*, 2008). In the present study, blockade of the TP receptors (SQ29548) increased ACh-induced relaxation in mesenteric arteries from all O-DR. In the 3-month-old O-DR, this effect was similar to that produced by NS-398, suggesting that TxA<sub>2</sub> is the COX-2-derived



**Figure 6**

Representative Western blot for COX-1, COX-2,  $\alpha$ -actin protein expression and the positive control (C+) for COX-1 and COX-2 in mesenteric resistance arteries from 3-, 6- and 12-month-old offspring of control (O-CR) or diabetic (O-DR) rats (upper panel). Graph shows densitometric analysis of the Western blot for COX-1 and COX-2. Results (means  $\pm$  SEM) are expressed as the ratio between the signal for the COX-1 or COX-2 protein and the signal for  $\alpha$ -actin.  $n = 6$  animals in each group. ANOVA: \* $P < 0.05$  O-DR versus age-matched O-CR; † $P < 0.05$  6-month-old O-DR versus 3-month-old O-DR; # $P < 0.05$  12-month-old O-DR versus 6-month-old O-DR.

prostanoid involved in the endothelial dysfunction in this group. Indeed, in arteries from these rats, ACh-stimulated release of TxB<sub>2</sub> (the TxA<sub>2</sub> metabolite) was increased. On the other hand, it should be noted that ACh-induced relaxation in arteries from 6- and 12-month-old O-DR was not entirely restored by SQ29548. In vessels from 6-month-old O-DR, relaxation to ACh was completely restored by additional application of the EP receptor blocker, AH6809, while the combined treatment of SQ29548 plus AH6809 and AL8810 (an FP receptor antagonist) was required to fully restore this response in arteries from 12-month-old O-DR. This implies that in arteries from 6-month-old O-DR, TxA<sub>2</sub> and PGE<sub>2</sub> contribute to the COX-2-dependent endothelial dysfunction while in 12-month-old O-DR, in addition to these two prostanoids, PGF<sub>2 $\alpha$</sub>  seems to play a role. In keeping with these functional findings, ACh-stimulated release of TxB<sub>2</sub> and PGE<sub>2</sub> was higher in arteries from 6-month-old O-DR. In addition to these two prostanoids, ACh-stimulated PGF<sub>2 $\alpha$</sub>  was also increased in arteries from 12-month-old O-DR. These findings are in accordance with previous results showing that COX-2 induces TxA<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub>  production under inflammatory conditions (Kimura *et al.*, 1994; Alvarez *et al.*, 2005; Blanco-Rivero *et al.*, 2005; Retailleau *et al.*, 2010).

PGI<sub>2</sub> is another prostanoid that could contribute to impaired endothelial function in arteries from O-DR. This prostanoid also produces vasoconstriction that is mediated by activation of TP and EP receptors (Gluais *et al.*, 2005; Xavier *et al.*, 2008; 2009). Thus, the improvement of relaxation to ACh in arteries from O-DR, obtained in the presence

of SQ29548 or AH6809, could be also due to blockade of the vasoconstrictor action of PGI<sub>2</sub>. In a previous study, we reported that COX-2 overexpression is accompanied by increased PGI<sub>2</sub> release and blunted ACh-induced relaxation in rat resistance vessels (Xavier *et al.*, 2008). In the current study, the amount of ACh-stimulated 6-keto-PGF<sub>1 $\alpha$</sub>  in arteries from O-DR was comparable with that observed in arteries from age-matched O-CR. Nevertheless, the involvement of PGI<sub>2</sub> in the impaired ACh-induced relaxation in vessels from O-DR cannot be completely ruled out. Duong Van Huyen *et al.* (2010) reported that exposure to maternal diabetes is associated with decreased vascular expression of IP and impaired relaxation to a PGI<sub>2</sub> analogue, which in turn could contribute to the impaired endothelium-dependent relaxation.

Taken together our findings provide the first demonstration of the participation of an age-dependent increased COX-2-derived TxA<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub>  in the changes in endothelial function in resistance vessels from adult offspring of diabetic rats, supporting the possible relevance of these prostanoids in cardiovascular changes induced by exposure *in utero* to hyperglycaemia. The fact that exogenously administered PGE<sub>2</sub>, PGF<sub>2 $\alpha$</sub>  or the TP receptor agonist U46616 produced similar contractions in arteries from all groups studied eliminates the possibility that EP, FP or TP receptor initiated signalling mechanisms are altered in mesenteric resistance arteries from O-DR.

In summary, the present study demonstrates an age-dependent up-regulation of COX-2 coupled to an enhanced formation of vasoconstrictor prostanoids in resistance arteries

from adult offspring of diabetic rats. This increased formation of vasoconstrictor prostanoids plays a key role in the pathogenesis of endothelial dysfunction, which in turn could contribute to progression of vascular dysfunction in these rats.

## Acknowledgements

This work was supported by grants from Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco (FACEPE). FER-A and DBdeQ were supported by a master's degree fellowship award from FACEPE. JS-R was supported by a scientific initiation fellowship award from PIBIC/UFPE. GPD and FEX are recipients of research fellowships from Conselho Nacional de Desenvolvimento Científico e Tecnológico. We are grateful to José Antonio de Albuquerque for his technical assistance.

## Conflict of interest

None.

## References

- Alexander SPH, Mathie A, Peters JA (2011). Guide to receptors and channels (GRAC), 5th edn. Br J Pharmacol 164: S1–S324.
- Alvarez Y, Briones AM, Balfagón G, Alonso MJ, Salaices M (2005). Hypertension increases the participation of vasoconstrictor prostanoids from cyclooxygenase-2 in phenylephrine responses. J Hypertens 23: 767–777.
- Amri K, Freund N, Vilar J, Merlet-Bénichou C, Lelièvre-Pégrier M (1999). Adverse effects of hyperglycemia on kidney development in rats: *in vivo* and *in vitro* studies. Diabetes 48: 2240–2245.
- Bagi Z, Erdei N, Toth A, Li W, Hintze TH, Koller A *et al.* (2005). Type 2 diabetic mice have increased arteriolar tone and blood pressure: enhanced release of COX-2-derived constrictor prostaglandins. Arterioscler Thromb Vasc Biol 25: 1610–1616.
- Barker DJ (1995). Fetal origins of coronary heart disease. BMJ 311: 171–174.
- Barker DJ (2004). The developmental origins of adult disease. J Am Coll Nutr 23: 588–595.
- Barker DJ, Eriksson JG, Forse'n T, Osmond C (2002). Fetal origins of adult disease: strength of effects and biological basis. Int J Epidemiol 31: 1235–1239.
- Blanco-Rivero J, Cachofeiro V, Lahera V, Aras-Lopez R, Márquez-Rodas I, Salaices M *et al.* (2005). Participation of prostacyclin in endothelial dysfunction induced by aldosterone in normotensive and hypertensive rats. Hypertension 46: 107–112.
- Blondeau B, Breant B (2005). Effect of nutrition on fetal development: a view on the pancreatic beta-cells. In: Djelmis J, Desoye G, Ivanisevic M (eds). Diabetology of Pregnancy. Karger: Basel, pp. 83–93.
- Blondeau B, Joly B, Perret C, Prince S, Bruneval P, Lelièvre-Pégrier M *et al.* (2011). Exposure in utero to maternal diabetes leads to glucose intolerance and high blood pressure with no major effects on lipid metabolism. Diabetes Metab 37: 245–241.
- Chahwala V, Arora R (2009). Cardiovascular manifestations of insulin resistance. Am J Ther 16: 14–28.
- Chen YW, Chenier I, Tran S, Scotcher M, Chang SY, Zhang SL (2010). Maternal diabetes programs hypertension and kidney injury in offspring. Pediatr Nephrol 25: 1319–1329.
- Duong Van Huyen JP, Vessières E, Perret C, Troise A, Prince S, Guihot AL *et al.* (2010). In utero exposure to maternal diabetes impairs vascular expression of prostacyclin receptor in rat offspring. Diabetes 59: 2597–2602.
- Elmarakby AA, Imig JD (2010). Obesity is the major contributor to vascular dysfunction and inflammation in high-fat diet hypertensive rats. Clin Sci (Lond) 118: 291–301.
- Félétou M, Vanhoutte PM (2006). Endothelial dysfunction: a multifaceted disorder (The Wiggers Award Lecture). Am J Physiol Heart Circ Physiol 291: 985–1002.
- Félétou M, Huang Y, Vanhoutte PM (2011). Endothelium-mediated control of vascular tone: COX-1 and COX-2 products. Br J Pharmacol 164: 894–912.
- Gluais P, Lonchamp M, Morrow JD, Vanhoutte PM, Feletou M (2005). Acetylcholine-induced endothelium-dependent contractions in the SHR aorta: the Janus face of prostacyclin. Br J Pharmacol 146: 834–845.
- Helmersson J, Vessby B, Larsson A, Basu S (2004). Association of type 2 diabetes with cyclooxygenase-mediated inflammation and oxidative stress in an elderly population. Circulation 109: 1729–1734.
- Holemans K, Van Bree R, Verhaeghe J, Meurrens K, Van Assche FA (1997). Maternal semistarvation and streptozotocin-diabetes in rats have different effects on the *in vivo* glucose uptake by peripheral tissues in their female adult offspring. J Nutr 127: 1371–1376.
- Holemans K, Gerber RT, Meurrens K, de Clerck F, Poston I, Van Assche FA (1999). Streptozotocin diabetes in the pregnant rat induces cardiovascular dysfunction in adult offspring. Diabetologia 42: 81–89.
- Johnstone MT, Creager SJ, Scales KM, Cusco JA, Lee BK, Creager MA (1993). Impaired endothelium-dependent vasodilation in patients with insulin-dependent diabetes mellitus. Circulation 88: 2510–2516.
- Kilkenny C, Browne W, Cuthill IC, Emerson M, Altman DG (2010) NC3Rs Reporting Guidelines Working Group. Br J Pharmacol 160: 1577–1579.
- Kimura I, Hata Y, Islam MA, Kimura M (1994). Diabetes mellitus-induced enhancement of prostaglandin F<sub>2</sub> alpha-responses is inhibited by lipoxygenase- but not cyclooxygenase-inhibitors in mesenteric veins and arteries of mouse and rat. Jpn J Pharmacol 64: 65–70.
- Liu X, Hering BJ, Brendel MD, Bretzel RG (1994). The effect of streptozotocin on the function of fetal porcine and rat pancreatic (pro-)islets. Exp Clin Endocrinol 102: 374–379.
- McGrath J, Drummond G, McLachlan E, Kilkenny C, Wainwright C (2010). Guidelines for reporting experiments involving animals: the ARRIVE guidelines. Br J Pharmacol 160: 1573–1576.
- Mulvany MJ, Halpern W (1977). Contractile properties of small arterial resistance vessels in spontaneously hypertensive and normotensive rats. Circ Res 41: 19–26.
- Nehiri T, Duong Van Huyen JP, Viltard M, Fassot C, Heudes D, Freund N *et al.* (2008). Exposure to maternal diabetes induces salt-sensitive hypertension and impairs renal function in adult rat offspring. Diabetes 57: 2167–2175.

- Parente L, Perretti M (2003). Advances in the pathophysiology of constitutive and inducible cyclooxygenases: two enzymes in the spotlight. *Biochem Pharmacol* 65: 153–159.
- Porto NP, Jucá DM, Lahlou S, Coelho-de-Souza AN, Duarte GP, Magalhães PJ (2010). Effects of K<sup>+</sup> channels inhibitors on the cholinergic relaxation of the isolated aorta of adult offspring rats exposed to maternal diabetes. *Exp Clin Endocrinol Diabetes* 118: 360–363.
- Retailleau K, Belin de Chantemèle EJ, Chanoine S, Guihot AL, Vessières E, Toutain B et al. (2010). Reactive oxygen species and cyclooxygenase 2-derived thromboxane A<sub>2</sub> reduce angiotensin II type 2 receptor vasorelaxation in diabetic rat resistance arteries. *Hypertension* 55: 339–344.
- Rocha SO, Gomes GN, Forti AL, Franco MC, Fortes ZB, Cavanal MF et al. (2005). Long-term effects of maternal diabetes on vascular reactivity and renal function in the rat male offspring. *Pediatr Res* 58: 1274–1279.
- Schein PS, Loftus S (1968). Streptozotocin: depression of mouse liver pyridine nucleotides. *Cancer Res* 28: 1501–1506.
- Segar EM, Norris AW, Yao J, Hu S, Koppenhafer SL, Roghair RD et al. (2009). Programming of growth, insulin resistance and vascular dysfunction in offspring of late gestation diabetic rats. *Clin Sci (Lond)* 117: 129–138.
- Siman CM, Eriksson UJ (1997). Vitamin E decreases the occurrence of malformations in the offspring of diabetic rats. *Diabetes* 46: 1054–1055.
- Simeoni U, Barker DJ (2009). Offspring of diabetic pregnancy: long-term outcomes. *Semin Fetal Neonatal Med* 14: 119–124.
- Tian X, Wong WT, Leung FP, Zhang Y, Wang YX, Lee HK et al. (2011). Oxidative stress-dependent cyclooxygenase-2-derived prostaglandin F<sub>2α</sub> impairs endothelial function in renovascular hypertensive rats. *Antioxid Redox Signal* 16: 363–373.
- Virdis A, Colucci R, Versari D, Ghisu N, Fornai M, Antonioli L et al. (2009). Atorvastatin prevents endothelial dysfunction in mesenteric arteries from spontaneously hypertensive rats: role of cyclooxygenase 2-derived contracting prostanoids. *Hypertension* 53: 1008–1016.
- Vlassara H, Fu H, Makita Z, Krungkrai S, Cerami A, Bucala R (1992). Exogenous advanced glycosylation end products induce complex vascular dysfunction in normal animals: a model for diabetic and aging. *Proc Natl Acad Sci USA* 89: 12043–12047.
- World Health Organization (2002). Integrated Management of Cardiovascular Risk. WHO CVD Program: Geneva.
- Wichi RB, Souza SB, Casarini DE, Morris M, Barreto-Chaves ML, Irigoyen MC (2005). Fetal physiological programming increased blood pressure in the offspring of diabetic mothers. *Am J Physiol Regul Integr Comp Physiol* 288: 1129–1133.
- Xavier FE, Aras-López R, Arroyo-Villa I, Campo LD, Salaices M, Rossini LV et al. (2008). Aldosterone induces endothelial dysfunction in resistance arteries from normotensive and hypertensive rats by increasing thromboxane A<sub>2</sub> and prostacyclin. *Br J Pharmacol* 154: 1225–1235.
- Xavier FE, Blanco-Rivero J, Ferrer M, Balfagón G (2009). Endothelium modulates vasoconstrictor response to prostaglandin I<sub>2</sub> in rat mesenteric resistance arteries: interaction between EP<sub>1</sub> and TP receptors. *Br J Pharmacol* 158: 1787–1795.

## 6. ARTIGO 2

# Increased Cyclooxygenase-2-Derived Prostanoids Contributes to the Hyperreactivity to Noradrenaline in Mesenteric Resistance Arteries from Offspring of Diabetic Rats

Fernanda E. Ramos-Alves<sup>3</sup>, Diego B. de Queiroz<sup>3</sup>, Juliana Santos-Rocha, Gloria P. Duarte, Fabiano E. Xavier\*

Departamento de Fisiologia e Farmacologia, Centro de Ciências Biológicas, Universidade Federal de Pernambuco, Recife, Brazil

## Abstract

This study analyzed the effect of *in utero* exposure to maternal diabetes on contraction to noradrenaline in mesenteric resistance arteries (MRA) from adult offspring, focusing on the role of cyclooxygenase (COX)-derived prostanoids. Diabetes in the maternal rat was induced by a single injection of streptozotocin (50 mg/kg body weight) on day 7 of pregnancy. Contraction to noradrenaline was analyzed in isolated MRA from offspring of diabetic (O-DR) and non-diabetic (O-CR) rats at 3, 6 and 12 months of age. Release of thromboxane A<sub>2</sub> (Tx<sub>A</sub><sub>2</sub>) and prostaglandins E<sub>2</sub> (PGE<sub>2</sub>) and F<sub>2α</sub> (PGF<sub>2α</sub>), was measured by specific enzyme immunoassay kits. O-DR developed hypertension from 6 months of age compared with O-CR. Arteries from O-DR were hyperactive to noradrenaline only at 6 and 12 months of age. Endothelial removal abolished this hyperreactivity to noradrenaline between O-CR and O-DR. Preincubation with either the COX-1/2 (indomethacin) or COX-2 inhibitor (NS-398) decreased noradrenaline contraction only in 6- and 12-month-old O-DR, while it remained unmodified by COX-1 inhibitor SC-560. In vessels from 6-month-old O-DR, a similar reduction in the contraction to noradrenaline produced by NS-398 was observed when TP and EP receptors were blocked (SQ29548+AH6809). In 12-month-old O-DR, this effect was only achieved when TP, EP and FP were blocked (SQ29548+AH6809+AL8810). Noradrenaline-stimulated TxB<sub>2</sub> and PGE<sub>2</sub> release was higher in 6- and 12-month-old O-DR, whereas PGF<sub>2α</sub> was increased only in 12-month-old O-DR. Our results demonstrated that *in utero* exposure to maternal hyperglycemia in rats increases the participation of COX-2-derived prostanoids on contraction to noradrenaline, which might help to explain the greater response to this agonist in MRA from 6- and 12-month-old offspring. As increased contractile response in resistance vessels may contribute to hypertension, our results suggest a role for these COX-2-derived prostanoids in elevating vascular resistance and blood pressure in offspring of diabetic rats.

**Citation:** Ramos-Alves FE, de Queiroz DB, Santos-Rocha J, Duarte GP, Xavier FE (2012) Increased Cyclooxygenase-2-Derived Prostanoids Contributes to the Hyperreactivity to Noradrenaline in Mesenteric Resistance Arteries from Offspring of Diabetic Rats. PLoS ONE 7(11): e50593. doi:10.1371/journal.pone.0050593

**Editor:** Yu Huang, The Chinese University of Hong Kong, Hong Kong

**Received** August 21, 2012; **Accepted** October 22, 2012; **Published** November 28, 2012

**Copyright:** © 2012 Ramos-Alves et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This work was supported by grants from Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco. FE Ramos-Alves and DB de Queiroz were supported by a master degree fellowship award from Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco. J Santos-Rocha was supported by a scientific initiation fellowship award from Programa de Bolsas de Iniciação Científica/Universidade Federal de Pernambuco. GP Duarte and FE Xavier are recipients of a research fellowship from Conselho Nacional de Desenvolvimento Científico e Tecnológico. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: fabianoxavier@ufpe.br

• These authors contributed equally to this work.

## Introduction

Experimental and epidemiological studies have provided evidence that intrauterine exposure to maternal hyperglycemia produces several effects on offspring including insulin resistance, change in glucose metabolism and increased risk of hypertension in adult life [1–5]. This phenomenon termed “fetal programming” refers to the observation that an adverse environmental stimulus experienced *in utero* during the critical period of development correlates with a number of adult chronic diseases, including coronary heart disease, stroke and hypertension [6,7]. In the offspring of diabetic rats, the hyperglycemia-programmed hypertension is age-dependent [1,8,9] and has been partially attributed to baroreflex dysfunction [10] and nephron deficiency [4,8,11]. In

addition, some studies have suggested that functional change in the vasculature could also contribute to increase blood pressure on offspring of diabetic rats [3,8]. These functional vascular changes are characterized by impaired endothelial function [8,9,12] and increased contractile responses to vasoconstrictor agents, such as noradrenaline and serotonin [2,12]. Recently, we have demonstrated that exposure to maternal hyperglycemia is associated with age-dependent vascular cyclooxygenase (COX)-2 up-regulation and enhanced formation of contracting prostanoids, which impairs the endothelium-dependent relaxation in resistance arteries from adult offspring [9].

COX catalyzes conversion of arachidonic acid to prostanoids, which is known as important inflammatory response mediators. In addition, endothelial prostanoids production contributes to vas-

cular tone regulation [13]. Two isoforms of COX have been identified in mammalian cells. COX-1 is constitutively expressed in most tissues, such as vascular endothelial cells, and is involved in homeostasis maintenance. Although COX-2 is expressed at low or undetectable level, it is readily upregulated by inflammatory, mitogenic and physical stimuli. Vascular COX-2 expression has been reported in several pathological conditions associated with cardiovascular risk, such as hypertension, diabetes, atherosclerosis, obesity and aging [13–20]. In addition to its contribution to impairment of the endothelium-dependent relaxations [9,17,19], COX-2-derived prostanoids have also been recognized as an important factor causing increased contraction in isolated arteries from hypertensive [15] and diabetic [18] rats. However, the impact of these endothelial factors on vascular contraction is specific to each experimental model or vascular bed studied. In this sense, we [19,21] and others [22] have demonstrated that in resistance arteries from spontaneously hypertensive rats the COX-mediated impairment of endothelium-dependent relaxation is accompanied by uncharged noradrenaline-induced contraction. In resistance arteries from offspring of diabetic rats the contribution of these prostanoids to noradrenaline-induced contraction is still unknown. Once sympathetic vasoconstriction in resistance arteries is an important peripheral blood pressure regulator, this study has important implications with regard to arterial diameter regulation and blood pressure in offspring of diabetic rats.

In view of that, the present experiments were designed to study whether *in utero* exposure to maternal hyperglycemia alters contractile responses to noradrenaline in resistance arteries from adult offspring as well as the possible role of COX-2-derived prostanoids in this effect. Considering that in offspring of diabetic rats the endothelial function change with age [9], in the current study we also investigated the influence of age on noradrenaline responses.

## Methods

### Animals

All experimental procedures were approved by the Animal Care and Use Committee of the *Universidade Federal de Pernambuco* (approval reference number: 23076.015755/2008-96) and were conducted in accordance with the Guide for the Care and Use of Laboratory Animals published by the US National Institute of Health (NIH Publication No. 85-23, revised 1996).

Adult offspring of control (O-CR) and diabetic (O-DR) pregnant Wistar rats were studied. Diabetes mellitus in the maternal rat was induced experimentally by a single i.p. injection of streptozotocin (50 mg/kg body weight) on day 7 of pregnancy. The diabetes was confirmed by measuring plasma glucose concentrations (ACCU-CHEK®, Roche Diagnostics, Mannheim, Germany) on day 14 of pregnancy. Only pregnant females whose plasma glucose ranged between 250 and 500 mg/dL were included in the study. After birth, each litter was randomly reduced to six pups and restricted to male offspring only. The excess pups were sacrificed by CO<sub>2</sub> inhalation followed by cervical dislocation. When the male number was not enough to complete six, females were used but discarded at weaning. All rats were housed at a constant room temperature (21°C), humidity (55%) and light cycle (12 h light/dark) and had free access to tap water and standard rat chow *ad libitum*. In this study we used O-CR and O-DR with 3, 6 and 12 months of age.

### Arterial Blood Pressure Measurement

Rats were anesthetized with ketamine, xylazine and acetopromazin mixture (64.9; 3.2 and 0.78 mg/Kg<sup>-1</sup>, respectively, *i.p.*).

The right carotid artery was cannulated with a polyethylene catheter (PE-50) that was exteriorized in the mid scapular region. Adequacy of anaesthesia was assessed by monitoring withdrawal reflexes. After 24 hours, arterial pressure was measured in conscious, freely moving rats. The arterial cannula was connected to a transducer and pressure signals were recorded for a 60-min period using a data acquisition system model PowerLab 4/35 (ADIInstruments Pty Ltd, Castle Hill, Australia).

### Vessel Preparation

Rats were anesthetized with ketamine, xylazine and acetopromazin mixture (64.9; 3.2 and 0.78 mg/Kg<sup>-1</sup>, respectively, *i.p.*) and killed by exsanguination. The mesenteric vascular bed was removed and placed in cold (4°C) Krebs Henseleit solution (KHS; in mmol/L: 115 NaCl, 2.5 CaCl<sub>2</sub>, 4.6 KCl, 1.2 KH<sub>2</sub>PO<sub>4</sub>, 1.2, MgSO<sub>4</sub>.7H<sub>2</sub>O, 25 NaHCO<sub>3</sub>, 11.1 glucose, and 0.03 EDTA). For reactivity experiments the third order branch of the mesenteric arcade was dissected and cut in segments of approximately 2 mm in length. Segments of mesenteric resistance arteries were mounted in a small vessel chamber myograph (Danish Myo Technology A/S, Aarhus, Denmark) to measure isometric tension according to the method described by Mulvany and Halpern [23]. After a 15-min equilibration period in oxygenated KHS at 37°C and pH 7.4, segments were stretched to their optimal lumen diameter for active tension development. Optimal lumen diameter was determined based on the internal circumference/wall tension ratio of the segments by setting the internal circumference, L<sub>0</sub>, to 90% of what the vessels would have if they were exposed to a passive tension equivalent to that produced by a transmural pressure of 100 mmHg [23]. Optimal lumen diameter was determined using specific software for normalization of resistance arteries (DMT Normalization Module; ADIInstruments Pty Ltd, Castle Hill, Australia). Segments were washed with KHS and left to equilibrate for 30 min. Vessel contractility was then tested by an initial exposure to a high-K<sup>+</sup> (120 mmol/L) solution.

### Experimental Protocols

After washout, segments were contracted with a concentration of noradrenaline that induced approximately 50% of the KCl contraction, and then acetylcholine (1 μmol/L) was added to assess the integrity of the endothelium. After 60 min, cumulative concentration-response curves for noradrenaline (10 nmol/L - 0.1 mmol/L) were generated. The role of the endothelium in the noradrenaline-induced contraction was evaluated in segments subjected to mechanical endothelium removal. The absence of endothelium was confirmed by the inability of acetylcholine (1 μmol/L) to induce relaxation and smooth muscle integrity was confirmed by the maintenance of the KCl- (120 mmol/L) induced contraction. The effect of the nonselective nitric oxide synthase inhibitor N-nitro-l-arginine methyl ester (L-NAME, 0.1 mmol/l) on concentration-response curves for noradrenaline was investigated.

The possible role of cyclooxygenase metabolites in noradrenaline-induced contraction was investigated in segments preincubated with either indomethacin (a COX-1 and COX-2 inhibitor, 10 μmol/L), [5-(4-chlorophenyl)-1-(4-methoxyphenyl)-3-trifluoromethylpyrazole] SC-560 (a COX-1 inhibitor, 1 μmol/L), N-(2-cyclohexyloxy-4-nitrophenyl) methansulfonamide (NS-398, a COX-2 inhibitor, 10 μmol/L), [1S-[1a,2a(Z),3a,4a]]-7-[3-[[2-(phenylamino) carbonyl]hydrazino]methyl]-7-oxabicyclo [2.2.1] hept-2-yl]-5-heptanoic acid (SQ29548, a TxA<sub>2</sub> receptor (TP) antagonist, 1 μmol/L), 6-Isopropoxy-9-oxoanthene-2-carboxylic acid (AH6809, a PGE<sub>2</sub> receptor (EP<sub>1</sub>, EP<sub>2</sub> and EP<sub>3</sub>) antagonist, 30 μmol/L) or 9α,15R-dihydroxy-11β-fluoro-15-(2,3-

dihydro-1H-inden-2-yl)-16,17,18,19,20-pentanor-prosta-5Z,13E-dien-1-oic acid (AL8810, a PGF<sub>2α</sub> receptor (FP) antagonist, 10 μmol/L). All drugs were added 30 min before the concentration-response curve to noradrenaline.

In another set of experiments, the vasoactive response to the TP receptor agonist (15)-hydroxy-11,9-(epoxymethano)prosta-5,13-dienoic acid (U46619, 1 nmol/L - 10 μmol/L), PGE<sub>2</sub> (10 nmol/L - 10 μmol/L) or PGF<sub>2α</sub> (10 nmol/L to 10 μmol/L), was analyzed in endothelium-denuded arteries from 6- and 12-month old rats.

### Prostanoids Production

To measure the release of TxA<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2α</sub>, we used specific enzyme immunoassay kits (Cayman Chemical Company, Ann Arbor, MI, USA). The second, third and fourth branches of mesenteric artery were preincubated for 45 min in 200 μl of KHS at 37°C and continuously gassed with a 95% O<sub>2</sub> - 5% CO<sub>2</sub> mixture (stabilization period). Afterward, three washout periods of 7 min in a bath of 200 μl of KHS were run before incubation with noradrenaline (10 nmol/L - 0.1 mmol/L). The medium was collected 10 min after the last dose of noradrenaline. The different assays were performed following the manufacturer's instructions. Results were expressed as pg.ml<sup>-1</sup> mg wet tissue<sup>-1</sup>.

### Drugs

Drugs used were noradrenaline hydrochloride, acetylcholine chloride, indomethacin, L-NAME (Sigma; St. Louis, MO, U.S.A.), NS-398, SC-560, SQ29548, AH6809, AL8810, PGE<sub>2</sub>, PGF<sub>2α</sub> (Cayman Chemical Company; Ann Arbor, Michigan, U.S.A.) and U46619 (Calbiochem-Novabiochem GmbH). Stock solutions of acetylcholine was made in distilled water, noradrenaline was dissolved in a NaCl (0.9%)-ascorbic acid (0.01% vv<sup>-1</sup>) solution, indomethacin, SQ29548, PGE<sub>2</sub> and PGF<sub>2α</sub> were dissolved in ethanol and AH6809, AL8810 and U46619, which were dissolved in dimethyl sulfoxide. These solutions were kept at -20°C and appropriate dilutions were made on the day of the experiment.

### Statistical Analysis

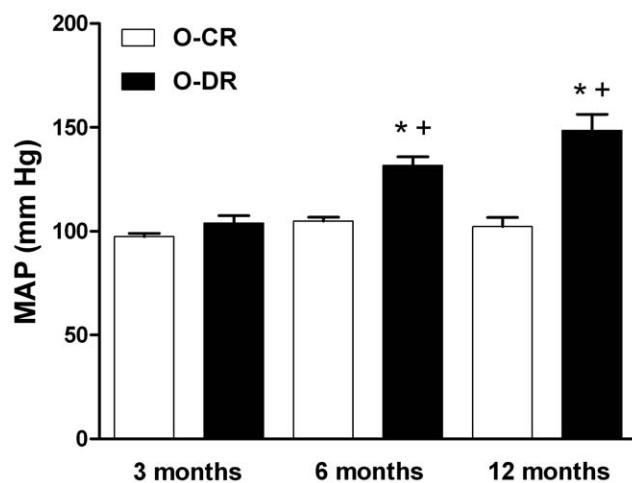
Noradrenaline, U46619, PGE<sub>2</sub> and PGF<sub>2α</sub> contractile responses were expressed as a percentage of the maximum response produced by KCl.

All values are expressed as means±S.E.M. of the number of animals used in each experiment. Results were analyzed using two-way ANOVA for comparison between groups. When ANOVA showed a significant treatment effect, Bonferroni's post hoc test was used to compare individual means. Differences were considered statistically significant at P<0.05.

### Results

O-DR presented higher blood pressure in adulthood. Although the mean arterial pressure of 3-month-old rats was similar in both groups (Figure 1) it was significantly increased in both 6 and 12 months (Figure 1) of age in O-DR compared to O-CR. The heart rate was similar in all O-DR groups compared to their respective age-matched O-CR (results not shown).

KCl (120 mmol/L) evoked similar contractions in vessels from all groups studied (3-month-old rats, O-CR: 3.15±0.04 vs. O-DR: 3.19±0.07 mN/mm; 6-month-old rats, O-CR: 3.28±0.12 vs. O-DR: 3.19±0.09 mN/mm; 12-month-old rats, O-CR: 3.21±0.02 vs. O-DR: 3.25±0.13 mN/mm; ANOVA (two-way), P>0.05). However, exposure to maternal diabetes promoted an increase in the contractile response to noradrenaline in a time-dependent manner in arteries from the offspring (Figure 2). While in arteries from 3-month-old O-DR the noradrenaline-induced contraction



**Figure 1. Effect of maternal diabetes on blood pressure levels in the offspring.** Mean arterial pressure (MAP) significantly increased from 6 months in the offspring from diabetic mothers (O-DR) compared to control mother offspring group (O-CR). Results are expressed as means±SEM, N=7–8 animals in each group. ANOVA (two-way): \*P<0.05 O-DR vs. age-matched O-CR; +P<0.05 6- or 12-month-old O-DR vs. 3-month-old O-DR.

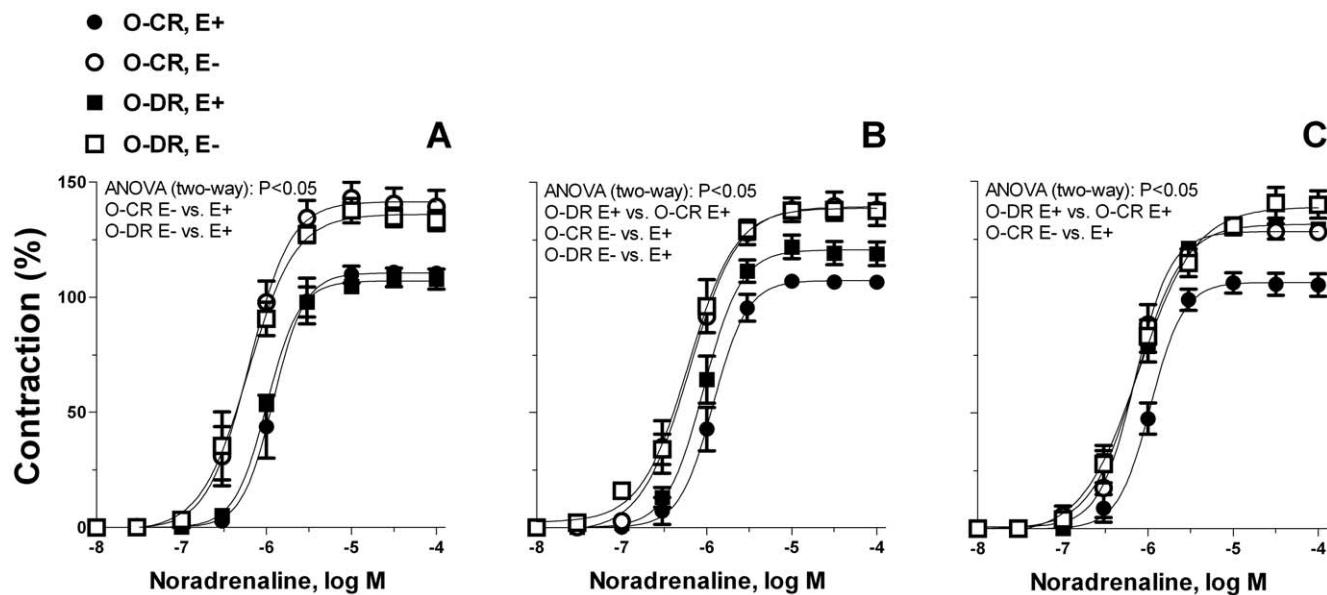
doi:10.1371/journal.pone.0050593.g001

remained unmodified (Figure 2A), in both 6- and 12-month-old O-DR this response was increased when compared to age-matched O-CR (Figure 2B and C). In arteries without endothelium the contractile responses produced by noradrenaline were similar in all groups studied (Figure 2).

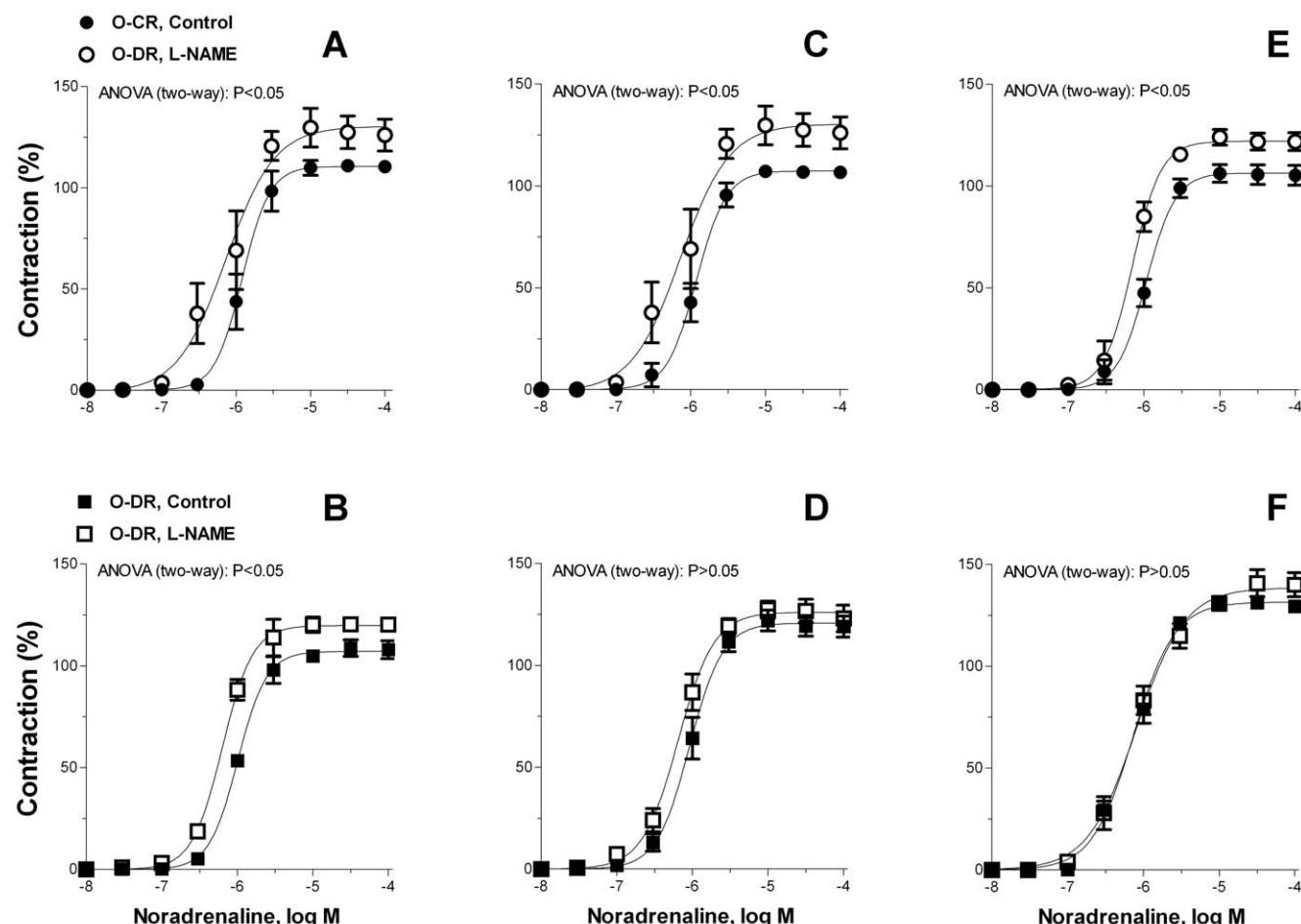
To assess the contribution of endothelium-derived nitric oxide to the noradrenaline responses, segments were incubated with the nitric oxide synthase inhibitor L-NAME. This drug similarly increased the response to noradrenaline in arteries from 3-, 6- and 12-month-old O-CR and 3-month old O-DR. (Figure 3). In arteries from 6- and 12-month-old O-DR L-NAME failed to produce significant increases of noradrenaline-induced contraction (Figure 3).

In mesenteric resistance arteries from 6- and 12-months old rats, incubation with either the unspecific COX inhibitor indomethacin or the specific COX-2 inhibitor NS-398 decreased the contractile response to noradrenaline only in O-DR (Figure 4C and D). In 3-month-old O-CR and O-DR, the contraction to noradrenaline did not change in presence of indomethacin or NS-398 (Figure 4A and B). Preincubation with the specific COX-1 inhibitor SC-560 did not alter the contraction to noradrenaline in any experimental group (results not shown).

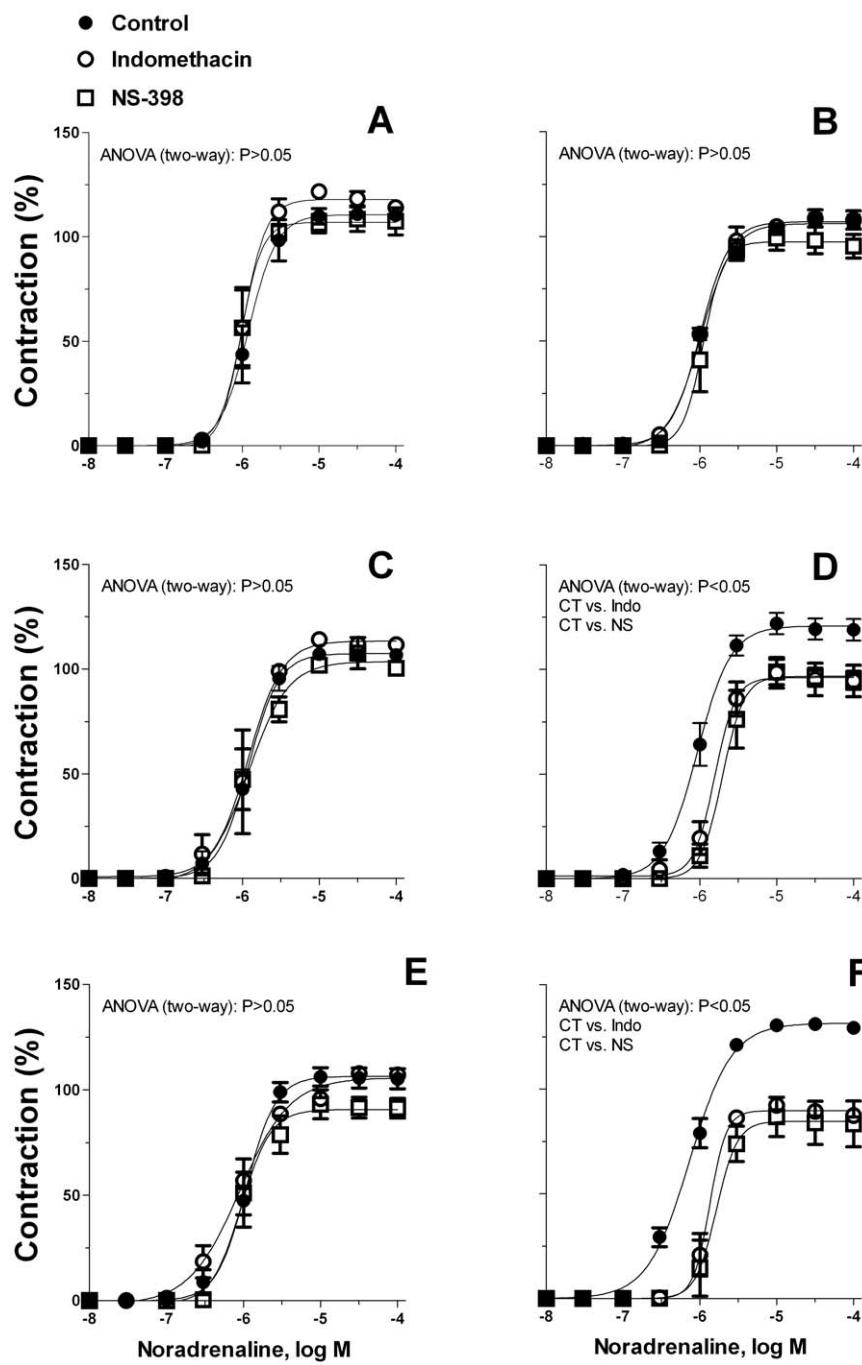
To identify the contractile prostanoid involved in noradrenaline responses, arteries from 6- and 12-month-old O-DR were incubated with the TP receptor antagonist SQ29548, the EP<sub>1</sub>, EP<sub>2</sub> and EP<sub>3</sub> antagonist AH6809 and the FP antagonist AL8810. In 6-month-old O-DR, SQ29548 decreased contraction to noradrenaline, but its effect was lower than to those obtained with NS-398 (Compare figure 4D with 5A). When co-incubated with SQ29548, AH6809 reduced the response to noradrenaline to similar values to those obtained with NS-398. AL8810 failed to decrease the effect of SQ29548 plus AH6809 (Figure 5A). In arteries from 12-month-old O-DR, SQ29548 partly decreased contraction to noradrenaline (Figure 5B). In arteries from this group, although co-incubation with AH6809 have increased the effect of SQ29548, an additional decreased of the noradrenaline response was observed when arteries were treated with SQ29548 in combination with AH6809 and AL8810 (Figure 5B).



**Figure 2. Concentration-dependent contraction to noradrenaline in endothelium-intact (E+) and denuded (E-) mesenteric resistance arteries from 3- (A), 6- (B) and 12-month-old offspring of diabetic (O-DR) and non-diabetic rats (O-CR). Results (mean±S.E.M.) were expressed as a percentage of the initial contraction elicited by KCl. N=6–7 animals each curve.**



**Figure 3. Effect of L-NAME (0.1 mmol/L) on the concentration dependent contraction to noradrenaline in mesenteric resistance segments (A) 3-month-old O-CR, (B) 3-month-old O-DR, (C) 6-month-old O-CR, (D) 6-month-old ODR, (E) 12-month-old O-CR and (F) 12-month-old ODR. Results (mean±S.E.M.) are expressed as a percentage of the initial contraction elicited by KCl. N=7–8 animals each curve.**



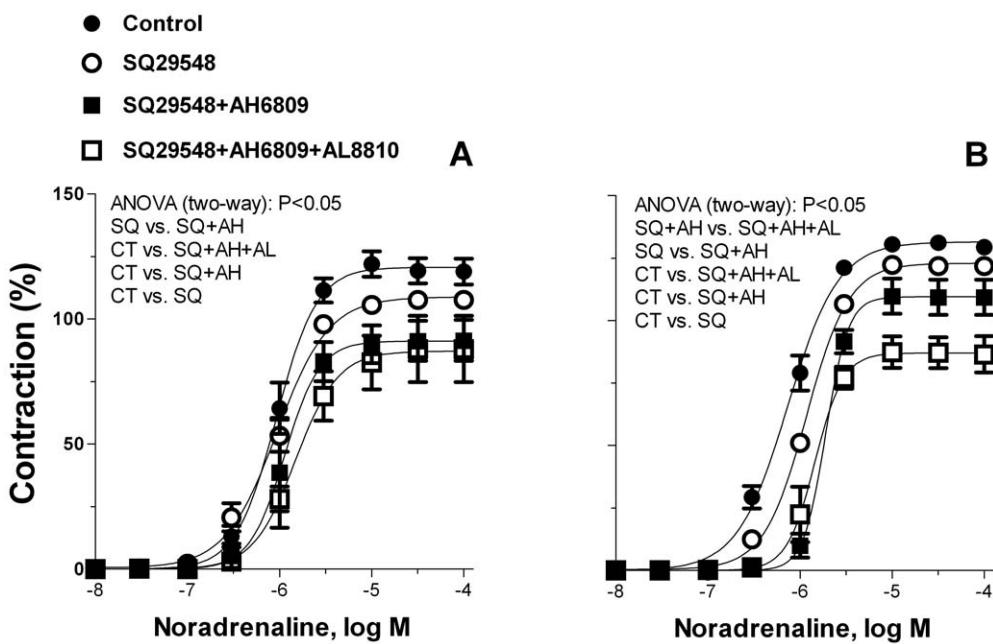
**Figure 4. Effect of indomethacin (Indo, 10  $\mu\text{mol/L}$ ) or NS-398 (NS, 10  $\mu\text{mol/L}$ ) on the concentration dependent contraction to noradrenaline in mesenteric resistance segments (A) 3-month-old O-CR, (B) 3-month-old O-DR, (C) 6-month-old O-CR, (D) 6-month-old ODR, (E) 12-month-old O-CR and (F) 12-month-old ODR. Results (mean  $\pm$  S.E.M.) are expressed as a percentage of the initial contraction elicited by KCl. N = 6–7 animals each curve.**

doi:10.1371/journal.pone.0050593.g004

U46619, PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub>  caused cumulative concentration-dependent contractions in quiescent resistance arteries. These contractions were not significantly different between arteries of O-CR and O-DR (Figure 6). Contraction to U46619, PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub>  were inhibited by SQ29548, AH6809 and AL8810, respectively (Figure 6).

In mesenteric resistance arteries from all groups, noradrenaline increased the release of TxB<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub>

(Figure 7). The noradrenaline-stimulated TxB<sub>2</sub> levels were higher in arteries from both 6- and 12-month-old O-DR compared to their respective age-matched O-CR (Figure 7A). In arteries from 6- and 12-month-old O-DR, the levels of noradrenaline-stimulated PGE<sub>2</sub> were higher compared to O-CR (Figure 7B and C). The noradrenaline-stimulated PGF<sub>2 $\alpha$</sub>  levels were higher only in arteries from 12-month-old O-DR (Figure 7C).



**Figure 5. Effect of SQ29548 (SQ, 1  $\mu\text{mol/L}$ ) alone or in association with AH6809 (AH, 30  $\mu\text{mol/L}$ ) or AH6809 plus AL8810 (AL, 10  $\mu\text{mol/L}$ ) on the concentration-dependent contraction to noradrenaline in mesenteric resistance segments from 6- (B) and 12-month (mo)-old (C) diabetic (O-DR) offspring rats.** Results (mean  $\pm$  S.E.M.) are expressed as a percentage of the initial contraction elicited by KCl. N=6–7 animals in each group.

doi:10.1371/journal.pone.0050593.g005

## Discussion

Results presented here demonstrate that *in utero* exposure to maternal hyperglycaemia in rats increases the participation of COX-2-derived TxA<sub>2</sub>, PGE<sub>2</sub> and PGF<sub>2 $\alpha$</sub>  on contraction to noradrenaline. This participation, associated to decrease in nitric oxide modulation, might help to explain the greater noradrenaline responses observed in mesenteric resistance arteries from 6- and 12-month-old offspring of diabetic rats, which could partially contribute to the development of hypertension and vascular disease in these rats.

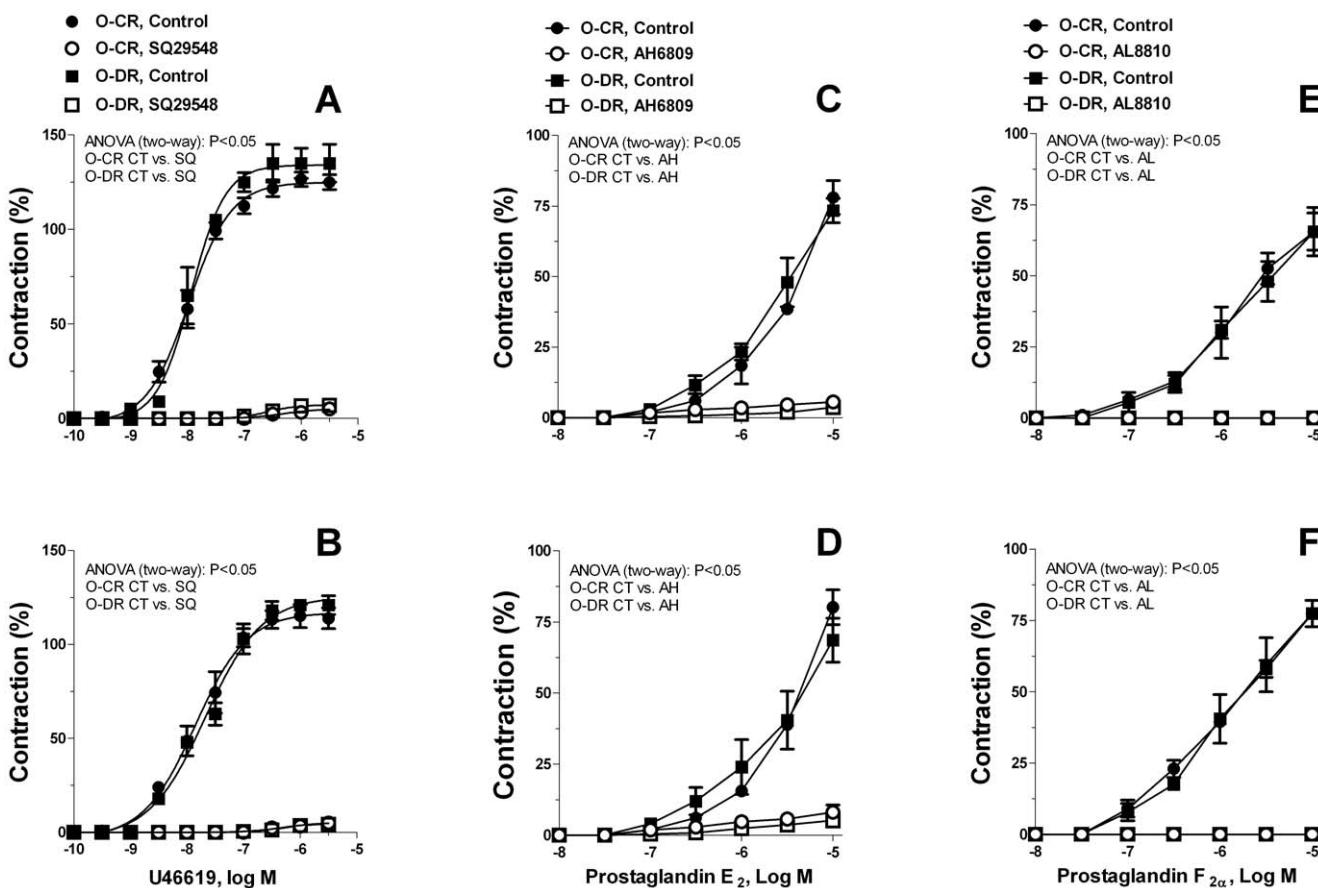
The intrauterine exposure to maternal hyperglycaemia is a significant risk factor for development of metabolic syndrome components, including glucose intolerance, insulin resistance and hypertension [5,24]. The mechanisms of this hyperglycemia-programmed hypertension are complex and involve renal, neural and vascular factors [1,2,4,8,10]. One of the most remarkable vascular changes in rats born from diabetic mothers is the reduction of endothelium-dependent relaxation [2,8,12]. Ingram *et al.* [25] reported that *in utero* exposure to diabetic environment severely results in general decline in proliferative capacity of fetal circulating endothelial progenitor cells, producing a decrease in the number of endothelial cells and their premature senescence. These alterations may predispose infants born from diabetic mothers to develop endothelial dysfunction and ultimately cardiovascular disease.

In a recent study we reported that offspring of diabetic rats manifest glucose intolerance, insulin resistance and hypertension at 6 and 12 months of age [9]. In addition, resistance arteries from these rats also displayed impaired endothelial function, associated with increased COX-2-derived vasoconstrictor prostanoids formation [9]. Several studies have demonstrated a correlation between endothelial dysfunction and the hyperreactivity to the vasoconstrictor agents [15,26–28]. Herein, we have provided clear evidence that compared to O-CR, mesenteric resistance arteries

isolated from O-DR is indeed hyperreactive to noradrenaline. This effect is endothelium-dependent, since in endothelium-denuded arteries noradrenaline contraction was similar between O-CR and O-DR. The unmodified K<sup>+</sup>-induced contraction in resistance arteries from O-DR suggests no alteration in contractile ability of the smooth muscle. Our results also show that the increased response to noradrenaline in arteries from O-DR correlates with changes of blood pressure in these rats, suggesting that this mechanism could be responsible for the enhanced arteriolar tone and consequently elevated systemic blood pressure in these rats.

The next objective was to determine possible quantitative or qualitative differences in the participation of endothelial factors in this contractile response in both O-CR and O-DR. Nitric oxide synthesis inhibition (L-NAME) increased the vasoconstriction to noradrenaline in arteries from all O-CR groups, while in 6- and 12-month-old O-DR it failed to produce significant increases of noradrenaline-induced contraction. This suggests a reduction on nitric oxide release/bioavailability in arteries from 6- and 12-month-old O-DR, which in turn could contribute to the augmented noradrenaline response in these rats.

COX-1 and COX-2 activity may contribute to underlying vascular hyperreactivity in some cardiovascular disease models [15,18,29–32]. Many studies have shown that COX-2 is induced during some inflammatory process and prostanoids produced by this isoform are responsible for many inflammatory signs [13,33]. In this sense, COX-2-derived prostanoids have been shown to be associated with the development of vascular complications under insulin resistance conditions and cardiovascular risk [13,14,16,20,34,35]. Results presented here and previously [9] provide several lines of evidence suggesting that COX-2 contributes to the hyperreactivity to noradrenaline in mesenteric resistance arteries from O-DR. First, the immunoblot analysis clearly shows that COX-2, but not COX-1, is up-regulated in



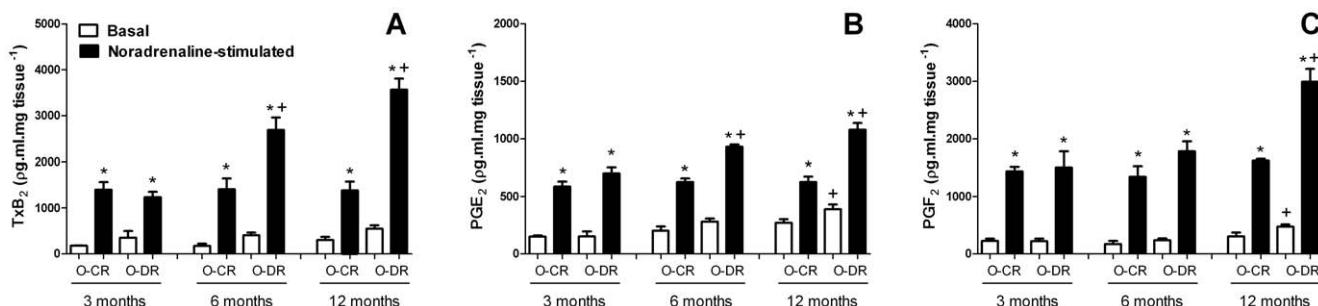
**Figure 6. Concentration-dependent contraction to U46619 (A and B), PGE<sub>2</sub> (C and D) and PGF<sub>2α</sub> (E and F) in endothelium-denuded mesenteric resistance arteries from 6- (A, C and E) and 12-month-old (B, D and F) offspring of diabetic (O-DR) and non-diabetic rats (O-CR). Curves to U46619, PGE<sub>2</sub> and PGF<sub>2α</sub> were performed in absence (Control, CT) and in presence of SQ29548 (SQ), AH6809 (AH) or AL8810 (AL), respectively.** Results (mean±S.E.M.) are expressed as a percentage of the initial contraction elicited by KCl. N=4–6 animals in each group.

doi:10.1371/journal.pone.0050593.g006

arteries from O-DR [9]. Second, the time course of COX-2 up-regulation [9] coincides with the appearances of hyperreactivity to noradrenaline in O-DR (6 and 12 months of age). Third, selective inhibition of COX-2 activity with indomethacin or NS-398, but not the inhibition of COX-1 with SC-560, significantly reduced the noradrenaline-induced contractile responses in arteries isolated from 6- and 12-month-old O-DR, whereas these drugs did not

affect the contractions in arteries from O-CR. Taken together, these results suggest that the selective COX-2 up-regulation in mesenteric resistance arteries is responsible for the contractile hyperreactivity in offspring of diabetic rats.

Data from our group [9,19] and others [15,35,36,37] have showed that COX-2 up-regulation is associated with increased production of the vasoconstrictor prostanooids, such as TxA<sub>2</sub>. This



**Figure 7. Basal and noradrenaline-stimulated production of thromboxane B<sub>2</sub> (TxB<sub>2</sub>, A), prostaglandin E<sub>2</sub> (PGE<sub>2</sub>, B) and prostaglandin F<sub>2α</sub> (PGF<sub>2α</sub>, C) in mesenteric resistance segments from 3-, 6- and 12-month-old control (O-CR) and diabetic (O-DR) offspring rats.** N=6–7 animals in each group. ANOVA (two-way): \*P<0.05 noradrenaline-stimulated vs. basal; +P<0.05 O-DR vs. age-matched O-CR.

doi:10.1371/journal.pone.0050593.g007

denotes that the increased production of  $\text{TxA}_2$  is one potential mechanism mediating COX-2-induced vascular hyperreactivity. Indeed, TP receptor blockade by SQ29548 diminished the vascular smooth muscle hyperreactivity in both 6- and 12-month-old O-DR. In addition, noradrenaline-stimulated release of  $\text{TxB}_2$  (the  $\text{TxA}_2$  metabolite) was increased in arteries from those rats. However, it should be noted that in arteries from 6 and 12-month-old O-DR the decrease in noradrenaline-induced contraction produced by SQ29548 was lower than that observed in the presence of indomethacin or NS-398. In mesenteric arteries from 6-month-old O-DR, reduction in the noradrenaline response to the level produced by COX-2 inhibition occurred when these vessels were preincubated with SQ29548 plus EP receptor blocker, AH6809. Among 12-month-old O-DR the combined treatment of SQ29548 plus AH6809 and AL8810 (a FP receptor antagonist) was required to reduce noradrenaline response to the level produced by NS-398 or indomethacin. This indicates that in resistance arteries from 6-month-old O-DR the increase in noradrenaline-induced contraction was produced by an increase in both  $\text{TxA}_2$  and  $\text{PGE}_2$  while in arteries from 12-month-old O-DR the hyperactivity to noradrenaline was produced by an increase in  $\text{PGE}_2$ ,  $\text{TxA}_2$ , and  $\text{PGF}_{2\alpha}$  release. Consistent with these functional findings, in addition to  $\text{TxA}_2$ , noradrenaline-stimulated  $\text{PGE}_2$  release was also higher in arteries from 6- and 12-month-old O-DR. Besides these two prostanoids, noradrenaline-stimulated  $\text{PGF}_{2\alpha}$  was also increased in arteries from 12-month-old O-DR. The fact that exogenous  $\text{PGE}_2$ ,  $\text{PGF}_{2\alpha}$  or TP receptor agonist U46616 administration have produced similar contraction in endothelium-denuded arteries in all groups studied discard the possibility that EP, FP or TP receptor initiated signaling mechanisms are altered in mesenteric resistance arteries from O-DR.

As reported by several studies, increased activation of TP, EP and/or FP receptors represents a key mechanism to increased vascular tone and blood pressure in several models of cardiovascular disease, including hypertension and diabetes [38–46]. Rutkai *et al.* [38] demonstrated that EP receptor activation in resistance arteries may contribute to the development of high blood pressure in type-2 diabetic mice. In these animals the augmented pressure- and angiotensin II-induced arteriolar tone, as well as the elevated systolic blood pressure were normalized by the EP antagonist, AH6809. Similarly, Guan *et al.* [39] demonstrated that EP receptor activation contributes to hypertension in two well-established experimental models: SHR and chronic angiotensin II-infused mice. Respect to  $\text{TxA}_2$ , various studies have reported

## References

- Nehiri T, Duong Van Huyen JP, Viltard M, Fassot C, Heudes D, et al. (2008) Exposure to maternal diabetes induces salt-sensitive hypertension and impairs renal function in adult rat offspring. *Diabetes* 57: 2167–2175.
- Segar EM, Norris AW, Yao JR, Hu S, Koppenhafer SL, et al. (2010) Programming of growth, insulin resistance and vascular dysfunction in offspring of late gestation diabetic rats. *Clin Sci* 117: 129–138.
- Simeoni U, Barker DJ (2009) Offspring of diabetic pregnancy: Long-term outcomes. *Seminars in Fetal & Neo Med* 14: 119–124.
- Chen YW, Chenier I, Tran S, Scotcher M, Chang SY, et al. (2010) Maternal diabetes programs hypertension and kidney injury in offspring. *Pediatr Nephrol* 25: 1319–1329.
- Blondeau B, Joly B, Perret C, Prince S, Bruneval P, et al. (2011) Exposure in utero to maternal diabetes leads to glucose intolerance and high blood pressure with no major effects on lipid metabolism. *Diabetes Metab* 37: 245–251.
- Barker DJ, Eriksson JG, Forsen T, Osmond C (2002) Fetal origins of adult disease: strength of effects and biological basis. *Int J Epidemiol* 31: 1235–1239.
- Barker DJ (2004) The developmental origins of adult disease. *J Am Coll Nutr* 23: 588–595.
- Rocha SO, Gomes GN, Forti AL, Franco MC, Fortes ZB, et al. (2005) Long-term effects of maternal diabetes on vascular reactivity and renal function in the rat male offspring. *Pediatr Res* 58: 1274–1279.
- Ramos-Alves FE, de Queiroz DB, Santos-Rocha J, Duarte GP, Xavier FE (2012) Effect of age and cyclooxygenase-2-derived prostanoids on the progression of adult vascular dysfunction in offspring diabetic rats. *Br J Pharmacol* 166: 2198–2208.
- Wichi RB, Souza SB, Casarini DE, Morris M, Barreto-Chaves ML, et al. (2005) Fetal Physiological Programming Increased blood pressure in the offspring of diabetic mothers. *Am J Physiol Regul Integr Comp Physiol* 288: 1129–1133.
- Amri K, Freund N, Vilar J, Merlet-Bénichou C, Lelièvre-Pégorier M (1999) Adverse effects of hyperglycemia on kidney development in rats: in vivo and in vitro studies. *Diabetes* 48: 2240–2245.
- Holemans K, Gerber RT, Meurrens K, de Clerck F, Poston L, et al. (1999) Streptozotocin diabetes in the pregnant rat induces cardiovascular dysfunction in adult offspring. *Diabetologia* 42: 81–89.
- Félétou M, Huang Y, Vanhoutte PM (2011) Endothelium-mediated control of vascular tone: COX-1 and COX-2 products. *Br J Pharmacol* 164: 894–912.
- Helmersson J, Vessby B, Larsson A, Basu S (2004) Association of type 2 diabetes with cyclooxygenase-mediated inflammation and oxidative stress in an elderly population. *Circulation* 109: 1729–1734.
- Alvarez Y, Briones AM, Balfagón G, Alonso MJ, Salaices M (2005) Hypertension increases the participation of vasoconstrictor prostanoids from cyclooxygenase-2 in phenylephrine responses. *J Hypertens* 23: 767–777.

that TP receptor activation contributes to the severity of blood pressure elevation as well as cardiac hypertrophy and increased vascular responsiveness in hypertensive rats [40,41,42,43].  $\text{PGF}_{2\alpha}$  has also been involved in the pathogenesis of hypertension. Tian *et al.* [44] demonstrated that COX-2-derived  $\text{PGF}_{2\alpha}$  contributes to endothelial dysfunction and elevation of blood pressure in renovascular hypertensive rats. In addition, Yu *et al.* [45] have also reported that FP receptor deletion decreased blood pressure in mice, indicating that  $\text{PGF}_{2\alpha}$  may act via FP receptor to modulate blood pressure regulation.

Results presented here provide evidences of participation of COX-2-derived  $\text{TxA}_2$ ,  $\text{PGE}_2$  and  $\text{PGF}_{2\alpha}$  in the altered regulation of mesenteric arterial responsiveness in offspring of diabetic rats. As abnormal vascular reactivity may contribute to the etiology of hypertension [47–49], our results suggest a crucial role for these COX-2-derived prostanoids in elevating vascular resistance and systemic blood pressure in offspring of diabetic rats. However, these results should be carefully interpreted once blood pressure was not analyzed after chronic COX-2 inhibition or TP, EP and FP receptors blockade. Further studies have yet to be performed to demonstrate this effect in offspring of diabetic rats.

Collectively, our results suggest that a) age-dependent hyperreactivity to noradrenaline is present in mesenteric resistance arteries of adult offspring of diabetic rats, b) it may result from a reduction on nitric oxide release/bioavailability and increased  $\text{TxA}_2$ ,  $\text{PGE}_2$  and  $\text{PGF}_{2\alpha}$  vasoconstrictor prostanoid releasing, and c) the mechanisms underlying this hyperreactivity may involve up-regulation of COX-2 [9]. However, further studies need to be carried out to determine whether and for what extent changes in mesenteric prostaglandin synthesis might contribute to blood pressure alterations in these rats. Nevertheless, changes in COX-dependent modulation of contractile response should be taken into consideration in future studies with offspring born from diabetic mothers.

## Acknowledgments

We are grateful to José Antonio de Albuquerque for his technical assistance.

## Author Contributions

Conceived and designed the experiments: FEX GPD. Performed the experiments: FERA DBQ JSR. Analyzed the data: FERA DBQ JSR. Contributed reagents/materials/analysis tools: FEX GPD. Wrote the paper: FERA DBQ FEX.

16. Bagi Z, Erdei N, Toth A, Li W, Hintze TH, et al. (2005) Type 2 diabetic mice have increased arteriolar tone and blood pressure: enhanced release of COX-2-derived constrictor prostaglandins. *Arterioscler Thromb Vasc Biol* 25: 1610–1616.
17. Blanco-Rivero J, Cachofeiro V, Lahera V, Aras-Lopez R, Marquez-Rodas I, et al. (2005) Participation of prostacyclin in endothelial dysfunction induced by aldosterone in normotensive and hypertensive rats. *Hypertension* 46: 107–112.
18. Shi Y, Vanhoutte PM (2008) Oxidative stress and COX cause hyper-responsiveness in vascular smooth muscle of the femoral artery from diabetic rats. *Br J Pharmacol* 154: 639–651.
19. Xavier FE, Aras-López R, Arroyo-Villa I, Campo LD, Salaices M, et al. (2008) Aldosterone induces endothelial dysfunction in resistance arteries from normotensive and hypertensive rats by increasing thromboxane A<sub>2</sub> and prostacyclin. *Br J Pharmacol* 154: 1225–1235.
20. Virdis A, Colucci R, Versari D, Ghislu N, Fornai M, et al. (2009) Atorvastatin prevents endothelial dysfunction in mesenteric arteries from spontaneously hypertensive rats: role of cyclooxygenase 2-derived contracting prostanoids. *Hypertension* 53: 1008–1016.
21. Xavier FE, Blanco-Rivero J, Avendaño MS, Sastre E, Yela R, et al. (2011) Aldosterone alters the participation of endothelial factors in noradrenaline vasoconstriction differently in resistance arteries from normotensive and hypertensive rats. *Eur J Pharmacol* 654: 280–288.
22. Diederich D, Yang ZH, Bühl FR, Lüscher TF (1990) Impaired endothelium-dependent relaxations in hypertensive resistance arteries involve cyclooxygenase pathway. *Am J Physiol* 258: H445–451.
23. Mulvany MJ, Halpern W (1977) Contractile properties of small arterial resistance vessels in spontaneously hypertensive and normotensive rats. *Circ Res* 41: 19–26.
24. Fetita LS, Sobngwi E, Serradas P, Calvo F, Gautier JF (2006) Consequences of fetal exposure to maternal diabetes in offspring. *J Clin Endocrinol Metab* 91: 3718–3724.
25. Ingram DA, Lien IZ, Mead LE, Estes M, Prater DN, et al. (2008) In vitro hyperglycemia or a diabetic intrauterine environment reduces neonatal endothelial colony-forming cell numbers and function. *Diabetes* 57: 724–731.
26. Dohi Y, Kojima M, Sato K (1996) Endothelial modulation of contractile responses in arteries from hypertensive rats. *Hypertension* 28: 732–737.
27. Wang D, Iversen J, Strandgaard S (1999) Contractility and endothelium-dependent relaxation of resistance vessels in polycystic kidney disease rats. *J Vasc Res* 36: 502–509.
28. Matsumoto T, Miyamori K, Kobayashi T, Kamata K (2006) Apocynin normalizes hyperreactivity to phenylephrine in mesenteric arteries from cholesterol-fed mice by improving endothelium-derived hyperpolarizing factor response. *Free Radic Biol Med* 41: 1289–1303.
29. Lagaud GJ, Masih-Khan E, Kai S, van Breemen C, Dube GP (2001) Influence of type II diabetes on arterial tone and endothelial function in murine mesenteric resistance arteries. *J Vasc Res* 38: 578–589.
30. Xavier FE, Davel AP, Rossoni LV, Vassallo DV (2003) Time-dependent hyperreactivity to phenylephrine in aorta from untreated diabetic rats: role of prostanoids and calcium mobilization. *Vascul Pharmacol* 40: 67–76.
31. Guo Z, Su W, Allen S, Pang H, Daugherty A, et al. (2005) COX-2 up-regulation and vascular smooth muscle contractile hyperreactivity in spontaneous diabetic db/db mice. *Cardiovasc Res* 67: 723–735.
32. Wenceslau CF, Davel AP, Xavier FE, Rossoni LV (2011) Long-term ouabain treatment impairs vascular function in resistance arteries. *J Vasc Res* 48: 316–326.
33. Parente L, Perretti M (2003) Advances in the pathophysiology of constitutive and inducible cyclooxygenases: two enzymes in the spotlight. *Biochem Pharmacol* 65: 153–159.
34. Elmarakby AA, Imig JD (2010) Obesity is the major contributor to vascular dysfunction and inflammation in high-fat diet hypertensive rats. *Clin Sci* 118: 291–301.
35. Retailleau K, Belin de Chantemèle EJ, Chanoine S, Guihot AL, Vessières E, et al. (2010) Reactive oxygen species and cyclooxygenase 2-derived thromboxane A<sub>2</sub> reduce angiotensin II type 2 receptor vasorelaxation in diabetic rat resistance arteries. *Hypertension* 55: 339–344.
36. Shiokoshi T, Ohsaki Y, Kawabe J, Fujino T, Kikuchi K (2002) Downregulation of nitric oxide accumulation by cyclooxygenase-2 induction and thromboxane A<sub>2</sub> production in interleukin-1beta-stimulated rat aortic smooth muscle cells. *J Hypertens* 20: 455–461.
37. Peçanha FM, Wiggers GA, Briones AM, Perez-Giron JV, Miguel M, et al. (2010) The role of cyclooxygenase (COX)-2 derived prostanoids on vasoconstrictor responses to phenylephrine is increased by exposure to low mercury concentration. *J Physiol Pharmacol* 61: 29–36.
38. Rutkai I, Feher A, Erdei N, Henrion D, Papp Z, et al. (2009) Activation of prostaglandin E<sub>2</sub> EP1 receptor increases arteriolar tone and blood pressure in mice with type 2 diabetes. *Cardiovasc Res* 83: 148–154.
39. Guan Y, Zhang Y, Wu J, Qi Z, Yang G, et al. (2007) Antihypertensive effects of selective prostaglandin E<sub>2</sub> receptor subtype 1 targeting. *J Clin Invest* 117: 2496–2505.
40. Himmelstein SI, Klotman PE (1989) The role of thromboxane in two-kidney, one-clip Goldblatt hypertension in rats. *Am J Physiol* 257: F190–196.
41. Boussairi EH, Sacquet J, Bassard J, Benzoni D (1994) Thromboxane A<sub>2</sub>-prostaglandin H<sub>2</sub> and renovascular hypertension in rats. *Am J Physiol* 267: R1190–1197.
42. Francois H, Makhanova N, Ruiz P, Ellison J, Mao L, et al. (2008) A role for the thromboxane receptor in L-NAME hypertension. *Am J Physiol Renal Physiol* 295: 1096–1102.
43. Kawada N, Dennehy K, Solis G, Modlinger P, Hamel R, et al. (2004) TP receptors regulate renal hemodynamics during angiotensin II slow pressor response. *Am J Physiol Renal Physiol* 287: 753–759.
44. Tian X, Wong WT, Leung FP, Zhang Y, Wang YX, et al. (2011) Oxidative Stress-dependent Cyclooxygenase-2-derived Prostaglandin F<sub>2α</sub> Impairs Endothelial Function in Renovascular Hypertensive Rats. *Antioxid Redox Signal* 16: 363–373.
45. Yu Y, Lucitt MB, Stubbe J, Cheng Y, Friis UG, et al. (2009) Prostaglandin F<sub>2α</sub> elevates blood pressure and promotes atherosclerosis. *Proc Natl Acad Sci USA* 106: 7985–7990.
46. Félixou M, Köhler R, Vanhoutte PM (2010) Endothelium-derived vasoactive factors and hypertension: possible roles in pathogenesis and as treatment targets. *Curr Hypertens Rep* 12: 267–275.
47. Bohr DF, Webb RC (1984) Vascular smooth muscle function and its changes in hypertension. *Am J Med* 77: 3–16.
48. Kishi K, Inoue T (1990) Possible mechanisms of abnormal norepinephrine sensitivity and reactivity of resistance vessels and the development of hypertension in spontaneously hypertensive rats. A hypothesis. *Am J Hypertens* 3: 202–205.
49. dos Santos L, Xavier FE, Vassallo DV, Rossoni LV (2003) Cyclooxygenase pathway is involved in the vascular reactivity and inhibition of the Na<sup>+</sup>, K<sup>+</sup>-ATPase activity in the tail artery from L-NAME-treated rats. *Life Sci* 74: 613–627.

## 7. CONCLUSÕES

Os resultados obtidos neste estudo demonstram que a exposição intrauterina e perinatal à hiperglicemia materna induz hipertensão arterial, associada à disfunção endotelial e hiper-reatividade vascular à noradrenalina em artérias de resistência da prole adulta. Este efeito é influenciado pela idade e é mediado por um aumento, nestas artérias, da expressão protéica da COX-2 e conseqüente liberação de prostanoídes vasoconstritores, como o TxA<sub>2</sub>, PGE<sub>2</sub> e a PGF<sub>2α</sub>.